Temporal study of natural populations of Heterorhabditid and Steinernematid nematodes in horticultural crop soils

Fernando GARCÍA DEL PINO and Alejandro PALOMO

Departamento de Biología Animal, Facultad de Ciencias, Universidad Autónoma de Barcelona, 08193 Bellaterra, Barcelona, Spain,

Accepted for publication 13 December 1996.

Summary – The dynamics of natural populations of entomopathogenic nematodes in horticultural crop soils were studied via monthly sampling of eight sites in Catalonia (NE Spain) over 14 months. Entomopathogenic nematodes were found at six of the eight sites and they continued to be detected regardless of the agricultural practices carried out at the sites. During the study these sites were ploughed, which destroyed the natural habitat of the nematodes, then left fallow for several months, but this did not have any significant effect on the presence of the nematodes. However, some seasonal fluctuations were observed with lower populations during the hotter summer months. This seasonality also appeared to affect the vertical distribution: the nematodes migrated to deeper layers during summer, presumably to avoid the damaging effects of temperature and lack of humidity. The results of this study show that natural populations of entomopathogenic nematodes are capable of persisting and surviving for a long time in the soil, by adapting to the fluctuating and adverse conditions of their habitat.

Résumé – Etude temporelle des populations naturelles de nématodes Heterorhabditides et Steinernematides dans les sols horticoles - La dynamique des populations de nématodes entomopathogènes dans des sols horticoles a été étudiée par des prélèvements mensuels en huit sites de Catalogne (nord-est de l'Espagne) pendant 14 mois. Ces nématodes entomopathogènes ont été détectés dans six des huit sites et ont continué à l'être quels qu'aient été les traitements agricoles pratiqués sur ces sites. Pendant cette étude, les sites ont été labourés, détruisant ainsi l'habitat naturel des nématodes, puis laissés en jachère pendant plusieurs mois, sans que la présence des nématodes n'en paraisse affectée. Cependant, une influence saisonnière peut être observée, la présence des nématodes étant plus faible pendant les mois d'été où la température est élevée. Cette influence saisonnière apparaît également affecter la répartition verticale des nématodes qui migrent vers les couches plus profondes du sol, vraisemblablement pour éviter les effets néfastes de la température et du manque d'humidité. Les résultats de cette étude montrent que les populations naturelles de nématodes entomophathogènes sont capables de persister et de survivre dans le sol pendant de longues périodes en s'adaptant aux conditions fluctuantes et adverses de leur habitat naturel.

Key-words: ecology, entomopathogenic nematodes, Heterorhabditidae, persistence, Steinernematidae, vertical distribution.

Entomopathogenic nematodes of the families Steinernematidae and Heterorhabditidae are obligate and lethal parasites of insects. They possess many of the attributes of an ideal biological control agent as they have a wide host spectrum, are environmentally safe, can be produced in large-scale bioreactors, are easily applied, and are compatible with most chemical pesticides. These organisms are used as biological control agents of different insect pests in diverse climatic conditions. By using the Galleria trap method (Bedding & Akhurst, 1975), it is possible to estimate the occurrence of entomopathogenic nematode species in the soil, and this technique has been used in many parts of the world (Poinar, 1990). García del Pino and Palomo (1996) observed a wide distribution of these nematodes in Catalonia (Spain). However, none of these studies examined the population dynamics of the nematodes. A few temporal studies of natural populations of heterorhabditid and steinernematid

nematodes (Hominick & Briscoe, 1990), and of their dynamics in cultivated soils (Glazer *et al*, 1996; Sturhan, 1996) have been published. As stated by Hominick and Reid (1990), the study of population dynamics of entomopathogenic nematodes is fundamental to understanding their persistence, distribution, and effect on insect populations and for the development of predictive models for control programmes.

