Introduction

Tropical rain forests are the most biodiverse terrestrial ecosystems on Earth, containing over 50% of known terrestrial biodiversity packed in just 7-10% of the Earth's surface [1,2]. The level of biodiversity is remarkable both locally (alpha diversity) and in terms of variation in space (beta diversity; e.g. [3,4]). Despite the importance of rain forests for terrestrial biodiversity, the drivers of diversity gradients within and between the world's main rain forest areas remain poorly understood [5,6]. In addition, large areas of rain forests, among which the central African block, are poorly explored by scientists, giving a fragmentary view of spatial diversity patterns, even for well-studied organisms such as plants [7].

In the last century, rain forests have been overexploited in many parts of the world leading to their alteration, fragmentation, and in some areas, complete destruction [7]. The consequences of these changes include biodiversity loss and increased atmospheric carbon dioxide concentration resulting in climate change, due to the conversion of high-carbon storage forest to low-carbon storage agriculture [8]. As a consequence, there is an urgent need to better understand the processes that sustain the biological diversity in tropical rain forests [9].

Biodiversity is classically assessed at species level (e.g. [10]), from the observation of the presence/absence of species (i.e. species occurrence) or species abundance in transects or ecological plot surveys (e.g. [11]). However, biodiversity assessments based on species counts and their relative abundance statistics provide little information regarding the functional diversity of the ecosystem under study, since they do not acknowledge the variation in their ecological niches [12,13,14,15]. Estimation of phylogenetic or functional diversity in addition to species diversity has been recognized as improving our understanding of the niche-based...
processes leading to the observed patterns of present day biodiversity [15,16]. Those estimations also help to better conserve phylogenetic relatedness (e.g. [14,15]). In this context, phylogenetic relatedness is classically considered as a proxy of functional relatedness, because the closer the species are in the phylogeny, the more likely they have inherited similar traits from a common ancestor. As a consequence the consideration of phylogenetic diversity informs on ecosystem functioning and adaptability. This ‘ecophylogenetic’ approach is therefore a relevant basis for conservation purposes [17].

Recently, progress in phylogeny reconstruction either from DNA sequences or from existing resources has accelerated theoretical and methodological advances in ecophylogenetics [18]. The number of studies on phylogenetic alpha and beta diversity in tropical tree communities has increased in the last two or three years [9]. Those studies have focused on partitioning diversity into spatial and environmental components (e.g. [19,20,21]) or by comparing the alpha and beta components of phylogenetic diversity (e.g. [18,20]).

Recent studies carried out on tropical rain forest trees reported that (i) phylogenetic turnover (i.e. a spatial turnover of the dominance of clades) is associated with habitat or environmental differentiation [19,20,22,23]; (ii) functional traits [22] and climate niche proxies [23] usually display a significant, although sometimes weak, phylogenetic structure; (iii) phylogenetic turnover parallels functional turnover, although with a weaker strength [22]. It is noteworthy that most of these studies were carried out in regions with substantial climatic and/or edaphic gradients, so that environmental filtering effects favoring functional and phylogenetic clustering may predominate over competitive exclusion effects that might lead to functional and phylogenetic over-dispersion (but see [24]).

In this paper we will focus on a region located in the margin of the Congo Basin which is home to the second largest tropical rain forest after the Amazon basin, with a high level of species diversity and endemism [25]. In recent years, several countries have created national parks in an effort to conserve rain forest biodiversity (e.g. Gabon). The Dja Faunal Reserve (DFR) is a UNESCO world heritage reserve of 526,000 ha located 250 kilometers south east from the Cameroon capital Yaoundé. The reserve was established in 1950 and is the largest protected rain forest in Cameroon [26]. There is no steep climatic or edaphic macrogeographic gradient across the DFR. However, its topography characterized by half-orange shaped hills generates a heterogeneity of soil hydromorphy, so that distinct edaphic habitats can be recognized. Besides, natural or past human disturbances (gap dynamics; ancient agricultural fields) are recognized locally by affecting the structure of the vegetation. In a previous study, Hardy and Sonke [27] assessed the role of dispersal and niche differentiation in shaping tree species turnover along nine transects covering the DFR. To this end, they quantified the impact of spatial distance and habitat differentiation on the probability that pairs of individuals are conspecific. With the exception of pioneer species, they found a pattern of isolation by distance, i.e. spatial species clustering due to the combined effect of limited dispersal and local ecological drift [10]. Habitat differentiation was also found to be a major determinant of the spatial pattern but had a lower impact than spatial distance per se. These results suggest that in the DFR, the degree of species aggregation might be better determined by dispersal-assemblage rules rather than by niche-assembly rules, at least for the common species. Because this study, which was based on taxonomic diversity, did not take into account phylogenetic diversity between species, a reanalysis of the dataset used in Hardy and Sonke [27], but accounting for phylogenetic variation will provide further insights in how niche-based processes constrains the composition of communities [23]. Because macrogeographic environmental gradients are weak across the DFR compared to most previous works carried out at a similar scale in tropical rain forests, patterns of phylogenetic structure might differ.

