Effect of temperature on the in vitro multiplication of seven Radopholus similis isolates from different banana producing zones of the world

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Summary – A study was carried out to compare the multiplication rate as a function of temperature of several isolates of the burrowing nematode Radopholus similis cultured monoxenically on carrot discs. These isolates were collected on banana roots in seven production areas of the world (Costa Rica, Martinique, Guadeloupe, Guinea, Ivory Coast, Sri Lanka, and Queensland). The study was divided into two parts. In the first part, each isolate was studied at four temperatures regimes: 21, 24, 27 and 30 °C. In the second part, the seven isolates were studied concurrently at two temperatures: 30 and 33 °C. All the isolates showed a similar pattern of behaviour in relation to the temperature. Multiplication rate was very low at 21 °C, increased rapidly in relation to the temperature to reach a maximum level at 30 °C, falling sharply at 33 °C. On the other hand, the different isolates had very different intrinsic multiplication rates at all temperatures. The isolates from Ivory Coast, Costa Rica and Guinea showed the highest multiplication rate, whereas the isolates from Martinique and Queensland had the lowest. These results confirm the wide range of intraspecific biological diversity of Radopholus similis.


Key-words : banana, biological diversity, multiplication rate, Radopholus similis, temperature.

Abundant information is available on the biological and physiological diversity of nematodes (Sturhan, 1971; Dropkin, 1988). The knowledge of the intraspecific diversity, especially in relation to pathogenicity, is essential for research concerning plant-nematode interactions leading towards successful plant breeding for resistance and effective integrated nematode control through cultural practices, such as crop rotation (Wallace, 1963; Sturhan, 1971; Dropkin, 1988).

The biological diversity of Radopholus similis was first studied on isolates from Central America and the Caribbean (Edwards & Wehunt, 1971; Pinochet, 1979, 1988; Tarté et al., 1981; Rivas & Román, 1985). These studies revealed a large diversity in pathogenicity to banana plants, in direct relation to the multiplication rate. Recently, it has been shown that the diversity of this species extends to other areas of the world (Sarah et al., 1992, 1993). R. similis diversity is apparently due to divergent evolution under different environmental and host conditions. Of the environmental conditions, temperature might play an important role.

Studies on influence of temperature on sub-specific biological diversity performed on nematodes species are relatively rare (Fagbenle et al., 1989; Jaehn & Lordello, 1990; Rutherford et al., 1992). To our knowledge, no studies have been carried out to date on R. similis.

Materials and methods

The study was performed with seven R. similis isolates from different banana growing areas of the world. Five of these, Martinique (Morne Rouge), Guadeloupe (Neufchâteau), Costa Rica (Talamanca), Ivory Coast (Angouédédou) and Sri Lanka (Hantane), had been previously studied in relation to their pathogenicity on banana plants (Sarah et al., 1993). Two further isolates, one from Guinea (Balikouré) and the other from Aus-
ultural (Mac Kay State, Queensland), were included in this study. All seven isolates were extracted from roots of banana plants (Musa AAA) and maintained in monoxenic culture on carrot discs (O’Bannon & Taylor, 1968) at 27 °C. The use of carrot discs to study the multiplication rates of nematodes is a commonly used technique and very useful (Verdejo-Lucas & Piñochet, 1992; Piñochet et al., 1994, 1995).

In 100 ml flasks were placed 5 ml of 1 % agar solution and four or five carrot discs; 1 cm in diameter (total weight: 5-7 g). An average of 70 nematodes (90 % females, 10 % males and juveniles) were placed on the carrot discs in each flask.

Two separate studies were carried out. In the first one, each isolate was studied at four temperature regimes: 21, 24, 27 and 30 °C. In the second experiment, the seven isolates were studied concurrently at two temperatures: 30 and 33 °C. Population levels were evaluated at 30, 40 and 50 days after inoculation in the first study and at 20, 30 and 40 days in the second.