In this study, we investigated the dynamics of natural populations of steinernematid and heterorhabditid nematodes through a temporal survey lasting 30 months with monthly sampling during a 14-month period at eight horticultural crop sites. The presence of these nematodes and their vertical distribution were analysed in relation to the environmental and agricultural conditions throughout the four seasons in which they were studied.

Materials and methods

The presence of entomopathogenic nematodes was monitored in eight sites (P1 to P8), each with an area of approximately 50 m², situated in horticultural fields in Baix Llobregat (Catalonia, Spain). The samples were taken monthly from September 1990 to October 1991, and on separate occasions during August 1992 and January 1993.

At each site, each sample was composed of three sub-samples taken at 1 to 3 m intervals. The status of the crop growing at that time was noted. Each subsample consisted of 1 kg of soil taken from a 1 m² area, at a depth of 5-20 cm. The three sub-samples were pooled and placed in a plastic bag for transport to the laboratory, where the nematodes were extracted through a modification of the "Galleria-trap" method (Bedding & Akhurst, 1975). The sample was divided into three equal fractions, approximately 1 kg each, and these were placed in plastic containers of 1 dm³ capacity with six late instar Galleria mellonella larvae at the bottom of each container. When necessary, the samples were moistened to allow mobility of nematodes. The containers were kept at a temperature of 23 ± 2 °C and sealed to prevent desiccation of the sample. After 7 days, the Galleria larvae were recovered and partially dissected to test for the presence of entomopathogenic nematodes. To quantify the presence of nematodes at every site, the three fractions of each sample containing nematodes were recorded.

The climatic data was recorded throughout the survey, including minimum, maximum, and mean air temperatures and rainfall. The soil at the eight sites was analyzed for granulometry and fertility. During the survey, no nematicides were applied to the soil, and it was not disinfected. However, it was impossible

to prevent the application of other chemical pesticides (insecticides, fungicides, etc.).

To study the vertical distribution of natural populations of entomopathogenic nematodes, depth sampling was carried out by boring during the months of April 1991, August 1991, October 1991, and January 1993, only in those fields where nematodes had been detected during the monthly samplings. Depth sampling consisted of ten soil samples, each 3 cm in diameter and 30 cm deep, extracted by using a gouge auger. Each core sample was divided into 5 cm long sections. Each section was placed in a Petri dish 9 cm in diameter and labelled for transport to the laboratory. The "Galleria-trap" method was used by adding four late instar G. mellonella larvae in each dish for extracting the nematodes. The dishes were kept at 23 ± 2 °C and covered with a plastic bag to prevent desiccation. After 7 days, the larvae were recovered and partially dissected to detect the presence of nematodes, and the number of larvae parasitized by nematodes was recorded at each depth.

Results

The granulometry analysis indicated that the sampled soils were loam or silt loam. The fertility analysis revealed that their salinity and pH range were adequate for nematodes survival and that they had a high organic matter content (see Table 1).

The climatic conditions that we recorded -viz., minimum, maximum and mean monthly air temperatures and monthly precipitation during the 14 months of the study- are given in Fig 1. A clear seasonality was present with a minimum winter temperature of 1.2 °C, but the average monthly minimum temperature in winter did not drop below 5 °C and the monthy

Table 1. Granulometry and fertility analyses of the soils sampled.

	Granulometry analysis					Fertility analysis				
	Fine sand (0.05 - 0.5 mm)	Coarse sand (0.5 - 2 mm)	Silt (0.002 - 0.05 mm)	Clay (<0.002 mm)	Classif. USDA*	E. C. (dS/m)	pН	O. M. (% p/p)	P (mg./kg.)	K (mg./kg.)
	13.61	14.99	60.95	24.06	L-S	0.52	8.01	3.84	147.70	907.51
P2	15.04	16.61	60.87	22.52	L-S	0.42	8.10	3.41	106.12	725.45
P3	34.65	36	51.21	12.79	L-S	0.60	8.16	2.43	50.97	624.14
P4	29.78	37.97	42.36	19.67	L	0.51	7.98	4.58	137.94	864.65
P5	31.34	33.54	48.38	19.08	L	0.40	7.99	3.16	49.49	299.46
P6	30.32	34.35	44.49	21.16	L	0.77	7.51	4.10	101.34	487.37
P7	9.25	11.14	59.97	28.89	L-C-S	0.77	8.16	3.25	55.72	386.57
P8	12.93	13.23	61.57	25.20	L-S	0.60	8.18	2.44	48.80	258.71