The main objective of the paper was to investigate the relative importance of niche-based and dispersal-based processes governing tree species assemblage within the Dja Faunal Reserve. We used an ecophylogenetic approach [14,15] to provide a phylogenetic quantification of biodiversity in this area for better conservation strategies. We wanted to address the question: can we detect phylogenetic or species turnover across the reserve? Specifically, we elaborated our approach to test the following four hypotheses:

<table>
<thead>
<tr>
<th>Hypothesis</th>
<th>Permutations</th>
<th>Tests</th>
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<tbody>
<tr>
<td>(i) If community assembly is dominated by limited dispersal, no phylogenetic structure should be detected among transects, while isolation-by-distance is expected in taxonomic beta diversity</td>
<td>Model 1-3x Whole dataset - transect. Permutation of individuals between transects within habitat types.</td>
<td>$I_{ST}$</td>
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<tr>
<td>(ii) Isolation-by-distance is expected in taxonomic beta diversity.</td>
<td>Model 1 s Whole dataset-transect. Permutation of species in phylogeny.</td>
<td>$B_{ST} \cdot \Pi_{ST}$</td>
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<td>(iii) If competitive exclusion/niche differentiation prevents the co-occurrence of related species locally, generating a patchwork distribution of functionally equivalent species, phylogenetic overdispersion might be detected within transects, at least within a habitat type</td>
<td>Model 1 s - Whole dataset-habitat. Permutations of species in phylogeny.</td>
<td>$B_{ST} \cdot \Pi_{ST}$</td>
</tr>
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<td>(iv) Environmental filtering and competitive exclusion may simultaneously occur and cancel out to yield apparently “neutral” patterns</td>
<td>Model 1 s - TPF. Permutations of species in phylogeny within TPF only.</td>
<td>$B_{ST} \cdot \Pi_{ST}$</td>
</tr>
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</table>

Table 1. Permutation used to test taxonomic and phylogenetic structure in relation with our hypotheses.
(i) If community assembly is dominated by limited dispersal, no phylogenetic structure should be detected between transects, while isolation-by-distance is expected in taxonomic beta diversity.

(ii) If environmental filtering differed among habitat types a pattern of phylogenetic clustering in habitat should be detected. This interpretation assumes phylogenetic niche conservatism of relevant traits [22,23].

(iii) If competitive exclusion prevents the co-occurrence of related species locally, generating a patchwork distribution of functionally equivalent species, phylogenetic overdispersion might be detected within transects, at least within a habitat type.

(iv) Environmental filtering and competitive exclusion may simultaneously occur and cancel each other out to yield apparent “neutral” patterns.

A critical issue for testing the relative imprint of these processes is to define sampling units that are relevant according to the scale of the processes. Here we considered the nine transects of Hardy and Sonké [27] that provide information on forest tree composition at two levels: among transects and among habitat types.

To address the four hypotheses above, we therefore partitioned taxonomic and phylogenetic diversity within and between transects, as well as within and between habitat types. For this we applied the statistical framework developed by Hardy and Senterre [20] for characterizing and testing the phylogenetic structure of transects and habitats types using appropriate randomization procedures [28] (Table 1, Figure 1).

Figure 1. Schema of the 3 types of randomization used to test taxonomic and phylogenetic structure. In model 1-3x, individuals were randomized among transects or species within each habitat type (a). In model 2-3x, individuals or species were randomized among habitats (c). These models of permutation aimed to test for taxonomic turnover using \( s_{T} \). Phylogenetic structure (\( B_{T} \) and \( \Pi_{T} \)) was tested using a model 1 s which randomized the observed species across the tips of the phylogenetic tree (b, d, e). Randomization were respectively done for the whole dataset (a,b), for the habitat data set (c, e) and for TPF only (d).

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Materials and Methods

1. Study site and tree communities

The Dja Faunal Reserve (DFR) is situated between latitudes 2°50’–3°30’ N and longitudes 12°20’–13°40 E in southeastern Cameroon. About two-thirds of the reserve’s perimeter is demarcated by the Dja River, forming a natural boundary. In this reserve, tree species have been inventoried along nine 5 km long and 5 m wide transects (Figure 2), in which all species with a diameter at breast height bigger than 10 cm were identified and mapped. This dataset (sampling, preliminary taxonomic analysis) is described in Sonke [29] and Sonke and Couvreur [30]. In total 11546 individuals were inventoried belonging to 312 identified species and 60 taxa identified to generic level only (and considered as a morphospecies, 372 total taxa included in the analyses). All nomenclatural criteria regarding species names and families followed Sonke and Couvreur [30]. The vegetation in the reserve has a 30–40 m tall canopy with emergent trees rising up to 60 m [26]. Detailed descriptions of the structure and species composition of the mixed forests can be found in Sonke [31] and Sonke and Couvreur [30].

