

Research Article

OPEN ACCESS

Landmark and outline-based geometric morphometrics analysis of three *Stomoxys* flies (Diptera: Muscidae)

Tanasak Changbunjong^{1,2}, Suchada Sumruayphol³, Thekhawet Weluwanarak², Jiraporn Ruangsittichai³ and Jean-Pierre Dujardin⁴

¹Department of Pre-clinic and Applied Animal Science, Faculty of Veterinary Science, Mahidol University, Nakhon Pathom, Thailand;

²The Monitoring and Surveillance Center for Zoonotic Diseases in Wildlife and Exotic Animals, Faculty of Veterinary Science, Mahidol University, Nakhon Pathom, Thailand;

³Department of Medical Entomology, Faculty of Tropical Medicine, Mahidol University, Bangkok, Thailand;

⁴Institut de Recherches pour le Développement, Campus International de Baillarguet, Montpellier, France

Abstract: Adult flies of the genus *Stomoxys* Geoffroy, 1762 (Diptera: Muscidae), especially *S. pullus* Austen, 1909, *S. uruma* Shinonaga et Kano, 1966 and *S. indicus* Picard, 1908, are morphologically similar and sometimes difficult to distinguish when using external morphological characteristics. These species may act as vectors and/or potential vectors of many pathogens (virus, bacteria and protozoa). Their correct identification is important to target the vectors involved in the transmission of the pathogens and also helps in the fly control program. The aim of the present study was to distinguish three species which are difficult to separate using traditional diagnostic characters for species of *Stomoxys* such as colour patterns and body proportions. Modern morphometrics, both landmark and outline-based, was used to access wing geometry of *S. pullus*, *S. uruma* and *S. indicus*. A total of 198 and 190 wing pictures were analysed for landmark- and outline-based approaches, respectively. Wing shape was able to separate species and sexes of the three *Stomoxys* flies with highly significant difference of Mahalanobis distances. The cross-validated classification scores ranged from 76% to 100% for landmark and 77% to 96% for outline-based morphometrics. The geometry of wing features appears to be a very useful, low-cost tool to distinguish among the vectors *S. pullus*, *S. uruma* and *S. indicus*.

Keywords: muscidae, landmarks, outlines, *Stomoxys pullus*, *Stomoxys uruma*, *Stomoxys indicus*

Flies of the genus *Stomoxys* Geoffroy, 1762 (Diptera: Muscidae) are haematophagous flies of considerable medical and veterinary importance. They are classified into the subfamily Muscinae, tribes Stomoxyini, with 18 species having been described (Zumpt 1973). In Thailand, 6 species of *Stomoxys*, namely *Stomoxys calcitrans* (Linnaeus, 1758), *S. sitiens* Rondani, 1873, *S. bengalensis* Picard, 1908, *S. indicus* Picard, 1908, *S. pullus* Austen, 1909 and *S. uruma* Shinonaga et Kano, 1966, have been recorded from different geographical areas (Tumrasvin and Shinonaga 1978, Masmeatathip et al. 2006, Muenworn et al. 2010; Changbunjong et al. 2012).

The adult flies feed on the blood of humans and animals, making them a nuisance to humans, a major irritant pest of both livestock and wildlife, and they also act as vectors and potential vectors of many pathogens (Zumpt 1973, Baldacchino et al. 2013). They have been implicated as mechanical vectors of viruses (Equine infectious anemia virus, African swine fever virus, West Nile fever virus and Bo-

vine leukosis virus), bacteria (*Bacillus anthracis* and *Anaplasma marginale*) and protozoa (species of *Trypanosoma* Gruby, 1843 and *Besnoitia* Henry, 1913). Moreover, they also act as biological vectors of the helminth *Habronema microstoma* Schneider, 1866 (see Baldacchino et al. 2013).

Species identification of adult *Stomoxys* is based mainly on body colour and pattern, leg colour, frons width proportions, curvature and setation of certain wing veins, occurrence or form of various bristles and hairs on parts of the legs, and also genital structure (Crosskey 1993). Morphological species identification is a gold standard for any taxonomic system, but it might become difficult or unsatisfactory for distinction of cryptic species. Correct identification not only permits critical access to the broad body of literature available on a particular taxon but also permits the implementation of adequate control measures to contend with species of medical and veterinary importance.

Some *Stomoxys* species such as *S. pullus*, *S. uruma* and *S. indicus* have morphological similarity, especially the

Address for correspondence: T. Changbunjong, Department of Pre-clinic and Applied Animal Science, Faculty of Veterinary Science, Mahidol University, Nakhon Pathom 73170, Thailand. Phone: +66-2-4415238; Fax: +66-2-4415238; E-mail: tanasak.cha@mahidol.edu

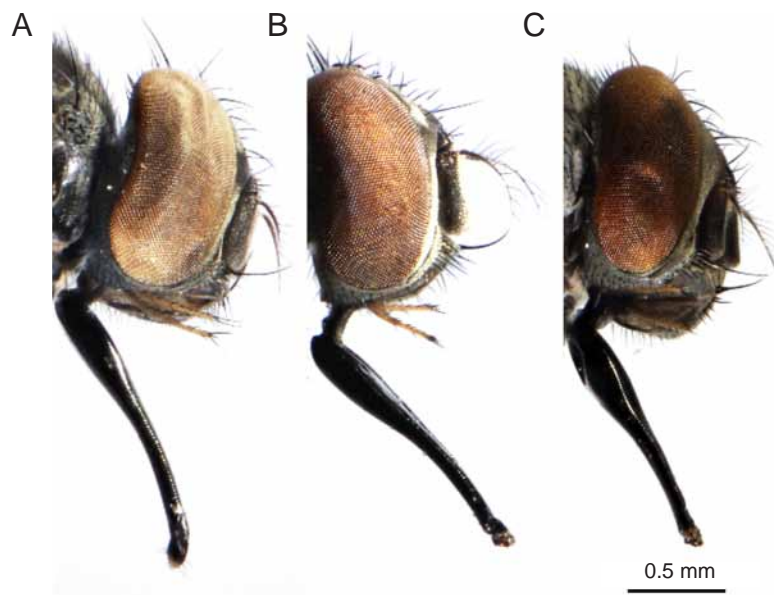


Fig. 1. Morphological characters of palpi used to separate *Stomoxys pullus* Austen, 1909 (A), *S. uruma* Shinonaga et Kano, 1966 (B) and *S. indicus* Picard, 1908 (C).

