

# Protein Quality of Amaranth Grains Cultivated in Ethiopia as Affected by Popping and Fermentation

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## Abstract

The effect of popping and fermentation on protein quality of three different varieties of amaranth grains cultivated in Ethiopia was evaluated. Total lysine content of the grains was higher than that of commonly available cereals but close to that of legumes. Methionine and cysteine contents in the grains were also higher than that found in cereal and legume proteins. Percentage of total indispensable amino acids, excluding tryptophan, was 43% - 49%, which was higher than WHO reference pattern (31%). Popping resulted in 36% and 37% reduction in total lysine and cysteine contents, respectively, whereas fermentation reduced cysteine, lysine and methionine contents by 16%, 20% and 20%, respectively. From the free amino acids, histidine was the major indispensable amino acid but threonine was not detected. During popping, all free amino acids, except threonine, were reduced. On the other hand, fermentation significantly increased ( $p < 0.01$ ) most amino acids except arginine, which was significantly decreased ( $p < 0.01$ ), and tyrosine and glutamic acid, for which no change was observed. Popping decreased *in vitro* protein digestibility (IVPD) by 8.3% - 17.1% while fermentation increased IVPD by 4.8% - 7.5%. Substitution of amaranth for wheat and/or maize during complementary food formulation could contribute much to the daily requirements of indispensable amino acids of young children.

## Keywords

Amaranth Grain, Indispensable Amino Acids, Popping, Fermentation, Protein Digestibility,

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## Complementary Food

### 1. Introduction

The genus *Amaranthus* belongs to the family Amaranthaceae. It has a huge biodiversity and several of them are cultivated as leafy vegetables, grains, and ornamental plants while others are weeds [1]. Among all the species, *Amaranthus caudatus*, *Amaranthus hypochondriacus* and *Amaranthus cruentus* are mainly cultivated for their seeds [2] [3].

Amaranth was neglected from the food table for many decades after the arrival of the Spanish conquistadors in Latin America as it was used in ceremonial dishes associated with human sacrifice by the Aztecs [1]. It is known for its high tolerance to arid conditions and poor soils, its resistance to drought, heat, and pests, and its ability to adapt to environments where conventional cereal crops do not grow well. Amaranth can thus contribute to food security, especially in resource poor setting [1] [4]. Despite the very wide distribution of amaranth in Ethiopia, it is cultivated as intercropped with sorghum and maize mainly by Me'enit people who live in the Southern Nations, Nationalities and Peoples Region (SNNPR). The most frequent mode of consumption of amaranth grains by these people is after popping and milling, and by mixing the flour with other cereal flours such as teff, sorghum, barley and wheat to prepare bread, injera and porridge.

Concerning the nutritional quality, protein content of amaranth grains is higher than that of most common cereals [5] [6]. In addition, amaranth grains contain useful amino acid profile [6]-[8]. Despite the high protein content and interesting amino acid profile of amaranth grains, the overall protein quality is dependent on the digestibility. In this regard, popping, the most commonly used processing method for amaranth, and fermentation, which is widely used in Africa to prepare cereal based foods like injera, can influence protein digestibility [9]-[12]. This is due to the fact that these techniques are associated with the decrease in the content of exogenous factors such as tannin, phytate and trypsin inhibitors which are responsible for reducing protein digestibility.

Other processing techniques such as boiling, microwave cooking, and autoclaving also affect protein digestibility. According to El-Adawy *et al.* (2002) [10], boiling, microwave cooking and autoclaving improved the digestibility of chickpea protein. On the other hand, Nunes *et al.* (2004) [13] reported that cooking decreased the digestibility of sorghum and maize protein. Similar controversies surround the effect of popping on the digestibility of amaranth protein. According to a study conducted by Gamel *et al.* (2004) [6], popping improved the IVPD whereas Pedersen *et al.* (1987) [5] and Písaříková *et al.* (2005) [8] reported a decrease in IVPD during popping of amaranth. On the other hand, fermentation has been found to improve *in vitro* protein digestibility of cereals due to the degradation of protein binding molecules [9] [11] [12].

In developing countries like Ethiopia, complementary foods for young children are mainly prepared from mixes of cereals and legumes. It is known that among essential amino acids, legumes are rich sources of lysine but lack methionine and cysteine, while cereals are rich in these two amino acids but are poor sources of lysine. Mixing legumes and cereals during complementary food formulation is a common strategy to meet essential amino acid requirements. However, a too high proportion of legumes sometimes compromise the acceptability of the product and, in addition, legumes contain high levels of antinutritional factors. It is thus advisable to reduce the proportion of legumes in complementary food formulation.