The nine transects were established across five broad types of forest (i.e. habitat) depending on soil hydromorphy and vegetation structure [32]. Terra firme forests were established on non-hydromorphic soils and subdivided into three successional types, namely (i) terra firme primary forest (74% of total individuals); (ii) secondary forest (8% of total individuals); (iii) gaps (4% of total individuals). Conversely, two hydromorphic types were defined as (iv) swamps (11%) and (v) flooded forest (3%).

We considered two sampling levels for subsequent phylogenetic and taxonomic composition analyses. First, we analyzed the variation within and between the 9 transects, such as the comparison of transects represented the largest spatial scale of beta variation. Second, the variation across the five habitats within transects conveyed the imprint of local niche-based processes. All analyses were performed on three datasets: 1) all individuals within each transect (= whole data - transect); 2) only individuals found in habitat (i) terra firme primary forest (= “TPF”) within each transect; 3) the five different habitats were treated independently of transects (i.e. transects were merged = “habitat”) (= whole data – habitat). This allowed us to test the different assumptions stated in the introduction (Table 1). Our analysis is different than the one of Hardy and Sonke [27] who estimated the probability that two randomly chosen individuals belong to the same species according to the distance separating them (1) on the combined samples of all transects using individual coordinates and then (2) within the three main habitat types.

2. Phylogenetic relationships of the DFR tree community

A phylogenetic tree of 372 species was generated in three steps. First, all species were grafted onto a comprehensive phylogenetic tree using the program PHYLOMATIC v3 [33] (http://
phyldiversity.net/phylomatic/). The program generated a tree in which the family relationships of the sampled species followed the angiosperm phylogeny APG III [34] version R20120829. We then manually resolved the generic relationships within most of the families based on specific molecular phylogenies (Table 2), using the software Mesquite [35]. Only relationships that were supported with bootstrap values of more than 70% were taken into account. For families where no phylogenetic information was available or for which the published phylogeny did not provide enough insights into the relationships between genera, generic relationships were left unresolved (polytomies). Finally, we used the branch adjustment algorithm BLADJ implemented in Phylocom [36] to scale the branch lengths based on a set of node age estimates from several publications (Table 2). For this part, we first used the dated phylogeny of Wikstrom et al [37] for major nodes. We also used family level dated trees to further constrain certain nodes (Table 2). Intra- and interspecific branch lengths were assumed to be 0 (i.e. relationships between species and within species are unknown and unresolved).

3. Species and phylogenetic structure analyses

We used the measures of phylogenetic distinctness and differentiation within and between transects/habitats introduced by Hardy & Senterre [20]. These statistics are based on the additive partitioning of Rao entropy [38], which lead to differentiation coefficients between transects/habitats that are analogous to the coefficients expressing genetic differentiation among populations in population genetics.

Tests of phylogenetic structure can be biased when there is a non-random phylogenetic distribution of species abundance at regional scale (i.e., in the overall dataset) [28]. Therefore, to test if abundant species were randomly distributed across the phylogeny, we first calculated the Abundance Phylogenetic Deviation (APD) statistic [28]. When APD <0, species abundances are over-dispersed, whereas when APD >0, species abundances are clustered (abundant species mainly belong to one or a few clades).

We re-estimated taxonomic diversity for phylogenetic analyses since we used a different strategy from the one used in Hardy and Sonké [27]. We calculated the probabilities that two individuals belonged to different species (Simpson-Gini diversity index) within a transect/habitat (\(D_{IS}\)) and between transects/habitats (\(D_{IT}\)), as well as the mean phylogenetic distances (based on the divergence time) between individuals (an index of phylogenetic diversity) within transect/habitat (\(D_{PS}\)), and between transects/habitats (\(D_{PT}\)). \(I_{ST} = (D_{IT}-D_{IS})/D_{IT}\) then expresses the species turnover between transects/habitats. Taxonomic clustering in transects/habitat is expected to be reflected by \(I_{ST}>0\), while taxonomic overdispersion should result in \(I_{ST}<0\). \(P_{ST} = (D_{PT}-D_{PS})/D_{PT}\) expresses the combined effect of species and phylogenetic turnover. However, as it is difficult to interpret, we do not consider this quantity in our interpretations. In addition, we estimated the mean phylogenetic distances between two non-conspecific individuals sampled at local (i.e. within transect/habitat) or regional scale (i.e. between transects/habitats), respectively denoted as \(D_{BS}\) and \(D_{BT}\), so that \(B_{ST} = (D_{BT}-D_{BS})/D_{BT}\) expressed phylogenetic turnover between transects/habitats independently of species turnover [38]. \(B_{ST}>0\) under local phylogenetic clustering while \(B_{ST}<0\) under local phylogenetic overdispersion.