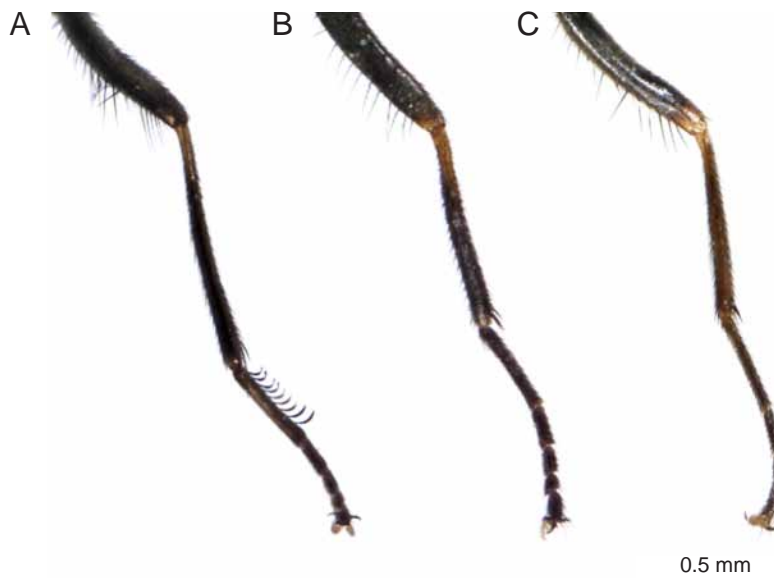


Fig. 2. Morphological characters of tibia and tarsus used to separate *Stomoxys pullus* Austen, 1909 (A), *S. uruma* Shinonaga et Kano, 1966 (B) and *S. indicus* Picard, 1908 (C).

body colour and abdominal pattern (Zumpt 1973), and the body size, as well as the frontal index (the ratio of the smallest width of the frons and the greatest length of the eye), cannot clearly separate them (Changbunjong et al. 2013). As an additional difficulty, these species can also be found in the same areas (Changbunjong et al. 2012).

According to Tumrasvin and Shinonaga (1978), *S. pullus* can be distinguished from *S. uruma* and *S. indicus* by the length of maxillary palpi exceeding the fore margin of the mouth and the different colour at the basal part of the third antennal segment. *Stomoxys indicus* can be distinguished from *S. uruma* by the yellowish colour of tibiae and tarsi (Figs. 1, 2). These morphological traits may be used as the primary method for screening specimens in the field, or for identifying uncomplicated specimens. Howev-

er, the morphological-based identification of these flies is often impeded by polymorphism, overlapping morphological characteristics and damage caused to specimens during collection.

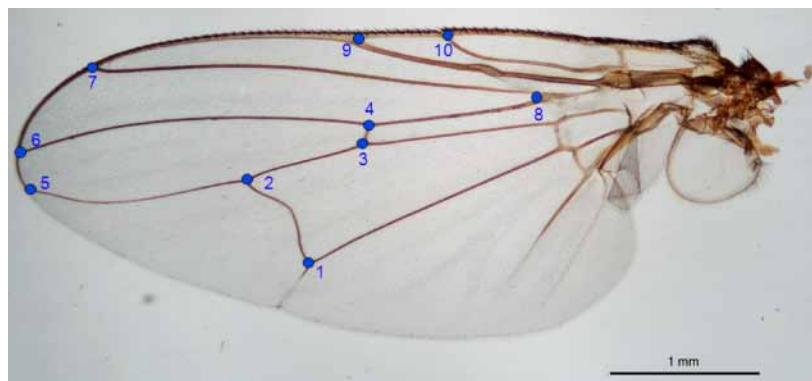
Molecular-based identification can resolve various problems encountered during morphology-based identification, especially of morphologically close species (Hebert 2003a,b), but its use is expensive and requires specialised training (Müller et al. 2013). Geometric morphometrics is increasingly applied to medically and economically important insects to distinguish morphologically similar species, especially cryptic taxa, and to detect intraspecific variation (Dujardin 2008, Ruangsittichai et al. 2011, Lorenz et al. 2012, Dujardin and Kitthawee 2013, Morales Vargas et al. 2013, Demari-Silva et al. 2014, Jaramillo-O et al. 2015,

Table 1. Number of flies of species *Stomoxys* Geoffroy, 1762 used for geometric morphometrics analysis.

Species (sex)	Number	
	Landmark-based method	Outline-based method
<i>S. pullus</i> Austen, 1909 (male)	35	34
<i>S. pullus</i> (female)	35	33
<i>S. uruma</i> Shinonaga et Kano, 1966 (male)	34	32
<i>S. uruma</i> (female)	34	31
<i>S. indicus</i> Picard, 1908 (male)	30	30
<i>S. indicus</i> (female)	30	30
Total	198	190

Table 2. Description of landmarks on wings of species of *Stomoxys* Geoffroy, 1762 (see Fig. 3).

Landmark	Description of the landmark
1	medial vein 3 and cubital vein 1
2	medial cross vein
3	midpoint branch of medial vein
4	radio-medial cross vein
5	distal end of medial vein 1 and 2
6	distal end of the radial vein 4 and 5
7	distal end of the radial vein 2 and 3
8	origin of radial vein 2 and 3
9	intersection of costa and radial vein 1
10	intersection of costa and subcosta

**Fig. 3.** Ten landmarks digitised on wings of species of *Stomoxys* Geoffroy, 1762 flies for landmark-based geometric morphometrics analysis (see Table 2 for description).

Sumruayphol et al. 2016). Wing veins provide many well-defined landmarks suitable for the landmark-based approach (Villegas et al. 2002). In addition to landmark-based morphometrics, outline-based morphometrics has been shown to be a reliable method for characterising various wingless insects and some arthropods (other than insects) with poorly defined landmarks (Dujardin et al. 2014).

A recent study has shown the efficacy of outline-based morphometrics for discriminating between closely related species, or between conspecific populations, of various arthropods including kissing bugs, tsetse flies, mosquito and soft ticks (Dujardin et al. 2014). The combined use of landmarks and outlines could represent a better method for discrimination between species (Francoy et al. 2012). Although geometric morphometrics does not reach the level of molecular accuracy, the present findings show that it can be highly and quickly informative at low cost. In the present study, we used landmark and outline-based geometric morphometrics to identify vector species of the genus *Stomoxys* focusing on three closely related species in Thailand.