The aim of the present study was, therefore, to evaluate the protein quality of three different varieties of amaranth grains cultivated in Ethiopia in the raw state and after popping or fermentation by analyzing the contents of free and total amino acids and *in vitro* protein digestibility of amaranth grain porridge to assess its potential application as an ingredient for complementary food formulation.

## 2. Materials and Methods

### 2.1. Materials

Three different varieties of amaranth grains, white, red and brown in color, were purchased from six farmers in Chat Kebelle, Bench Majji Zone, in the Southern Nations, Nationalities and Peoples region, Ethiopia. Composite samples of each variety prepared from the grains of the six origins were sorted, cleaned and washed to remove immature seeds, sand and soil. The washed seeds were sun dried, milled and sieved in a 0.425 mm sieve. The flour was stored at 4°C until further analysis.

## 2.2. Solvents, Chemicals and Reagents

Citrate buffer pH 2.2 and ninhydrin reagent were purchased from Biochrom (France). Sodium hydroxide (BioX-tra,  $\geq 98\%$  acidimetric), pellets (anhydrous), L-Norleucine (N8513 suitable for amino acid analysis), aminobutyric acid (A2129;  $\geq 99\%$ ), methanesulfonic acid (M4141; 4 M with 0.2% (w/v) tryptamine) were purchased from Sigma-Aldrich (Saint-Louis, Missouri, USA). Amino acid standards (AA-S18; analytical standard), hydrochloric acid 0.1 N were purchased from Fluka (Fluka Chemicals, Buchs, Switzerland). All reagents and chemicals used were of analytical grade.

## 2.3. Processing Methods

### 2.3.1. Popping

Sun dried seeds were placed in a hot clay pan for 10 - 15 seconds until they popped. The popped grains were milled to pass through a 0.425 mm sieve and stored in polyethylene bags at 4°C.

### 2.3.2. Fermentation

Natural fermentation was carried out according to the method described in [14] with modifications. Briefly, 250 g of flour prepared from raw amaranth grains was mixed with 500 ml distilled water and then left to ferment spontaneously for 48 h at room temperature. The sample was mixed, transferred into aluminum cups and dried in a hot oven (Heraeus UT 5042, Germany) at 50°C for 20 h. The dried sample was then ground to pass through a 0.425 mm sieve and stored in polyethylene bags at 4°C.

### 2.3.3. Preparation of Gruels

Amaranth gruels were prepared using the method described in [15]. Briefly, the flour was mixed with cold deionized water into slurry and cooked with continuous stirring over a hot plate for 5 min after the mixture starts to boil. The gruel was allowed to cool and sample was taken for protein digestibility experiment.

## 2.4. Chemical Analysis

### 2.4.1. Protein and Dry Matter Measurement

The nitrogen content of all Samples was determined using Kjeldhal method and a conversion factor of 5.85 was used [16]. The dry matter content was determined by oven drying at 105°C to a constant weight.

### 2.4.2. Free and Total Amino Acid Analysis

- 1) Extraction procedure
  - a) Free amino acids

Free amino acids were analyzed following the method used by Moore *et al.* (1958) [17] with modifications. Briefly, 150 mg of sample was weighed and placed in a sealable test tube. To this, 50  $\mu$ l of internal standard Norleucine (25  $\mu$ M) and 4.95 ml of citrate buffer (pH 2.2) were added. The solution was mixed for 1 h on a rotational shaker. All extractions were performed in triplicate.

- b) Total amino acids

Samples (10 - 20 mg) of dried flour were weighed in a Schlenk tube and 50  $\mu$ l of 25  $\mu$ M Norleucine and 450  $\mu$ L of 4 M methanesulfonic acid were added. The tube was flushed with nitrogen, closed and heated at 150°C for 2 h. After cooling, 450  $\mu$ L of 4 M NaOH was added to the hydrolysate, which was diluted to 5 ml with a loading buffer (citrate buffer at pH 2.2). All extractions were performed in triplicate.

- 2) Amino acid analysis

Sample extracts for free and total amino acid analysis were filtered using a 0.45  $\mu$ m membrane filter and injected into the amino acid analyzer (Biochrom 30+, Biochrom, France), using a lithium cation exchange resin column and ninhydrin as detection compound. All total and free amino acid contents were determined except total tryptophan, which is the most fragile amino acid and is destroyed by the extraction procedure. Amino acid standards were also run in a similar way as the samples.