These estimators require abundance data, and rare species are underemphasized, while the distribution of rare species can also bring useful information on species assembly rules. Thus, we used measures of phylogenetic distinctness based on species incidence [20]. \(A_{PS}\) is defined as the mean phylogenetic distance between distinct species within transects/habitats and \(A_{PT}\) between transects/habitats (i.e. mean phylogenetic distance between

<table>
<thead>
<tr>
<th>Family</th>
<th>Phylogenetic relationships</th>
<th>Calibration points</th>
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<tbody>
<tr>
<td>angiosperms</td>
<td>[37]</td>
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<tr>
<td>Annonaceae</td>
<td>[52]</td>
<td>[48]</td>
</tr>
<tr>
<td>Apocynaceae</td>
<td>[58]</td>
<td></td>
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<tr>
<td>Bignoniaceae</td>
<td>[59]</td>
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<tr>
<td>Chrysobalanaceae</td>
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<tr>
<td>Clusiaceae</td>
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</tr>
<tr>
<td>Combretaceae</td>
<td>[62]</td>
<td></td>
</tr>
<tr>
<td>Euphorbiaceae</td>
<td>[63]</td>
<td></td>
</tr>
<tr>
<td>Leguminosae: Caesalpinioideae</td>
<td>[64];</td>
<td></td>
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<tr>
<td>Leguminosae: Mimosoideae</td>
<td>[65]</td>
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<tr>
<td>Malvaceae</td>
<td>[66] and <a href="http://www.malvaceae.info">http://www.malvaceae.info</a></td>
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<tr>
<td>Meliaceae</td>
<td>[67,68]</td>
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<tr>
<td>Myristicaceae</td>
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<tr>
<td>Mytaceae</td>
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<td>Oleaceae</td>
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</tr>
<tr>
<td>Rutaceae</td>
<td>[73]</td>
<td></td>
</tr>
<tr>
<td>Sapindaceae</td>
<td>[74]</td>
<td></td>
</tr>
</tbody>
</table>

Table 2. References to phylogenetic trees and chronograms used to manually resolve relationships and identify calibration points in families with three or more species sampled in this study.
distinct species sampled from two transects/habitats, averaged over all pairs of transects/habitats. Hence a coefficient analogue to $B_{ST}$ is defined, $\Pi_{ST} = (\Delta_{PT} - \Delta_{PS})/\Delta_{PT}$ which expresses phylogenetic turnover by the gain of phylogenetic distance between species occurring in different sites compared with species occurring in the same site. $\Pi_{ST}$ is equivalent to $B_{ST}$ but neglects species abundances. This coefficient excludes comparisons of a species with itself. All the estimations were performed using the software SPACoDi [39].

4. Testing species and phylogenetic structure

To test for species turnover and phylogenetic structure, we used 3 models of randomization (Table 1, Figure 1). In model 1–3x, we randomized individuals among transects or species within each habitat type (Figure 1a). In model 2–3x, we randomized individuals or species among habitats (Figure 1c). These models of permutation aimed to test for taxonomic turnover using $I_{ST}$. The model 3x in which community composition was randomized but not the position of taxa in the phylogeny has been shown to be biased to test the phylogenetic structure [28]. Thus, to test for phylogenetic structure, we used a third permutation model (model 1s, Figure 1b, d, e). The model 1s randomizes the observed species across the tips of the phylogenetic tree and allowed testing for phylogenetic structure using $B_{ST}$ and $\Pi_{ST}$ (Table 1).

We undertook 999 permutations for each model, providing 999 estimations of the above differentiation coefficients under those null models. Deviations of observed coefficient from random coefficients were used to test whether $I_{ST} = 0$, $B_{ST} = 0$ or $\Pi_{ST} = 0$. A significant test for $I_{ST}$ is expected at least under hypothesis (i) between transects or habitats with the model 1–3x. Under hypothesis (ii) (habitat filtering dominates), we expect $B_{ST} > 0$ and $\Pi_{ST} > 0$ with the whole dataset-habitat (model 1s); under hypothesis (iii) (competitive exclusion between related species dominates), we expect $B_{ST} < 0$ and $\Pi_{ST} < 0$ but transects at least within TPF habitat; while no phylogenetic significant tests should be obtained under hypotheses (i, neutral assembly with limited dispersal) and (iv, compensation between ii and iii) (Table 1). Mantel tests were used to test the relations between pairwise taxonomic ($I_{ST}$) or phylogenetic distances ($B_{ST}$ and $\Pi_{ST}$) and geographic distances among the 9 transects using the R package vegan [40]. A significant test with $I_{ST}$ but not with $B_{ST}$ or $\Pi_{ST}$ is expected under hypothesis (i).