MATERIALS AND METHODS

Specimen collection

Flies were collected between July 2014 and August 2015 in two localities in western and northeastern Thailand: *S. pullus* and *S. uruma* from Nakhon Ratchasima Province (14°24'55"N;

101°22'33"E) and *S. indicus* from Kanchanaburi Province (14°25'54"N; 98°48'35"E) using Vavoua traps (Laveissière and Grebaut 1990). The traps were placed at the collection sites from 6:00 AM to 6:00 PM over a two-day-period (Changbunjong et al. 2012). Species were identified with a stereomicroscope based on the taxonomic key of Zumpt (1973) and Tumrasvin and Shinonaga (1978).

Sample preparation and data collection

The left wings of males and females belonging to *S. pullus*, *S. uruma* and *S. indicus* were dissected from the body and mounted by Hoyer's medium on microscopic slides. The wings were placed at the center of the visual view to avoid peripheral optical distortion. All slides were photographed using a digital camera connected to a stereomicroscope (Nikon AZ 100, Nikon Corp, Tokyo, Japan) at 10× magnification. A total of 198 and 190 wing pictures of the three species of *Stomoxys* flies were performed for landmark and outline-based methods, respectively. The simple external contour of the wings was used for outlines, but in eight wings the contours were damaged and so were not satisfactory for outline analysis. Otherwise, the same set of wing pictures was used to compare both methods (Table 1).

Geometric morphometrics analysis

Landmark-based method

The coordinates of ten wing landmarks (Table 2 and Fig. 3) were selected and digitised for geometric morphometrics analysis. The wing size was estimated using the isometric estimator of

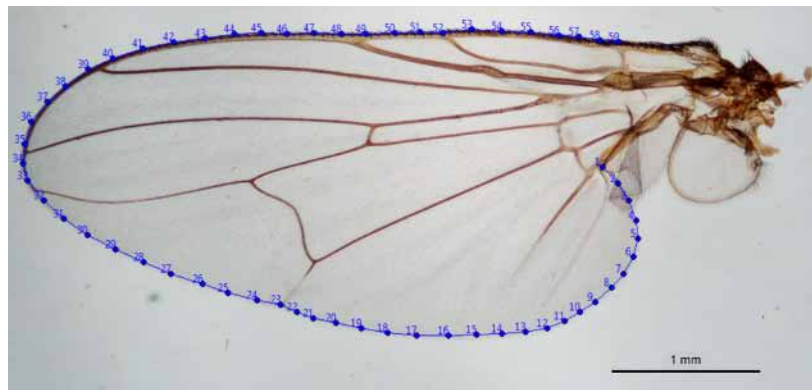


Fig. 4. Contour digitised on *Stomoxys* Geoffroy, 1762 flies wing for outline-based geometric morphometrics analysis. A short, artificial segment is computed by the digitising program to completely close the contour.

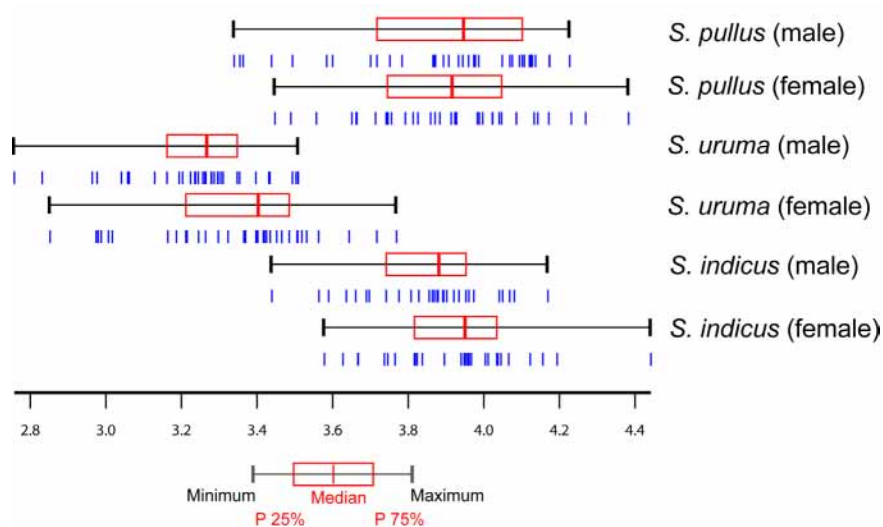


Fig. 5. Centroid size variation of the wings between species and sexes, shown as quartile boxes. Each box shows the group median separating the 25th and 75th quartiles. Vertical bars under the boxes represent the wing (units as mm).

the centroid size (CS) derived from data on coordinates. The centroid size is defined as the square root of the sum of the squared distances between the centre of the configuration of landmarks and each separate landmark (Bookstein 1991). The centroid size difference was compared among species and sexes (15 pairwise comparisons) by non-parametric tests based on 1000 permutations, with Bonferroni correction for test of significance at P -value of 0.05.

Generalised Procrustes Analysis (GPA) (Rohlf 1990) was performed to compute the wing shape variables and the centroid sizes. The Procrustes superimposition provided configurations for visual comparisons of the mean anatomical landmarks between species and sexes. The wing shape variables were initially computed from these configurations ('aligned' configurations) as the partial warps (PW) scores. Then their principal components or relative warps (RW) were used as input for the discriminant analyses (or canonical variate analysis). The discriminant analyses were illustrated by the factor maps. The statistical significance of shape differences among the species and sexes was obtained by non-parametric analyses based on 1000 permutations, with Bonferroni correction for test of significance at P -value of 0.05.

Outline-based method

The outline considered for species comparisons was the external contour of the wing (Fig. 4). The wing size may be estimated as the outline perimeter or by the square root of the first harmonic ellipse area. It was compared among species and sexes using non-parametric analyses in the same way as for CS.

Elliptic Fourier Analysis (EFA) (Kuhl and Giardina 1982) was performed to produce the wing shape variables. It provided configurations for visual comparisons of the outlines between species and sexes. The wing shape variables were computed as Normalised Elliptic Fourier coefficients. To deal with possible problems of multidimensionality, a reduced set of their principal components was used as input for the discriminant analyses (for methodological details, see Dujardin et al. 2014). Statistical comparisons of wing shape among the species and sexes were the same as those used for the landmark-based method.