### 2.4.3. Determination of *in Vitro* Protein Digestibility

*In vitro* protein digestibility was determined according to the method of Akeson and Stahmanna (1964) [18] with modifications. To 10 g of amaranth porridge (with approximately 10% assumed DM content), 15 ml of

0.16 M HCl containing 1.5 mg pepsin (Sigma, P-7000, 14,900 u/mL) was added and incubated at 37°C for 2 h in a shaking water bath. The resulting suspension was neutralized with 7.5 ml of 0.32 M NaOH and treated with 4 mg of pancreatin (Sigma, P-7545, 8\*USP specifications) in 7.5 ml of 0.32 M phosphate buffer (pH 8.0). The mixture was incubated for an additional 2 h at 37°C. Enzyme blank was prepared by incubation under the same conditions except that the sample was omitted. After incubation, the sample was treated with 10 ml of 10% trichloroacetic acid (Sigma) to remove undigested protein and larger peptides and centrifuged at 5000 g for 20 min at room temperature. The solution was filtered using Whatman No. 1 filter paper, and the protein in the supernatant was then determined using Kjeldahl method.

#### 2.4.4. Amino Acid Score and Protein Digestibility Corrected Amino Acid Score

The amino acid score (AAS) is the ratio of the amino acid content in the protein of a food/diet to the content of the same amino acid in the requirement pattern. The score determines the effectiveness with which absorbed dietary nitrogen can meet the indispensable amino acid requirements at the safe level of protein intake. This is thus a measure of the actual amounts of individual amino acids in a food with respect to the need for this amino acid. And the quality of the protein will finally depend on the indispensable amino acid for which the AAS is the lowest. However, AAS does not account for the digestibility of the protein. Therefore, another scale called the protein digestibility corrected amino acid score (PDCAAS) was adopted by WHO/FAO (2007) [19] to better explain digestibility in relation to the needs of humans and the scoring of foods (Equation (1)):

$$\text{PDCAAS} = \text{amino acid score} \times \text{digestibility} \quad (1)$$

### 2.5. Calculation of the Percentage Contribution of Amaranth to Estimated Amino Acid Requirements

From the amino acid data, intakes of indispensable amino acids were calculated for comparison with the WHO estimated needs [19], assuming breast-fed infants aged 6 - 23 months consume the recommended daily ration size of 50 g/d dry weight complementary food [20]. The amount of cereal used in complementary food formulation was assumed to be 64% [21] and, assuming that this amount of cereal is replaced by amaranth, its percent contribution to the daily indispensable amino acid requirements was calculated.

### 2.6. Data Analysis

All measurements were done in triplicate and statistical analyses were performed using the software Statgraphics plus 5.1 (Statpoint, Warrenton, USA). Two-way ANOVA was applied to the experimental data and means were separated using Fischer's least significant difference tests with a probability  $p < 0.05$ .

## 3. Results and Discussion

### 3.1. Protein, Total and Free Amino Acid Content in Amaranth Grains

**Table 1** indicates the crude protein, and total and free amino acid contents in raw amaranth grains. The protein contents of raw white, red and brown amaranth were 13.9, 15.2 and 15.5 g/100g DM, respectively. These results are in agreement with the results reported by Pedersen *et al.*, (1987) [5]. Kaur *et al.* (2010) [3] also reported that 14.8% - 16.8% protein has been obtained from 11 lines of *Amaranthus caudatus* species grown in India.

The amount of almost all essential amino acids in the raw samples was above the new FAO/WHO standard pattern for all age groups [19] implying that there is no limiting amino acid in amaranth grains considered in this study. Tryptophan, one of the indispensable amino acids, was not analyzed in this study but Gamel *et al.* (2004) [6] reported that their amaranth samples also contained appreciable amounts of tryptophan.

The lysine content of raw amaranth ranged from 65 to 74 mg·g<sup>-1</sup> protein which is close to the lysine contents in legumes (70 - 75 mg·g<sup>-1</sup> protein) including chickpeas, cowpeas and lentils but twice higher than that found in many other cereals, such as barley, wheat, rice, rye and maize: 22 - 37 mg·g<sup>-1</sup> protein. Methionine and cysteine contents were ranged from 24 to 29 and 43 to 51 mg·g<sup>-1</sup> protein, respectively, which were higher than that found in the legumes and cereals mentioned above [22] [23]. The concentrations of essential amino acid in amaranth grains cultivated in Ethiopia (**Table 1**) were in good agreement with those reported by Chavez-Jauregui *et al.* (2000) [24] and slightly higher than those reported by Gamel *et al.* (2004) [6] in *Amaranthus caudatus*.

The percentage of indispensable amino acids, excluding tryptophan, tryptophan, was highest in raw white

**Table 1.** Total and free amino acid profile ( $\text{mg}\cdot\text{g}^{-1}$  protein) of three varieties of raw amaranth grains<sup>†</sup>.