Finally, to assess the robustness of the results with respect to the taxonomic scale investigated, and possibly assess whether hypothesis (iv, compensatory effects between habitat filtering and competitive exclusion) might hold, partial randomization of the data between transects was performed on certain clades defined as species rich which were arbitrarily defined as families containing 10 or more sampled species. We also looked at Eudicot and Magnoliidae clades. For each clade the coefficients described above were calculated under the 1s model (999 randomization of tree tips). This was done by using the spacodi.per.nodes function in the SpacodiR [41].

Results

1. Phylogenetic tree of the DFR

For 17 families, phylogenetic studies allowed the resolution of most relationships between genera (Table 2). A total of 25 calibration points (Table 2) were used to generate the dated phylogenetic tree of the DFR. The tree was produced using the iTOL web application [42,43] (Figure 3).

2. Species and phylogenetic structure analyses

The Abundance Phylogenetic Deviation (APD) estimations were not significant for the 3 datasets (Table 3) indicating that abundant species were randomly distributed across the phylogeny at the scale of the study area.

The probability that two randomly selected individuals belonged to different species ($D_{PS}$) was high for all our 3 datasets (0.9805, 0.9789 and 0.9798) for the whole dataset-transect, TPF and the whole dataset-habitat respectively (Table 2). The mean divergence time between individuals was $D_{PS} = 129.49$ million years (Myr), 129.83 Myr and 126.84 Myr respectively for the three datasets. The mean divergence times between species ($D_{BS}$) was 129.75 Myr, 130.01 Myr and 128.97 Myr (Table 2). According to these coefficients, most diversity occurred within transect or habitat, the between contribution being always less than or equal to 0.6% for the whole dataset-transect ($I_{ST} = 0.0067$, $B_{ST} = 0.0002$, $\Pi_{ST} = 0.0002$, $0.7\%$ for TPF ($I_{ST} = 0.0079$, $B_{ST} = 0.0002$, $\Pi_{ST} = 0.0004$) and 1.1% for the whole dataset-habitat ($I_{ST} = 0.0079$, $B_{ST} = 0.0002$, $\Pi_{ST} = 0.0003$). When the coefficients were calculated using taxonomic ranks to produce a surrogate of phyletic distances, estimates of phylogenetic distinctness ($D_{PS}$ and $D_{PT}$) were only slightly different, and estimates of phylogenetic differentiation between transects ($\Pi_{ST}$) were only slightly affected (Table 3).

The distribution of divergence times between individuals within a transect and within habitat showed that more than half of the pairs of individuals diverged between 160 and 179 Myr ago (Figure 4).

3. Testing species and phylogenetic structure

Whatever the randomization model used (models 1 or 2–3x), all the tests done on taxonomic differentiation were significant ($I_{ST} > 0$), indicating species turnover (Table 4). In the case of model 1–3x, on the whole dataset at transect level, the test indicated limited dispersal between transects within habitat. The model 2–3x on the whole dataset at habitat level suggested that the turnover is also due to a habitat effect because in the habitat dataset geographic distances are meaningless. Concerning tests on phylogenetic structure, we found no significant deviations from random of the observed phylogenetic statistics under model 1 s at the transect level, so that $B_{ST} = 0$ and $\Pi_{ST} = 0$ for the whole dataset-transect and for the TPF dataset. When analyzing the whole dataset at habitat level, the model 1 s still indicated $\Pi_{ST} = 0$, but (marginally) significant $B_{ST} > 0$ indicating low phylogenetic turnover for abundant species among habitats (Table 4).

Mantel tests indicated that pairwise taxonomic distances ($I_{ST}$, species turnover across space) were significantly correlated to pairwise geographic distances both for the whole dataset-transect as well as for TPF only ($r = 0.74$, $p\text{-val}=9x10^{-6}$ and $r = 0.77$, $p\text{-val}=0.0013$). The relations between pairwise phylogenetic and geographic distances were never significant.