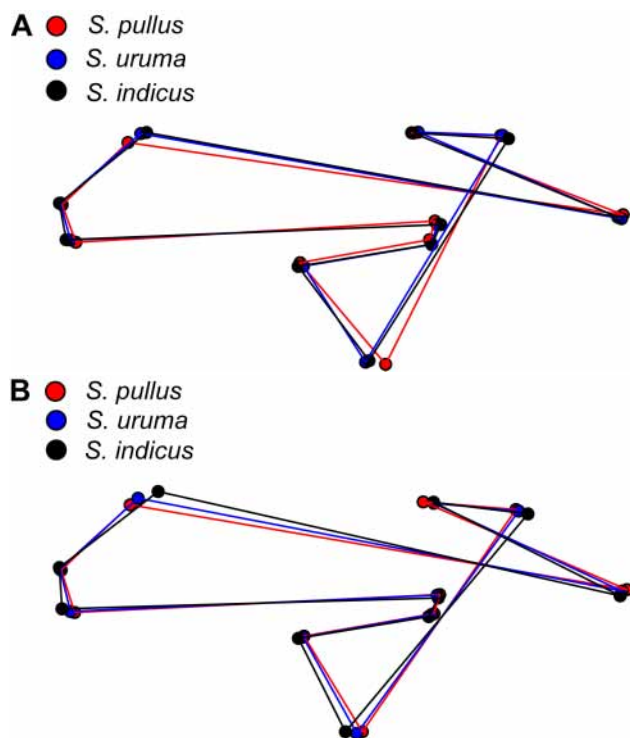
Validate classification

To test the accuracy of species classification yielded by geometric morphometrics, the Mahalanobis distances were used to perform a cross-validated classification (or jackknife classifica-

Table 3. *P*-values of mean centroid size differences among populations of *Stomoxys pullus* Austen, 1909, *S. uruma* Shinonaga et Kano, 1966 and *S. indicus* Picard, 1908.

Species (sex)	<i>S. pullus</i> (m)	<i>S. pullus</i> (f)	<i>S. uruma</i> (m)	<i>S. uruma</i> (f)	<i>S. indicus</i> (m)	<i>S. indicus</i> (f)
<i>S. pullus</i> (m)						
<i>S. pullus</i> (f)	NS					
<i>S. uruma</i> (m)	S	S				
<i>S. uruma</i> (f)	S	S	NS			
<i>S. indicus</i> (m)	NS	NS	S	S		
<i>S. indicus</i> (f)	NS	NS	S	S	NS	

The level of statistic significance was first computed from a non-parametric test, then corrected after Bonferroni test (see Materials and Methods). m – male; f – female; S – significant ($P < 0.05$); NS – not significant ($P > 0.05$).

**Fig. 6.** Configurations of the ten anatomical landmarks connected by a straight line after procrustes superimposition of three species of *Stomoxys* Geoffroy, 1762, in males (A) and females (B).**Table 4.** Landmark-based Mahalanobis distances between wing shapes of *Stomoxys pullus* Austen, 1909, *S. uruma* Shinonaga et Kano, 1966 and *S. indicus* Picard, 1908.

Species (sex)	<i>S. pullus</i> (m)	<i>S. pullus</i> (f)	<i>S. uruma</i> (m)	<i>S. uruma</i> (f)	<i>S. indicus</i> (m)	<i>S. indicus</i> (f)
<i>S. pullus</i> (m)	0.00					
<i>S. pullus</i> (f)	6.68	0.00				
<i>S. uruma</i> (m)	5.64	5.06	0.00			
<i>S. uruma</i> (f)	8.06	3.23	5.05	0.00		
<i>S. indicus</i> (m)	8.04	7.94	5.54	7.43	0.00	
<i>S. indicus</i> (f)	12.83	9.38	8.81	7.54	6.76	0.00

The level of statistic significance was first computed from a non-parametric test, then corrected after Bonferroni test (see Materials and Methods). Mahalanobis distances were highly significant for all pairwise comparisons. m – male; f – female.

tion), in which each individual is allocated to its closest group without being used to help determine a group centre (Manly 2004).

Software

Collections of anatomical landmarks, data analyses and graphical outputs were performed using the various modules of CLIC package version 97 (Dujardin et al. 2010); the Collection of Coordinates (COO) module for collecting landmarks and outlines; the Tabla, Espacios, Texto (TET) module for modifying the data; the Morformetria Geometrica (MOG) module for GPA analyses and generation of CS, PW and RW; the Fourier Outlines Graphics (FOG) module for EFA analyses; the Variation and variance (VAR) module for size analysis and the Permutaciones Analisis Discriminante (PAD) module for shape analysis.

RESULTS

Landmark-based geometric morphometrics

The largest wing (centroid size) was found in female *Stomoxys indicus* (3.88 mm), whereas the smallest wing was found in male *S. uruma* (3.22 mm). The remaining populations had following wing size: 3.87 mm (female *S. pullus*), 3.85 mm (male *S. pullus*), 3.82 mm (male *S. indicus*) and 3.32 mm (female *S. uruma*). The size relationships among samples is illustrated in Fig. 5, and their statistical significance is shown in Table 3.

The visual comparisons of the mean anatomical landmark positions between species and sexes showed most visible landmarks displacements in the upper and lower part of wing (landmarks 1, 7, 9, 10) (Fig. 6). Based on the Mahalanobis distances comparisons, the wing shape was significantly different among species and sexes of the three species of *Stomoxys* (Table 4). The discriminant analysis for the wing landmark-based shape showed that individuals clustered into distinct groups in males, whereas females of *S. pullus* and *S. uruma* showed some overlapping (Fig. 7). The accuracy scores after cross-validated classification test ranged from 76% to 100%, showing better values in males (Table 5).

Outline-based geometric morphometrics

For outline-based method, the largest wing (perimeter) was found in male *S. pullus* (10.95 mm), whereas the smallest wing was found in both male and female *S. uruma* (9.12 mm). The remaining populations had the following wing size: 10.92 mm (male *S. indicus*), 10.84 mm (female *S. indicus*) and 10.52 mm (female *S. pullus*). The size relationships among samples is illustrated in Fig. 8, and their statistical significance is shown in Table 6.