Protein (g/100g DM)	White amaranth		Red amaranth		Brown amaranth		Standard pattern <sup>‡</sup>
	13.88 ± 0.16		15.15 ± 0.08		15.53 ± 0.24		
Amino acid	Total	Free	Total	Free	Total	Free	
Cysteine <sup>*</sup>	50.7 ± 8.3	0.9 ± 0.0	42.7 ± 3.0	1.3 ± 0.2	45.5 ± 4.3	0.7 ± 0.1	
Histidine <sup>*</sup>	37.2 ± 3.1	1.4 ± 0.1	33.9 ± 2.5	1.5 ± 0.0	34.8 ± 2.0	1.2 ± 0.1	18.0
Isoleucine <sup>*</sup>	39.4 ± 3.0	0.4 ± 0.1	37.2 ± 2.6	0.4 ± 0.1	34.9 ± 2.4	0.3 ± 0.1	31.0
Leucine <sup>*</sup>	67.1 ± 2.9	0.4 ± 0.0	60.8 ± 2.6	0.4 ± 0.0	59.5 ± 0.2	0.2 ± 0.0	63.0
Lysine <sup>*</sup>	73.9 ± 2.3	1.2 ± 0.0	66.7 ± 7.2	1.1 ± 0.0	65.3 ± 1.6	0.9 ± 0.0	52.0
Methionine <sup>*</sup>	28.8 ± 0.5	0.4 ± 0.1	23.8 ± 3.4	0.5 ± 0.2	25.8 ± 0.7	0.3 ± 0.1	
Phenylalanine <sup>*</sup>	46.5 ± 3.6	0.2 ± 0.0	42.6 ± 1.4	0.2 ± 0.0	40.4 ± 3.5	0.1 ± 0.0	
Threonine <sup>*</sup>	40.3 ± 1.9	0.0 ± 0.0	35.0 ± 3.4	0.0 ± 0.0	36.7 ± 1.1	0.0 ± 0.0	27.0
Tryptophan <sup>*</sup>	ND	1.0 ± 0.0	ND	1.4 ± 0.1	ND	0.7 ± 0.0	7.4
Valine <sup>*</sup>	48.3 ± 1.2	0.7 ± 0.1	42.0 ± 4.6	0.7 ± 0.1	41.3 ± 2.7	0.5 ± 0.1	42.0
Alanine	44.0 ± 2.0	1.1 ± 0.0	38.2 ± 5.8	0.5 ± 0.1	39.0 ± 1.5	1.0 ± 0.0	
Arginine	109.8 ± 8.0	1.6 ± 0.0	102.2 ± 10.0	3.6 ± 0.3	100.6 ± 4.0	1.0 ± 0.0	
Aspartic acid	94.4 ± 5.2	1.1 ± 0.0	86.0 ± 9.3	2.2 ± 0.1	89.4 ± 10.0	1.2 ± 0.0	
Glutamic acid	203.5 ± 14.0	2.9 ± 0.0	183.5 ± 15.0	4.6 ± 0.3	181.8 ± 3.0	1.8 ± 0.0	
Glycine	100.1 ± 4.0	1.3 ± 0.0	89.1 ± 6.4	0.4 ± 0.1	100.0 ± 4.0	1.1 ± 0.0	
Proline	135.7 ± 25.6	2.4 ± 0.0	125.0 ± 30.0	3.7 ± 0.2	118.4 ± 17.4	1.5 ± 0.0	
Serine	65.5 ± 4.0	1.3 ± 0.2	58.5 ± 4.8	2.1 ± 0.1	68.9 ± 1.6	0.6 ± 0.0	
Tyrosine	52.9 ± 6.0	0.5 ± 0.0	45.8 ± 2.7	0.7 ± 0.0	49.7 ± 3.4	0.4 ± 0.0	
AAA <sup>§</sup>	99.4 ± 6.3		88.4 ± 2.6		90.1 ± 3.4		46.0
SAA <sup>¶</sup>	79.5 ± 7.9		66.5 ± 5.9		71.3 ± 4.9		26.0
NH <sub>3</sub>	78.3 ± 6.1	3.2 ± 0.0	70.0 ± 14.3	2.0 ± 0.1	76.6 ± 12.2	1.4 ± 0.1	

<sup>†</sup>Values are means of results obtained in triplicate ± SD expressed in  $\text{mg}\cdot\text{g}^{-1}$  protein and means followed by different letters in the same row are significantly different at  $p < 0.05$ . <sup>§</sup>AAA: aromatic amino acids (phenylalanine and tyrosine). <sup>¶</sup>SAA: sulfur amino acids (cysteine and methionine). ND: not determined. <sup>\*</sup>Indispensable amino acids [19]. <sup>‡</sup>Standard FAO/WHO reference pattern for indispensable amino acids for children aged 1 - 2 years [19].

amaranth (49%) followed by 43% in both raw red and brown amaranth. These values are higher than the reference protein pattern for 1 - 2 years old children (31%). The amaranth amino acid profile thus provides a good balance of total indispensable amino acids, and some of the limiting amino acids, especially lysine in cereals and methionine and cysteine in legumes, could be complemented by amaranth.