4. Variation in phylogenetic structure across the phylogeny

Ten families (sensu APG III) contained ten or more species [30]. Out of those, only two families exhibited a significant phylogenetic turnover between transects ($I_{ST}$ or $P_{ST}$) under the null model 1 s: Annonaceae ($B_{ST} = 0.0009$, p-value $= 0.04$; $I_{ST} = 0.0256$) and Apocynaceae ($B_{ST} = -0.0004$, p-value $= 0.019$; $I_{ST} = 0.0241$). Hence, small but significant phylogenetic clustering was identified in Annonaceae (as $B_{ST}$ is positive); whereas overdispersion was detected in Apocynaceae (as $B_{ST}$ is negative). This indicated that
Annonaceae species were more related within transects than species taken from different transects, while Apocynaceae species were more related among than within transects. Finally, our results show no significant phylogenetic turnover for most of the species rich families (8 out of 10) indicating a neutral pattern at the transect level, and probably contributing mostly to the global neutral observed pattern.

**Discussion**

The present study aimed to better understand the processes underlying taxonomic and phylogenetic diversity and turnover in the Dja Faunal Reserve. We investigated both the alpha and beta diversity components in order to infer mechanisms of local community assembly, as well as the nature of the turnover of species across space, based from the analysis on 9 transects.
Table 3. Partition of taxonomic and phylogenetic diversity within and between the 9 transects from the Dja Faunal Reserve (11538 individuals belonging to 372 species) for the whole dataset and for TPF only.

<table>
<thead>
<tr>
<th>Whole dataset</th>
<th>APD = −0.034562</th>
<th>(pval = 0.0750)</th>
</tr>
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<tbody>
<tr>
<td>Coefficients based on</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Average within site diversity</td>
<td>DWA = 0.9805</td>
<td>Total diversity (γ = γ(y))</td>
</tr>
<tr>
<td>Species identity and abundance</td>
<td></td>
<td>IST = 0.0067</td>
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<tr>
<td>Species phylogeny and abundance</td>
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<td>DTP = 130.3916</td>
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<tr>
<td></td>
<td>DWP = 132.097</td>
<td>DTP = 132.097</td>
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<tr>
<td>Species phylogeny and incidence</td>
<td>ΔPS = 129.7548</td>
<td>ΔPT = 129.7781</td>
</tr>
<tr>
<td>TPF dataset</td>
<td>APD = −0.030372</td>
<td>(pval = 0.0880)</td>
</tr>
<tr>
<td>Coefficients based on</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Local diversity</td>
<td>DWA = 129.8353</td>
<td>Total diversity (γ = γ(y))</td>
</tr>
<tr>
<td>Species identity and abundance</td>
<td></td>
<td>PST = 0.0081</td>
</tr>
<tr>
<td>Species phylogeny and abundance</td>
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<td>DTP = 130.8939</td>
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<td>DWP = 132.64</td>
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<td>ΔPS = 130.0129</td>
<td>ΔPT = 130.0629</td>
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<tr>
<td>Whole dataset-habitat</td>
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<td>(pvalue = 0.758)</td>
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<tr>
<td>Local diversity</td>
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<tr>
<td>Species phylogeny and abundance</td>
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<tr>
<td></td>
<td>DWP = 129.4628</td>
<td>DTP = 129.8765</td>
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<tr>
<td>Species phylogeny and incidence</td>
<td>ΔPS = 128.9733</td>
<td>ΔPT = 129.0418</td>
</tr>
</tbody>
</table>

TPF: terra firme primary forest; APD: mean abundance phylogenetic deviation index. doi:10.1371/journal.pone.0098920.t003

The existence of a dispersal limitation is further confirmed by the species turnover increasing with spatial distance (isolation by distance) for both datasets at transect levels, while the phylogenetic turnover is insensitive to distance. Using only taxonomic data for the same set of species and with different statistics, Hardy and Sonké [27] also found isolation by distance to be a major driver of community variation in the DFR. This pattern is consistent with a neutral model of community dynamics assuming species equivalence in terms of patterns but not automatically in terms of processes [44]. Previous analyses of beta diversity in tropical forests confirmed that the taxonomic turnover found between forest plots separated by 0.2 to 50 km are consistent with the expectation of the neutral model [10], indicating that dispersal is an important factor in community assemblage rules in these ecosystems. The sampled transects covered most of the delimited DFR area with a maximum distance between sites of 105 km (Figure 2) which justifies interpreting the results at the scale of the whole reserve. As phylogenetic turnover in tropical tree communities is expected to be related to species sorting along an environmental gradient (e.g. [22,23]), the absence of phylogenetic turnover among transects indicates that, at coarse grain, no environmental variation influenced the variation in forest composition across space. This would agree with the overall homogeneity of environmental conditions found in the DFR [27]. For example, elevation, which has been shown to be an important factor in species composition in tropical rain forests [20], varies between 600 and 700 meters, and does not generate significant environmental variation. Thus even though there is a significant species turnover across the reserve, phylogenetic relatedness remains equivalent at different places across the Dja rejecting our hypothesis (iii) of the presence of competitive exclusion, and confirming our hypothesis (i), that our communities are marked by dispersal assembly rules. However, we still need to be cautious with our conclusions since an alternative explanations of the fact that no phylogenetic sorting was found among transects might actually not have anything to do with community assembly mechanisms, but rather with the resolution of the phylogeny. Because relationships are only resolved to family/genus level, if species belonging to the same genus are functionally distinct and are sorted among sites we would not be able to tell it with this phylogeny.