The visual comparisons of contours between species and sexes are shown in Fig. 9. Subtle differences of contours were observed in the males of all three species, and female of *S. pullus* and *S. uruma*. Based on the Mahalanobis distances comparisons, the wing outline-based shape variables were significantly different among species and sexes of all taxa studied (Table 7). In both sexes, the factor map

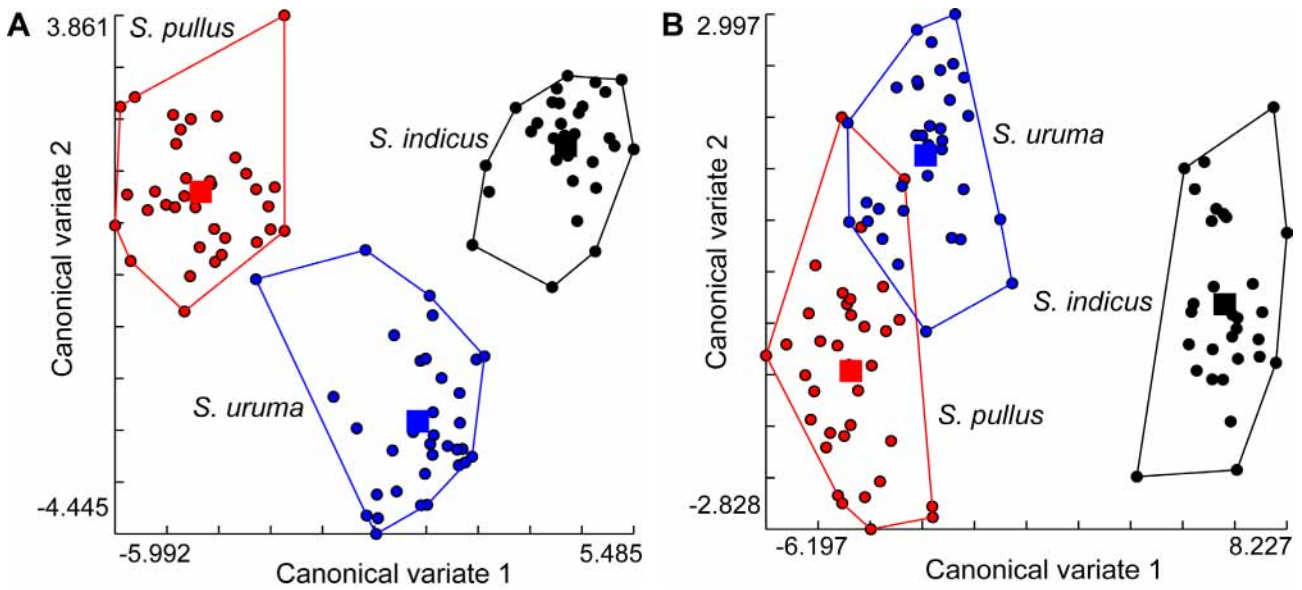


Fig. 7. Landmark-based discriminant analysis. Factor map of canonical variates resulting from comparison among the three species of *Stomoxys* Geoffroy, 1762, in males (A) and females (B).

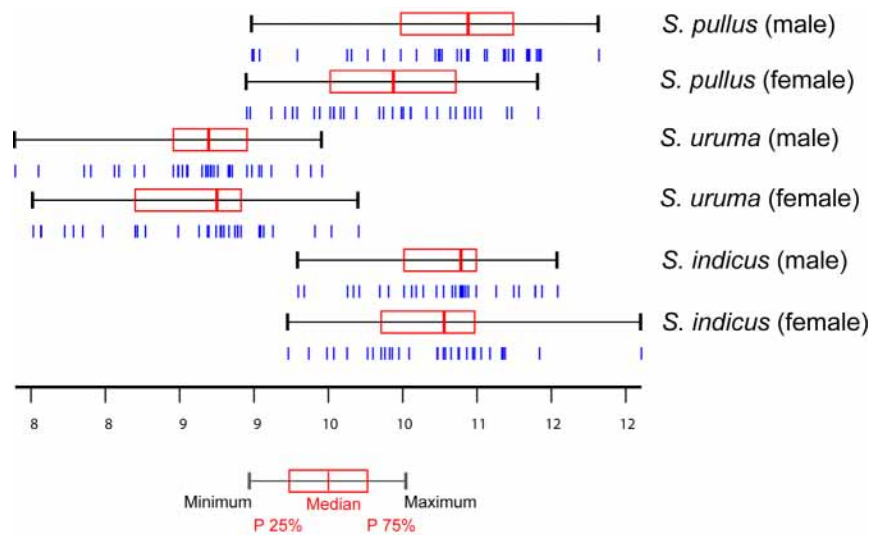


Fig. 8. Perimeter variation of the wings between species and sexes, shown as quartile boxes. Each box shows the group median separating the 25th and 75th quartiles. Vertical bars under the boxes represent the wing (units as mm).

Table 5. Cross-validated classification of *Stomoxys pullus* Austen, 1909, *S. uruma* Shinonaga et Kano, 1966 and *S. indicus* Picard, 1908 based on the shape of the wings.

Species	Landmark-based		Outline-based	
	Male	Female	Male	Female
<i>S. pullus</i>	97% (34/35)	91% (32/35)	91% (31/34)	81% (27/33)
<i>S. uruma</i>	97% (33/34)	76% (26/34)	84% (27/32)	77% (24/31)
<i>S. indicus</i>	100% (30/30)	96% (29/30)	86% (26/30)	96% (29/30)

Table 6. P-values of mean perimeter differences among populations of *Stomoxys pullus* Austen, 1909, *S. uruma* Shinonaga et Kano, 1966 and *S. indicus* Picard, 1908.

Species (sex)	<i>S. pullus</i> (m)	<i>S. pullus</i> (f)	<i>S. uruma</i> (m)	<i>S. uruma</i> (f)	<i>S. indicus</i> (m)	<i>S. indicus</i> (f)
<i>S. pullus</i> (m)						
<i>S. pullus</i> (f)	NS					
<i>S. uruma</i> (m)	S	S				
<i>S. uruma</i> (f)	S	S	NS			
<i>S. indicus</i> (m)	NS	NS	S	S		
<i>S. indicus</i> (f)	NS	NS	S	S	NS	

The level of statistic significance was first computed from a non-parametric test, then corrected after Bonferroni test (see Materials and Methods). m – male; f – female; S – significant ($P < 0.05$); NS – not significant ($P > 0.05$).