The free amino acid content of the three varieties of raw amaranth is listed in **Table 1**. The main essential free amino acid was histidine followed by lysine and tryptophan with 1.2 - 1.5, 0.9 - 1.2 and 0.7 - 1.4  $\text{mg}\cdot\text{g}^{-1}$  protein, respectively. A study by Nimbalkar *et al.* (2012) [25] on *Amaranthus hypocondriacus* grain reported that the top three essential free amino acids were threonine, phenylalanine, and methionine and that the amount of threonine was twice that of the other free amino acids. However, in the present study, no free threonine was detected in any of the three varieties of amaranth grains, and glutamic acid and proline were the main non-essential free amino acids with values in the range of 1.8 to 4.6 and 1.5 to 3.7  $\text{mg}\cdot\text{g}^{-1}$  protein, respectively.

### 3.2. Effect of Popping and Fermentation on Total Amino Acid Content

Popping decreased the amount of six amino acids including three indispensable amino acids: lysine, methionine,

and cysteine. The decrease was highest for lysine and cysteine and reached 36% and 37% reduction, respectively (Table 2). In a study conducted in two different species of amaranth (*Amaranthus caudatus* and *Amaranthus cruentus*) by Gamel et al. (2004) [6], the highest loss of amino acid was recorded for tyrosine followed by phenylalanine and methionine with 35%, 24% and 18% loss for *Amaranthus caudatus* and 32%, 20% and 19% for *Amaranthus cruentus*, respectively. Another study by Tovar et al. (1989) [26] reported highest loss for lysine (56%) followed by arginine (29%). Pisarikova et al. (2005) [8] also reported that the decrease in the concentration of indispensable amino acids due to heat treatment was highest for histidine followed by lysine and leucine. Despite the improvement in sensory attributes during popping, the associated loss of amino acids decreased the overall quality of amaranth proteins.

In the present study, fermentation also led to a significant decrease in the contents of the same six amino acids as that of popping (Table 2). However, after fermentation, the decrease in essential amino acids, lysine and cysteine with percent reduction of 20 and 16%, respectively, were significantly lower than after popping ( $p < 0.05$ ) and methionine was subject to a higher (20%) percent reduction although not statistically significant ( $p < 0.05$ ). The reduction could be ascribed to metabolism of amino acids by microorganisms into ammonia and volatile compounds that are responsible for the flavor of the fermented product [12].

### 3.3. Effect of Popping and Fermentation on Free Amino Acid Content

Popping and fermentation led to highly significant changes in free amino acid contents ( $p < 0.01$ ) (Table 3).

**Table 2.** Effect of popping and fermentation on total amino acid content ( $\text{mg}\cdot\text{g}^{-1}$  protein) of amaranth grains<sup>†</sup>.

Amino acid	p-value	Processing				
		Raw	Popped		Fermented	
		Mean $\pm$ SD	Mean $\pm$ SD	% variation	Mean $\pm$ SD	% variation
Cysteine	0.0023*	46.3 $\pm$ 6.0 a	29.4 $\pm$ 8.2 c	-37	38.9 $\pm$ 15.0 b	-16
Histidine	0.8097	35.3 $\pm$ 2.7	35.0 $\pm$ 1.3	-	34.6 $\pm$ 2.8	-
Isoleucine	0.1014	37.2 $\pm$ 3.0	33.7 $\pm$ 4.7	-	35.4 $\pm$ 3.2	-
Leucine	0.4034	62.5 $\pm$ 4.0	60.3 $\pm$ 4.3	-	60.8 $\pm$ 5.0	-
Lysine	0.0000*	68.6 $\pm$ 5.6 a	44.0 $\pm$ 10.1 c	-36	55.0 $\pm$ 16.8 b	-20
Methionine	0.0062*	26.1 $\pm$ 2.8 a	23.0 $\pm$ 4.1 b	-12	20.8 $\pm$ 5.1 b	-20
Phenylalanine	0.3940	43.2 $\pm$ 2.8	41.4 $\pm$ 3.3	-	41.4 $\pm$ 4.5	-
Threonine	0.0836	37.3 $\pm$ 3.1	35.1 $\pm$ 2.0	-	35.4 $\pm$ 3.3	-
Tyrosine	0.1473	49.5 $\pm$ 4.8	48.5 $\pm$ 2.7	-	45.7 $\pm$ 4.8	-
Valine	0.0832	43.9 $\pm$ 4.3	40.2 $\pm$ 5.5	-	42.2 $\pm$ 3.8	-
Alanine	0.3488	40.4 $\pm$ 4.2	38.7 $\pm$ 2.6	-	39.8 $\pm$ 2.6	-
Arginine	0.0003*	104.2 $\pm$ 8.0 a	88.6 $\pm$ 11.9 b	-15	83.2 $\pm$ 15.1 b	-20
Aspartic acid	0.0191*	89.9 $\pm$ 6.5 a	84.8 $\pm$ 2.7 b	-6	83.1 $\pm$ 5.5 b	-8
Glutamic acid	0.0711	189.6 $\pm$ 15.0	183.6 $\pm$ 6.8	-	176.3 $\pm$ 13.3	-
Glycine	0.6262	96.4 $\pm$ 7.1	93.8 $\pm$ 3.7	-	95.4 $\pm$ 7.3 a	-
Proline	0.0584	126.4 $\pm$ 23.0	112.9 $\pm$ 15.5	-	106.5 $\pm$ 8.4	-
Serine	0.0049*	64.3 $\pm$ 5.6 a	59.1 $\pm$ 3.2 b	-8	57.2 $\pm$ 3.7 b	-11
Ammonia	0.1330	75.0 $\pm$ 10.6	71.9 $\pm$ 9.5	-	80.7 $\pm$ 10.3	-