Even though our dataset contained only 9 transects, which is fairly limiting, we nevertheless have an extensive coverage of the reserve (Figure 2), and have sampled over 11 000 individuals for 372 species or morphospecies. In addition, significant values of $I_{ST}$ and significant Mantel tests between species and geographic distances among transects indicated that spatial variation among species has been captured in the study. $I_{ST}$ values pointed however towards lower values than those observed for more fragmented wet forests such as in the Western Ghats of India, where $I_{ST}$ values...
were found above 0.027 (computations from the results of [45], against 0.0067 to 0.0079 here (Table 3); \( I_{ST} \) is an increasing function of relative community differentiation). The same pattern is observed when considering \( P_{ST} \) (incidence data integrating phylogeny) which is \( 2 \times 10^{-4} \) in the Dja Reserve against \( 34 \times 10^{-4} \) for the dataset in India and \( 13 \times 10^{-4} \) in the Panama Canal watershed [23]. This may indicate that differentiation between localities is not very pronounced in the continuous Dja forest.

2. Phylogenetic structure among habitat types in the Dja Faunal Reserve

At habitat level, a signal of phylogenetic turnover was barely significant (P-value = 0.05) with abundance data but not with incidence data (\( B_{ST} > 0 \) and \( \Pi_{ST} \sim 0 \). Since the most constraining habitat types are of limited extent, this may explain why there are some significant patterns with abundance data and not with incidence data. This result suggests that species are sorting not just because of limited colonization but also because of environmental variation due to habitat heterogeneity. Indeed, phylogenetic
turnover was expected among habitats because some of the environmental gradients distinguishing between habitat types in the reserve, such as water availability and anoxic stresses (flooded or swamp vegetation versus terra firme vegetation), are known to be strong filtering factors influencing tropical forest community structures [19]. This confirms our second hypothesis (ii): environmental filtering differs among habitat types and relevant selected traits could be phylogenetically conserved although this would have to be further tested using traits dataset.

3. Variation in phylogenetic structure across the phylogeny

To date few studies have investigated community phylogenetic structure in African rain forests. In a study of 28 one-hectare plots in mature rain forest in Monte Aleón National Park (Equatorial Guinea), Hardy and Senterre [20] found a phylogenetic clustering structure which was attributed to adaptations of local species to elevation gradients. They also found that most of the signal is related to ancient clade subdivisions, with most of the individuals pairs (between and among plots) occurring between 100 and 120 Myr. In our analysis, we find a comparable situation where most subdivisions between individual pairs occur between 140–160 Myr. The difference could be related to the different calibrations used to date of the phylogenetic trees. Hardy and Senterre [20] used ages from Davis et al. [46] while we used the more conservative value of Wikstrom et al [37] to constrain the origin of the angiosperms (150 Myr versus 179 Myr) in addition to several other calibrations points based on detailed family-specific dating analyses (see methods). Approaches whereby DNA sequence data is generated for the whole sampling would possibly provide better resolution at shallower nodes and hence better address more recent signals [47]. However the resolution of our tree is good for ancient lineages but poor for recent ones. As a consequence this pattern should be interpreted with care.

Our results indicate that Annonaceae species are more related within transects than species taken from different transects. This result could be real and not just an artifact of phylogenetic resolution as the phylogenetic tree for Annonaceae is well resolved [48,49]. Clustering of Annonaceae was also found in another study of African phylogenetic structure in Equatorial Guinea [20] in which phylogenetic differentiation was shown to be correlated with elevation. They also indicated that the number of magnoliid (which includes Annonaceae) species per plot was correlated with altitude. Interestingly, the DFR has very little elevation variability [27] and thus the significant phylogenetic differentiation detected in Annonaceae of the Dja would have to result from a different process than altitudinal gradients. Moreover, in contrast to South America, lowland Annonaceae are more or as diverse at mid latitudinal as indicated in a survey of an elevation gradient in Mont Cameroon [50]. On the contrary, we observed an opposite pattern for Apocynaceae since species among transects appear more related than within indicating an overdispersion of phylogenetic pattern. This result might be linked to the fact that the species Tabernanthea crassa (Apocynaceae) is the most abundant species inventoried in the reserve and strongly present in transects [51]. The rest the Apocynaceae species are not well represented (1–64 individuals/species) and less well represented across all transects. In both cases, Annonaceae and Apocynaceae have a large number of liana species [52,53] that have not been inventoried in this study and thus more detailed sampling and tests should be carried out before a link to any evolutionary pattern can be done. However, the more in depth analyses looking at the species rich clade as the Annonaceae indicates that some phylogenetic sorting is occurring among transects and can be detected when more data is available.