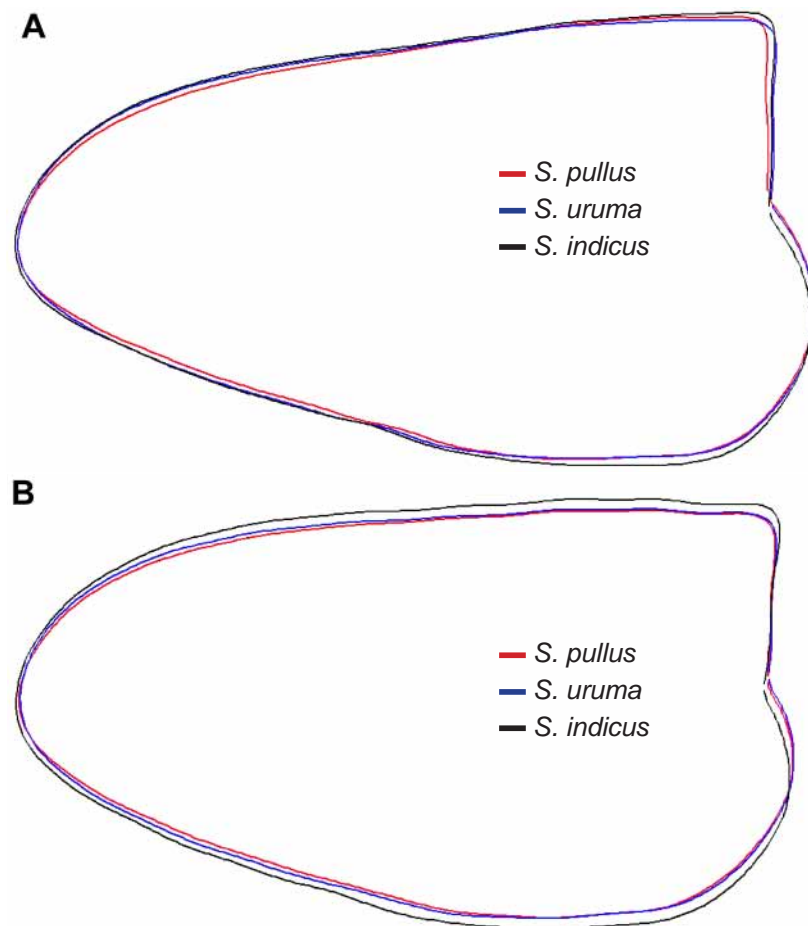


Fig. 9. Configurations of the outlines after Elliptic Fourier Analysis of *Stomoxys pullus* Austen, 1909, *S. uruma* Shinonaga et Kano, 1966 and *S. indicus* Picard, 1908, in males (A) and females (B). Areas outlined by different colours represent shape, not size.

Table 7. Outline-based Mahalanobis distance between outlines of *Stomoxys pullus* Austen, 1909, *S. uruma* Shinonaga et Kano, 1966 and *S. indicus* Picard, 1908.

Species (sex)	<i>S. pullus</i> (m)	<i>S. pullus</i> (f)	<i>S. uruma</i> (m)	<i>S. uruma</i> (f)	<i>S. indicus</i> (m)	<i>S. indicus</i> (f)
<i>S. pullus</i> (m)	0.00					
<i>S. pullus</i> (f)	9.14	0.00				
<i>S. uruma</i> (m)	4.73	7.62	0.00			
<i>S. uruma</i> (f)	9.76	2.66	7.56	0.00		
<i>S. indicus</i> (m)	6.30	8.44	4.55	8.62	0.00	
<i>S. indicus</i> (f)	11.13	6.31	9.20	6.01	8.80	0.00

The level of statistic significance was first computed from a non-parametric test, then corrected after Bonferroni test (see Materials and Methods). Mahalanobis distances were highly significant for all pairwise comparisons. m – male; f – female.

derived from the discriminant analysis for the wing shape showed slightly overlapping areas between *S. pullus* and *S. uruma* (Fig. 10). The accuracy scores after cross-validated classification test ranged from 77% to 96% (Table 5).

DISCUSSION

This research provides new information about the morphology of *Stomoxys* flies. Some species of *Stomoxys*, especially the females of *S. pullus* and *S. uruma*, are very similar and difficult to identify using taxonomic key (Changbunjong et al. 2013). Hence, accurate species identi-

fication of these flies is an important pre-requisite to help in fly control program (Bhakdeenuan et al. 2012). Our results revealed that both landmark and outline-based geometric morphometrics of the wings can distinguish the three species of *Stomoxys* (*S. pullus*, *S. uruma* and *S. indicus*).

The comparison of the wing size by using average centroid size or perimeter of the wing contour showed that the wing size of *S. indicus* and *S. pullus* was not significantly different, but consistently larger than of *S. uruma*. Our results indicated that wing size could help in distinguishing *S. pullus* from *S. uruma*, or *S. indicus* from *S. uruma*. However, wing size can have a major affect from environmental factors such as temperature, relative humidity and food availability (Jirakanjanakit et al. 2007, Morales-Vargas et al. 2010, Ayala et al. 2011). Moreover, *S. indicus* were collected in our study from different sites and seasons compared to *S. pullus* and *S. uruma* that may also have had an affect on wing size (Schachter-Broide et al. 2009, Prudhomme et al. 2012). These environmental factors can influence wing size variable much more than wing shape which often depends on genetic drift and evolutionary divergence (Dujardin 2008, Klingenberg 2010).

The discriminant analysis of landmark and outline-based methods showed that both sexes of *S. indicus* are well separated from *S. pullus* and *S. uruma*. This finding parallels previous results indicating that *S. indicus* can