<sup>†</sup>Values are means of results obtained in triplicate  $\pm$  SD for the three amaranth varieties. \*The effect of processing is significant ( $p < 0.05$ ) and means followed by different letters in the same row are significantly different at  $p < 0.05$ .

**Table 3.** Effect of popping and fermentation on free amino acid content ( $\text{mg}\cdot\text{g}^{-1}$  protein) of amaranth grains<sup>†</sup>.

Amino acid	p-value	Processing				
		Raw	Popped		Fermented	
		Mean $\pm$ SD	Mean $\pm$ SD	% variation	Mean $\pm$ SD	% variation
Arginine	p < 0.01	2.0 $\pm$ 1.2 a	0.8 $\pm$ 0.6 b	-60	0.3 $\pm$ 0.1 c	-85
Cysteine		1.0 $\pm$ 0.3 b	0.5 $\pm$ 0.1 c	-50	3.7 $\pm$ 0.7 a	270
Histidine		1.4 $\pm$ 0.2 b	0.4 $\pm$ 0.1 c	-71	4.6 $\pm$ 2.6 a	229
Isoleucine		0.4 $\pm$ 0.1 b	0.1 $\pm$ 0.1 c	-75	2.8 $\pm$ 0.5 a	600
Leucine		0.3 $\pm$ 0.1 b	0.1 $\pm$ 0.1 c	-67	6.5 $\pm$ 1.4 a	2067
Lysine		1.1 $\pm$ 0.2 b	0.7 $\pm$ 0.2 c	-36	2.0 $\pm$ 0.6 a	82
Methionine		0.4 $\pm$ 0.2 b	0.1 $\pm$ 0.1 c	-75	2.6 $\pm$ 0.5 a	550
Phenylalanine		0.2 $\pm$ 0.1 b	0.0 $\pm$ 0.0 c	-100	3.5 $\pm$ 0.8 a	1650
Threonine		0.0 $\pm$ 0.0 b	0.0 $\pm$ 0.0 b	-	0.2 $\pm$ 0.0 a	100
Tryptophan		1.0 $\pm$ 0.3 b	0.1 $\pm$ 0.1 c	-90	2.1 $\pm$ 0.7a	110
Tyrosine		0.5 $\pm$ 0.1 a	0.0 $\pm$ 0.1 b	-100	0.5 $\pm$ 0.4 a	-
Valine		0.6 $\pm$ 0.1 b	0.3 $\pm$ 0.2 c	-50	4.0 $\pm$ 0.7 a	344
Alanine		0.9 $\pm$ 0.3 b	0.5 $\pm$ 0.3 c	-44	5.2 $\pm$ 1.2 a	478
Aspartic acid		1.5 $\pm$ 0.5 b	0.5 $\pm$ 0.2 c	-67	2.3 $\pm$ 0.5 a	53
Glutamic acid		3.1 $\pm$ 1.2 a	0.6 $\pm$ 0.4 b	-81	3.2 $\pm$ 1.7 a	-
Glycine		0.9 $\pm$ 0.4 b	0.8 $\pm$ 0.5 c	-11	3.6 $\pm$ 1.2 a	300
Ornithine		0.2 $\pm$ 0.0 b	0.1 $\pm$ 0.0 c	-50	2.1 $\pm$ 1.0 a	950
Proline		2.5 $\pm$ 1.0 a	0.4 $\pm$ 0.3 c	-84	2.4 $\pm$ 1.3 b	4
Serine		1.3 $\pm$ 0.7 b	0.3 $\pm$ 0.2 c	-77	1.7 $\pm$ 0.3 a	31
Ammonia		2.2 $\pm$ 0.8 b	3.2 $\pm$ 0.6 c	-46	14.1 $\pm$ 2.3 a	541