To collect the nematodes for counting, the content of the flask was emptied into a blender (Braun MX 32) for maceration (three times 5 s at full speed). The flask was washed with water to remove any nematodes adhering to the wall. This water was added to the macerated contents of the flask on a sieve column of different pore sizes (250, 80, 50 and 32 μm). The nematodes were then recovered from the last three sieves of the column and separated from the tissue debris using the centrifugation/floation technique (Coolen & D’Herde, 1972).

In both studies, the flasks were arranged in a totally-randomized design with five replicates. Nematode counts were transformed to $\log_{10}(x + 1)$ for analysis of variance. Means were compared using the Newman-Keuls test ($P \geq 0.05$).

**Results**

In the first study (Fig. 1), nematode multiplication at 21 °C was very low regardless of the isolate and date considered. There were no significant differences of nematode number at 30 days. For three isolates (Ivory Coast, Martinique and Guinea), an average of 100-140 nematodes were recovered in each flask 50 days after inoculation. Population build-up of the Sri Lanka and the Guadeloupe isolates at 50 days was significantly higher than the previously mentioned isolates, but remained fairly low (220 and 340 nematodes recovered, respectively). At this same temperature, the highest reproduction rate occurred with the Queensland and the Costa Rica isolates, reaching 760 and 1320 nematodes per flask, respectively.

At higher temperatures (24, 27 and 30 °C) nematode multiplication was considerably more rapid and the rate increases in relation to the temperature. At 30 days, nematode number were still relatively low: from 700 to 4900 per flask, depending on the temperature and the isolate. There were no significant differences among isolates whatever the temperature at this date. At 40 days, the maximum of recovered nematode number were around 10,000 at 24 °C, 20,000 at 27 °C and 50,000 at 30 °C. At this date, differences among isolates appeared more clearly and more consistently. At 24 °C, the isolates from Guinea, Ivory Coast and Costa Rica significantly reached higher population levels than the others. At 27 °C the same three isolates, plus the Guadeloupe isolate, reproduced significantly more; the Queensland and Martinique isolates had the lowest nematode number, and the Sri Lanka isolate showed an intermediate level of reproduction. At 30 °C the population ranking was the same as at 27 °C but with more apparent separation between the different isolates. At 50 days, the population build-up made the separation between isolates clearer increasing the differences of nematode number among isolates consistently at any temperature. However, the Costa Rica isolate at 24 °C and those from Ivory Coast, Guinea and Costa Rica at 30 °C multiplied more slowly than the other isolates between 40 and 50 days. Degradation of carrot tissue occurred at 30 °C in most of the flasks where those three isolates were cultured. This degradation probably limited available substrate for population build-up and therefore the final nematode numbers should have been negatively affected.

In the second study (Table 1), there was almost no population build-up at 33 °C since, whichever the date of observation, levels were about the same as those at

<table>
<thead>
<tr>
<th>Isolates</th>
<th>30 °C</th>
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<th>30 °C</th>
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</tr>
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<tbody>
<tr>
<td>Ivory Coast</td>
<td>1200 a</td>
<td>120 a</td>
<td>5040 a</td>
<td>64 ab</td>
<td>46400 a</td>
<td>64 a</td>
</tr>
<tr>
<td>Costa Rica</td>
<td>1250 a</td>
<td>96 a</td>
<td>3480 b</td>
<td>128 a</td>
<td>41200 a</td>
<td>112 a</td>
</tr>
<tr>
<td>Martinique</td>
<td>640 b</td>
<td>96 a</td>
<td>1290 d</td>
<td>100 a</td>
<td>10000 c</td>
<td>80 a</td>
</tr>
</tbody>
</table>

Data are means of five replicates. Actual data are presented, but data were transformed to $\log_{10}(x + 1)$ for analysis. Means in the same column followed by the same letter are not different ($P \geq 0.05$) according to Newman and Keuls test.
Multiplication of *Radopholus similis* isolates