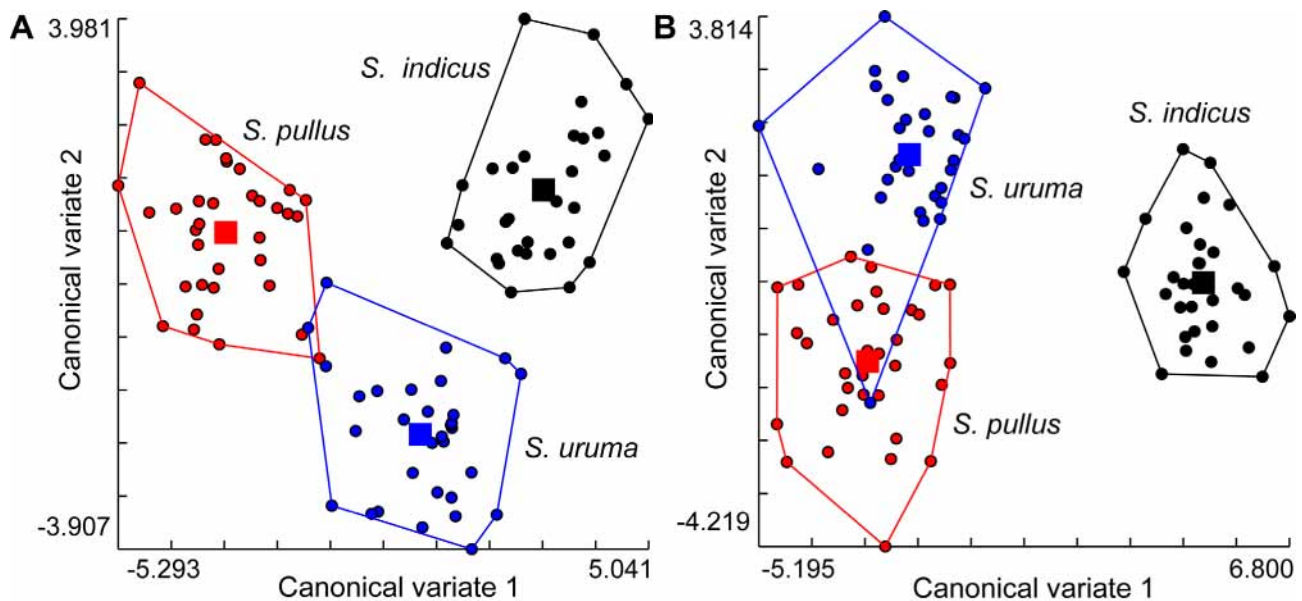


Fig. 10. Outline-based discriminant analysis. Factor map of canonical variates (i.e. discriminant factors) derived from the principal components of the Normalised Elliptic Fourier coefficients of three species of *Stomoxys* Geoffroy, 1762, in males (A) and females (B).

be distinguished by the adult external morphology (yellowish colour of tibiae and tarsi) (Tumrasvin and Shinonaga 1978). Additionally, the phylogenetic relationship based on cytochrome c oxidase subunit I (*COI*) showed that *S. pullus* and *S. uruma* have a closer genetic relationship than either has with *S. indicus* (T.C. – unpubl. data). They were occasionally misidentified, especially in the females as observed after cross-validated reclassification. Males of *S. pullus* and *S. uruma* were, however, quite clearly distinguished, especially by landmark analyses. The reclassification scores based on landmark analyses (76% to 100%) and outline analyses (77% to 96%) were quite similar. These results corresponded to those of the previous study of Dujardin et al. (2014). These authors showed that an outline-based approach could produce similar or even

better discrimination scores than landmarks for various arthropods including kissing bugs, tsetse flies, mosquito and soft ticks (Dujardin et al. 2014).

In conclusion, the landmark and outline-based geometric morphometrics of the wings proved to be a very useful tool to help in the morphological distinction of the vectors *S. pullus*, *S. uruma* and *S. indicus*. They have the potential to improve the vector surveillance, hence the planning of fly control programs.

Acknowledgements. This research is supported by Mahidol University. We would like to thank the Department of National Parks, Wildlife and Plant Conservation for permission to conduct the study in the area.

REFERENCES

- AYALA D., CARO-RIÑO H., DUJARDIN J.P., RAHOLA N., SIMARD F., FONTENILLE D. 2011: Chromosomal and environmental determinants of morphometric variation in natural populations of the malaria vector *Anopheles funestus* in Cameroon. *Infect. Genet. Evol.* 11: 940–947.
- BALDACCHINO F., MUENWORN, V., DESQUESNES M., DESOLI F., CHAROENVIRIYAPHAP T., DUVALLET G. 2013: Transmission of pathogens by *Stomoxys* flies (Diptera: Muscidae). *A review. Parasite* 20: 26.
- BHAKDEENUAN P., SIRIYASATIEN P., PAYUNGPORN S., PREATIVATANYOU K., THAVARA U., TAWATSIN A. 2012: Molecular analysis of medically and veterinary important muscid flies (Diptera: Muscidae) in Thailand. *Thai J. Vet. Med.* 42: 333–342.
- BOOKSTEIN F.L. (Ed.) 1991: *Morphometric Tools for Landmark Data: Geometry and Biology*. Cambridge University Press, New York, 435 pp.
- CHANGBUNJONG T., WELUWANARAK T., RATANAKORN P., MANEON P., APIWATHNASORN C., SUNGVORNYOTHIN S., SRIWICHAI P., SUMRUAYPHOL S., RUANGSITTICHAI J. 2012: Distribution and abundance of Stomoxyni flies (Diptera: Muscidae) in Thailand. *Southeast Asian J. Trop. Med. Publ. Hlth.* 46: 1400–1410.
- CHANGBUNJONG T., WELUWANARAK T., SEDWISAI P., CHAMSAI T. 2013: Stomoxyni fly fauna of the Khao Yai National Park, Thailand. *Asian Pac. J. Trop. Dis.* 3: 348–351.
- CROSSKEY R.W. 1993: Stable flies and horn flies (bloodsucking Muscidae). In: R.P. Lane and R.W. Crosskey (Eds), *Medical Insects and Arachnids*. Chapman & Hall, London, pp. 389–400.
- DEMARI-SILVA B., SUESDEK L., SALLUM M.A., MARRELLI M.T. 2014: Wing geometry of *Culex coronator* (Diptera: Culicidae) from South and Southeast Brazil. *Parasit. Vectors* 7: 174.
- DUJARDIN J.P. 2008: Morphometrics applied to medical entomology. *Infect. Genet. Evol.* 8: 875–890.
- DUJARDIN J.P., KABA D., HENRY A.B. 2010: The exchangeability of shape. *BMC Res. Notes* 3: 266.
- DUJARDIN J.P., KABA D., SOLANO P., DUPRAZ M., MCCOY K.D., JARAMILLO-O N. 2014: Outline-based morphometrics, an overlooked method in arthropod studies? *Infect. Genet. Evol.* 28: 704–714.
- DUJARDIN J.P., KITTHAWEE S. 2013: Phenetic structure of two *Bactrocera tau* cryptic species (Diptera: Tephritidae) infesting