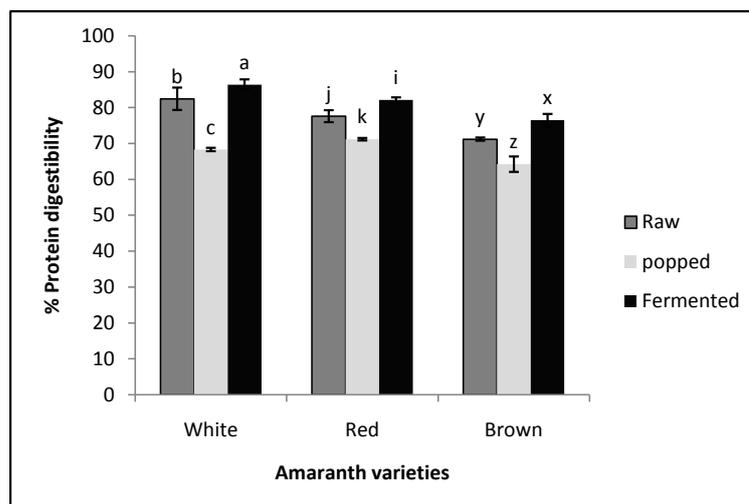
<sup>†</sup>Values are means of results obtained in triplicate  $\pm$  SD expressed for the three amaranth varieties. Means followed by different letters in the same row are significantly different at  $p < 0.01$ .

Popping led to a significant decrease in almost all free amino acid contents (**Table 3**). Free aromatic amino acids were strongly affected, phenylalanine and tyrosine completely vanished and 90% of tryptophan vanished during popping. Non-enzymatic browning reaction is the most probable explanation for the decrease in the level of free amino acids during heat treatment [27].

Conversely, fermentation increased the amount of almost all free amino acids, except tyrosine, glutamic acid and proline, which remained unchanged, and arginine, which strongly decreased (**Table 3**). Lysine and phenylalanine increased to a greater extent than the other free amino acids. Increased microbial enzyme activity coupled with protein hydrolysis is the likely explanation for the increase in the amount of free amino acids during fermentation [12] [28]. Hamad and Fields (1979) [29] also showed that there was a significant increase in free lysine content during fermentation in other type of cereals.

### 3.4. Effect of Popping and Fermentation on *in Vitro* Protein Digestibility (IVPD) and Protein Digestibility Corrected Amino Acid Score (PDCAAS)

*In vitro* protein digestibility (IVPD) is shown in **Figure 1**. When the digestibility of porridge made from raw amaranth samples was compared, it was highest for white (82.4%) followed by red (77.6%) and brown (71.2%) varieties. The lower IVPD in the colored varieties might be related to the presence of higher amount of dietary



**Figure 1.** Effect of processing on *in vitro* protein digestibility (%) in amaranth. Means with different letters are significantly different at  $p < 0.05$  for the same variety.

fiber and polyphenols (data not shown), which could bind the protein and reduce its susceptibility to enzymatic attack [5].

Both popping and fermentation significantly affected, ( $p < 0.05$ ), the IVPD (**Figure 1**). Popping decreased protein digestibility by 17.14%, 8.3% and 9.8% for white, red and brown colored amaranth, respectively. This is because amino acids undergo several chemical reactions during severe heat treatment, including Maillard reaction between amino acids and reducing sugars, which reduces the availability of amino acids [27]. Therefore, during dry heat processing (popping) where the temperature often exceeds  $150^{\circ}\text{C}$ , this reaction is likely to contribute to the decrease in protein digestibility. Unlike popping, fermentation improved IVPD by 4.8%, 5.9% and 7.5% for white, red and brown colored amaranth, respectively. This could be due to degradation of phytic acid (data not shown), which is a potent inhibitor of proteolytic enzymes. Besides, hydrolysis of tannins, which potentially complex proteins, may also increase the accessibility of protein molecules for enzymatic attack and hence increase IVPD [30].

In addition, hydrolysis of proteins by microorganisms during fermentation, evidenced by the increase in free amino acids, could increase IVPD. Studies by El Hag *et al.* (2002) [11], Pranoto *et al.* (2013) [12] and Hassan and El Tinay (1995) [9] on finger millet and sorghum, indeed showed that tannin hydrolysis occurs under the action of microorganisms during fermentation.

For all the indispensable amino acids, the PDCAAS, which takes both the amino acid score and protein digestibility into account, was highest for the white followed by the red and brown variety (**Table 4**). In all the three varieties, the popped samples had the lowest PDCAAS values due to the loss of heat labile amino acids during popping. Although there was a significant improvement ( $p < 0.05$ ) in protein digestibility during fermentation (**Figure 1**), the PDCAAS of almost all essential amino acids slightly increased or remained unchanged after fermentation except lysine in the brown variety, which decreased considerably. In the white and red varieties, leucine in raw samples, leucine and lysine in popped samples and leucine in fermented samples were decreased, while in the brown variety, leucine in raw samples and lysine in popped and fermented samples were decreased.