4. Phylogenetic diversity and conservation

A classical measure of phylogenetic diversity (signal) is Faith’s PD [12] which measures the total phylogenetic branch length (i.e. amount) of evolutionary history in the studied community. This measure is equivalent in our work to $D_{PS}$ which also does not account for species abundance. However, $D_{PS}$ is a measure of phylogenetic distinctness, but has the advantage not to be influenced by species richness [20].

Recent literature has debated the interest of adding phylogenetic diversity evaluation in conservation planning [17,54,55]. One general agreement in favor of taking phylogenetic diversity into account is to conserve all components of biodiversity including evolutionary information, and that the explicit consideration of biodiversity as comprising evolving and related lineages would add power and robustness to measures of biodiversity for conservation [17]. Specifically, adding phylogenetic estimation in conservation strategy would result in maximization of the set of species to be conserved [55]. According to our results, most

<table>
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<td>Whole data-transect</td>
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<td>Model 1s</td>
<td>$H_0: \Delta T = 0$  $p = 0.348$ NS</td>
<td>No phylogenetic turnover</td>
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<td>Whole data-habitat</td>
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<td>Taxonomic differentiation between habitats: reflect a filtering habitat effect on species</td>
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<td>$H_0: \Delta T = 0$  $p = 0.05$ * (H1: $\Delta T &gt; 0$)</td>
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<td>No phylogenetic turnover</td>
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<tr>
<td></td>
<td></td>
<td>$H_0: \Pi = 0$       $p = 0.064$ NS</td>
<td>No phylogenetic turnover</td>
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Details of model permutation are given in Table 1. P-values are given after 999 permutations of individuals or species in model of permutation. NS: non significant. Stars indicate the level of the significance. doi:10.1371/journal.pone.0098920.0004
diversity occurred within transect or habitat, the between contribution being always less than or equal to 1.1% (whole dataset-habitat).

Conclusions

The preservation of tropical rainforest is an ethical, political and practical concern and biodiversity assessment should be a major focus in nature preservation programs [56]. Indeed, faced with high anthropic pressure in tropical forest, the number and extent of protected areas have increased across the tropics [57]. The objective of such protected areas is to conserve a sufficient sample of the world’s biodiversity.

Few conservation policies consider phylogenetic diversity as an important component probably because the added value of phylogenetic diversity for nature conservation remains unclear [55] due to a lack of consensus between various measures and a difficulty to interpret the results in terms of conservation perspectives [54,55]. Here, we detected a random phylogenetic pattern between transects at the scale of the Dja Faunal Reserve, possibly because of a common history and weak environmental variation. We also showed that geographic distance encompassed species turnover. In addition, our phylogenetic based analysis added new results to the previous study of Hardy and Sonké [27] using the same dataset, by detecting a weak but significant phylogenetic turnover signal among habitats reflecting a filtering effect of the habitat. Our results can contribute to the conservation of the park by providing insights into the processes driving community assembly. Notably, the prevalence of patterns compatible with dispersal assembly highlights the need to conservation schemes that allow for sufficiently large conservation areas. Future studies should investigate more plots to be based on a hierarchical sampling plan considering spatial variation within transects in order to better interpret the phylogenetic structure.

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Author Contributions

Conceived and designed the experiments: BS. Performed the experiments: BS. Analyzed the data: TC SM OH. Contributed reagents/materials/analysis tools: FM. Wrote the paper: SM TC PC FM OH.

References

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57. Jenkins CN, Joppa L (2009) Expansion of the global terrestrial protected area


51. Sonke B, Couvreur TLP (2014) Tree diversity of the Dja Faunal Reserve, South

50. Bele MY, Focho DA (2011) Inventory and distribution of the Annonaceae along


43. Letunic I, Bork P (2011) Interactive Tree of Life v2: online annotation and

42. Letunic I, Bork P (2007) Interactive Tree Of Life (iTOL): an online tool for

41. Sonke B, Couvreur TLP (2014) Tree diversity of the Dja Faunal Reserve, South


38. Hardy OJ (2010) SPACoDi 0.10: a program for spatial & phylogenetic analysis


33. Letunic I, Bork P (2011) Interactive Tree of Life v2: online annotation and

32. Letunic I, Bork P (2007) Interactive Tree Of Life (iTOL): an online tool for

31. Sonke B, Couvreur TLP (2014) Tree diversity of the Dja Faunal Reserve, South


29. Hardy OJ (2010) SPACoDi 0.10: a program for spatial & phylogenetic analysis