- Momordica cochinchinensis* (Cucurbitaceae) in Thailand and Laos. *Zoology* (Jena) 116: 129–138.
- FRANCOY T.M., FRANCO F.F., ROUBIK D.W. 2012: Integrated landmark and outline-based morphometric methods efficiently distinguish species of *Euglossa* (Hymenoptera, Apidae, Euglossini). *Apidologie* 43: 609–617.
- HEBERT P.D.N., CYWINSKA A., BALL S.L., DE WAARD J.R. 2003a: Biological identifications through DNA barcodes. *Proc. Biol. Sci.* 270: 313–321.
- HEBERT P.D.N., RATNASINGHAM S., DEWAARD J.R. 2003b: Barcoding animal life: cytochrome c oxidase subunit 1 divergences among closely related species. *Proc. Biol. Sci.* 270: S96–99.
- JARAMILLO-O N., DUJARDIN J.P., CALLE-LONDOÑO D., FONSECA-GONZÁLEZ I. 2015: Geometric morphometrics for the taxonomy of 11 species of *Anopheles* (Nyssorhynchus) mosquitoes. *Med. Vet. Entomol.* 29: 26–36.
- JIRAKANJANAKIT N.L.S., THONGRUNGKIAT S., APIWATHNASORN C., SINGHANIYOM S., BELLEC C., DUJARDIN J.P. 2007: Influence of larval density or food variation on the geometry of the wing of *Aedes* (*Stegomyia*) *aegypti*. *Trop. Med. Int. Hlth* 12: 1354–1360.
- KLINGENBERG C.P. 2010: Evolution and development of shape: integrating quantitative approaches. *Nat. Rev. Genet.* 11: 623–635.
- KUHL F.P., GIARDINA C.R. 1982: Elliptic Fourier features of a closed contour. *Comput. Graph. Image Process* 18: 236–258.
- LAVEISSIÈRE C., GREBAUT P. 1990: The trapping of tsetse flies (Diptera: Glossinidae) improvement of a model: the Vavoua trap. *Trop. Med. Parasitol.* 41: 185–192.
- LORENZ C., MARQUES T.C., SALLUM M.A., SUESDEK L. 2012: Morphometrical diagnosis of the malaria vectors *Anopheles cruzii*, *An. homunculus* and *An. bellator*. *Parasit. Vectors* 5: 257.
- MANLY B.F.J. (Ed.). 2004: *Multivariate Statistical Methods: A Primer*. Chapman and Hall/CRC Press, Boca Raton, 224 pp.
- MASMEATATHIP R., KETAVAN C., DUVALLET G. 2006: Morphological studies of *Stomoxys* spp. (Diptera: Muscidae) in central Thailand. *Kasetsart J. (Nat.Sci.)* 40: 872–881.
- MORALES VARGAS R.E., PHUMALA-MORALES N., TSUNODA T., APIWATHNASORN C., DUJARDIN J.P. 2013: The phenetic structure of *Aedes albopictus*. *Infect. Genet. Evol.* 13: 242–251.
- MORALES-VARGAS E.R., YA-UMPHAN P., PHUMALA-MORALES N., KOMALAMISRA N., DUJARDIN J.P. 2010: Climate associated size and shape changes in *Aedes aegypti* (Diptera: Culicidae) populations from Thailand. *Infect. Genet. Evol.* 10: 580–585.
- MUENWORN V., DUVALLET G., THAINCHUM K., TUNTAKOM S., TANASILCHAYAKUL S., PRABARIPAI A., AKRATANAKUL P., SUKONTHABHIROM S., CHAREONVIRIYAPHAP T. 2010: Geographic distribution of *Stomoxys* flies (Diptera: Muscidae) and diurnal activity of *Stomoxys calcitrans* in Thailand. *J. Med. Entomol.* 47: 791–797.
- MÜLLER P., PFLÜGER V., WITTEWIT M., ZIEGLER D., CHANDRE F., SIMARD F., LENGELER C. 2013: Identification of cryptic *Anopheles* mosquito species by molecular protein profiling. *PLoS ONE* 8: e57486.
- PRUDHOMME J., GUNAY F., RAHOLA N., OUANAIMEI F., GUERNAOUI S., BOUMAZZOUH A., BANULS A.L., SERENO D., ALTEN B. 2012: Wing size and shape variation of *Phlebotomus papatasi* (Diptera: Phlebotomidae) populations from the south and north slopes of the Atlas Mountains in Morocco. *J. Vector Ecol.* 37: 137–147.
- ROHLF F.J. 1990: Rotational fit (Procrustes) methods. In: F.J. Rohlf and F.L. Bookstein (Eds.), *Proceedings of the Michigan Morphometrics Workshop*. Number 2, Ann Arbor, Michigan. The University of Michigan Museum of Zoology, Special Publication No. 2: 227–236.
- RUANGSITTICHAI J., APIWATHNASORN C., DUJARDIN J.P. 2011: Interspecific and sexual shape variation in the filariasis vectors *Mansonia dives* and *Ma. bonneae*. *Infect. Genet. Evol.* 11: 2089–2094.
- SCHACHTER-BROIDE J., GÜRTLER R.E., KITRON U., DUJARDIN J.P. 2009: Temporal variations of wing size and shape of *Triatoma infestans* (Hemiptera: Reduviidae) populations from northwestern Argentina using geometric morphometry. *J. Med. Entomol.* 46: 994–1000.
- SUMRUAYPHOL S., APIWATHNASORN C., RUANGSITTICHAI J., SRIWICHAI P., ATTRAPADUNG S., SAMUNG Y., DUJARDIN J.P. 2016: DNA barcoding and wing morphometrics to distinguish three *Aedes* vectors in Thailand. *Acta Trop.* 159: 1–10.
- TUMRASVIN W., SHINONAGA S. 1978: Studies of medically important flies in Thailand V. on 32 species belonging to the subfamilies Muscinae and Stomoxyinae including the taxonomic keys (Diptera: Muscidae). *Bull. Tokyo Med. Dent. Univ.* 25: 201–227.
- VILLEGAS J., FELICIANGELI M.D., DUJARDIN J.P. 2002: Wing shape divergence between *Rhodnius prolixus* from Cojedes (Venezuela) and *Rhodnius robustus* from Merida (Venezuela). *Infect. Genet. Evol.* 2: 121–128.
- ZUMPT F. (Ed.). 1973: *The Stomoxyine Biting Flies of the World*. Gustav Fischer Verlag, Stuttgart, 175 pp.

Received 6 June 2016

Accepted 12 September 2016

Published online 24 October 2016

Cite this article as: Changbunjong T., Sumruayphol S., Weluwanarak T., Ruangsittichai J., Dujardin J.-P. 2016: Landmark and outline-based geometric morphometrics analysis of three *Stomoxys* flies (Diptera: Muscidae) in Thailand. *Folia Parasitol.* 63: 037.