### 3.5. Percentage Contribution of Amaranth as an Ingredient in Complementary Foods to the Daily Amino Acid Requirement of 6 - 23 Months old Children

**Table 5** shows the percent contribution of amaranth to the daily indispensable amino acid requirements of 6 - 23 months old children. All the three varieties of amaranth contributed more than 50% of the daily indispensable amino acid requirements based on the consumption of 50 g of complementary foods per day of which amaranth accounts for 32 g. Compared to the cereals commonly used in complementary food formulation such as wheat and maize, the contribution of amaranth is appreciably higher for all indispensable amino acids except leucine

**Table 4.** Percent protein digestibility corrected amino acid score (PDCAAS) for three varieties of raw and processed amaranth grains\*.

Amino acid	White amaranth			Red amaranth			Brown amaranth		
	Raw	Popped	Fermented	Raw	Popped	Fermented	Raw	Popped	Fermented
AAA <sup>§</sup>	178	135	174	149	138	159	139	125	133
Histidine	170	128	170	146	143	162	138	126	139
Isoleucine	105	81	107	93	70	94	80	70	79
Leucine	88	68	89	75	66	79	67	61	69
Lysine	117	68	110	100	66	104	89	40	49
SAA <sup>§</sup>	252	157	248	198	134	224	195	119	99
Threonine	123	94	122	101	88	110	97	83	90
Valine	95	74	95	78	60	82	70	60	71

\*The scoring pattern for indispensable amino acids was considered for children aged 1 - 2 years [19]. <sup>§</sup>SAA: sulfur amino acids (cysteine and methionine). <sup>§</sup>AAA: aromatic amino acids (phenylalanine and tyrosine).

**Table 5.** Percentage contribution of raw amaranth to the daily indispensable amino acid requirements for children aged 6 - 23 months, compared to that of more commonly used cereals (maize or wheat)\*.

Amino acid	Amount of amino acid (mg/100g DM)					Requirement <sup>§</sup>	Percentage contribution (%) <sup>§</sup>				
	Wheat <sup>†</sup>	Maize <sup>†</sup>	Amaranth				Wheat	Maize	Amaranth		
			White	Red	Brown				White	Red	Brown
AAA	764	795	1379	1340	1400	458 - 476	51 - 53	53 - 56	92 - 96	90 - 93	94 - 98
His	274	244	516	514	540	171 - 178	49 - 51	44 - 46	92 - 96	92 - 97	98 - 102
Ileu	441	325	547	563	542	279 - 321	44 - 51	32 - 37	54 - 62	56 - 64	54 - 62
Leu	811	1129	932	921	925	566 - 642	40 - 46	56 - 64	46 - 53	46 - 52	46 - 53
Lys	334	244	1026	1011	1014	497 - 535	20 - 22	15 - 16	61 - 66	61 - 65	60 - 65
SAA	418	316	1103	1008	1108	241 - 262	51 - 56	39 - 42	136 - 148	124 - 135	135 - 147
Thr	346	334	559	531	570	264 - 273	40 - 42	39 - 41	65 - 67	62 - 64	67 - 70
Val	523	433	671	636	641	380 - 428	39 - 44	32 - 36	50 - 56	48 - 54	48 - 54

\*Assuming daily consumption of 50 g of complementary food on a dry weight basis formulated with 64% of cereals or amaranth. <sup>†</sup>Shewry *et al.* (2007) [23]. <sup>§</sup>The lower and upper values refer to the requirements for 6 and 23 months old children, respectively [19]. <sup>§</sup>The lower and upper values refer to the contribution of unprocessed cereals to the daily amino acid needs for 23 and 6 months old children, respectively.

(Table 5). Moreover, the daily needs of sulfur-containing amino acids were fully met by consuming the suggested amount of amaranth. However, if only the digestible fraction of protein is taken into consideration, the contribution to amino acid requirements would decrease by about 18% - 29%.

#### 4. Conclusion

All three varieties of amaranth investigated in this study can be considered as a potential source of protein. Due to its high essential amino acid contents, amaranth could potentially substitute other more common cereals used in complementary food formulation for young children and thus reduce the proportion of legumes. Although popping improved the sensorial attributes of amaranth porridge, it reduced protein quality through the loss of heat labile amino acids. On the other hand, fermentation was better than popping in maintaining the amino acid profile with its added advantage of improving the protein digestibility although it resulted in a marked decrease in lysine content.

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