Nematodes per flask

Days after inoculation

Fig. 1. Population build-up of seven *Radopholus similis* isolates at different temperatures on *carrot* discs. A: 21 °C; B: 24 °C; C: 27 °C; D: 30 °C. (■: Ivory Coast; □: Guinea; ●: Costa Rica; ▲: Guadeloupe; ▼: Martinique; ●: Sri Lanka; ○: Queensland). Means of five replicates. Actual data are presented, but data were transformed to log_{10} (x + 1) for analysis. Means at the same date with the same letter are not different according to Newman and Keuls test (P ≤ 0.05).

inoculation date. At 30 °C, the multiplication rates were very high and similar to those observed in the first study 30 and 40 days after inoculation whichever isolate was considered. This shows that the results obtained here are reliable and reproducible. Therefore, the results of the two studies may be combined to draw graphs of population levels observed at 40 days as a function of temperature, for the different isolates (Fig. 2). At 40 days no carrot tissue degradation was observed and the data obtained reflect the reproduction rate of each isolate. The
graphs are similar in appearance for all isolates, showing 40-day nematode number increasing with temperature to a maximum at 30 °C and falling back sharply at 33 °C.

The pooling of data from the two studies allowed good separation, by Newman and Keuls test, of the levels observed at 30 °C 40 days after inoculation (Fig. 2). The Ivory Coast and Costa Rica isolates reached the highest population levels. The Guinea isolate showed population levels lower than those of the Ivory Coast isolate, but not significantly different from those of the Costa Rica isolate. These were followed in order of decreasing levels by the Guadeloupe isolate, then the Sri Lanka isolate. The Queensland and Martinique isolates presented similar population levels, which were the lowest observed here.

Discussion

The general pattern of population build-up as influenced by temperature was roughly uniform among the different isolates of *R. similis* studied here. All the isolates reached highest numbers at 30 °C. The maximum temperature limit for multiplication was probably not far below 21 °C and the maximum temperature limit was close to 33 °C. This maximum temperature was not a lethal temperature, since the nematodes recovered were alive, but there was little or no reproduction at this temperature whatever the considered isolate. One should make the hypothesis that this homogeneity among isolates should have been induced by the common breeding temperature at which all the isolates were maintained for a period ranging from 3 years (Martinique, Ivory Coast) to 1 year (Queensland). Thermal acclimatization is widespread among invertebrates, and thermal preference for moves of some nematode species has been proved to be influenced by temperature of storage (Croll, 1967; Robinson, 1989; Robinson & Heald, 1989, 1993). However, the observed optimal temperature (30 °C) was higher than the breeding temperature (27 °C), and there was a good homogeneity of relative multiplication rate as a function of temperature for all isolates. Therefore this homogeneity would appear to be a character inherent to the species whose biological unity seems to have resisted modifications despite divergent evolution under different environmental conditions.

Although the general pattern of final population as a function of temperature is similar for all isolates, the intrinsic reproduction rate was highly variable depending on the geographical origin. There was a fairly low isolate temperature interaction between 24 and 30 °C which are the temperatures allowing high rates of multiplication. In this temperature interval, isolates ranked almost in the same order in terms of population build-up, variation being generally non-significant. The only exceptions were the number of nematodes of the Costa Rica isolate at 24 °C 50 days after inoculation and the reduction of multiplication rate of the isolates from Ivory Coast, Guinea and Costa Rica between 40 and 50 days related with degradation of carrot tissue at 30 °C. In any other case isolate ranking agrees with the relative degree of pathogenicity in banana plants observed in previous experiments (Sarah et al., 1993). The five isolates common to these previous experiments (Ivory Coast, Costa Rica, Guadeloupe, Sri Lanka and Martinique) were ranked in the same order considering decreasing multiplication rate which is strongly associated with pathogenicity. However, there was one exception. The Martinique isolate showed a higher reproduction rate than the Sri Lanka isolate in banana plants roots at 27 °C (Sarah et al., 1993). This correlation between multiplication rate *in vitro* on carrot discs and pathogenicity to banana roots indicates that the pathogenic diversity of *R. similis* is associated with intrinsic factors that point towards important genetic differences between the different isolates.

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References