^{*} L : Loam, S : Silt, C : Clay

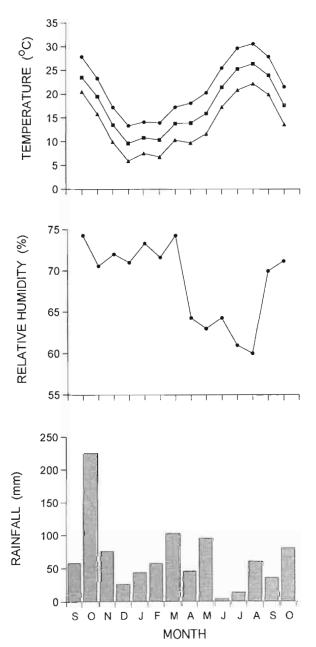


Fig. 1. Climatic conditions in the sites sampled. Temperature reported as monthly maximum (lacktriangle), minimum (lacktriangle), and mean (lacktriangle) temperatures.

mean temperature in winter was approximately 10 °C. During the summer, the maximum temperatures were up to 33 °C, although the average monthly maximum did not exceed 30.5 °C. The characteristic seasonality of the Mediterranean region was also evident in the recorded rainfall, with minimum precipitation in June and July (3.6 and 13.3 mm, respectively).

In two of the eight sites sampled (P7 and P8) no entomopathogenic nematodes were found during the entire period of the study. These two sites were not included in the subsequent calculations or figures.

In five of the six sites with entomopathogenic nematodes, nematodes belonging to the family Heterorhabditidae, i.e., Heterorhabditis bacteriophora, Poinar, 1976, were observed, while only site P3 contained nematodes belonging to the family Steinernematidae, i.e., Steinernema feltiae (Filipjev, 1934). During the survey, no changes were observed in the species originally detected at any of the sites analysed.

The graphic representation of the number of sites and the number of samples containing entomopathogenic nematodes during the 14 months of the survey (Fig. 2) reveals some seasonal fluctuations in the occurrence of nematodes throughout the year, which could be related with the seasonality of the environmental parameters described above (Fig. 1).

Observing the results for each of the sites during the study, we see that the presence of the entomopathogenic nematodes is variable (Fig. 3). In only two sites (P3 and P5) were the nematodes detected regularly, regardless of the agricultural practices being carried out on the sites at the time of the successive samplings. At site P3, at the time when the natural population of the nematode S. feltiae was detected, a crop of tomato plants harboring a potential supply of insect hosts was under cultivation. After two months (September and October) the crop was dug out and the site was left in this condition for two more months (November and December). After this, the field was ploughed, which altered the structure of the soil and destroyed the habitat that had been protecting the nematodes, thus exposing them to the effects of harmful agents (solar radiation, desiccation, etc.). The field remained fallow for 3 months (February to April), which means that the land was not watered for more than 6 months. As there was no crop during this period, the insect supply probably was minimal. After this, the land was ploughed again and then left fallow for another 3 months (June to August). Throughout this time, every monthly sample yielded nematodes until July and August, when none were detected. In September the land was sowed and the entomopathogenic nematodes reappeared and they were also detected in October. On the next two separate samplings (August 1992 and January 1993), no nematodes were detected in August 1992, but they reappeared in January 1993, 15 months after their last detection (October 1991). A similar trend was observed in site P5 (Fig. 3) with natural populations of H. bacteriophora.

The vertical distribution of entomopathogenic nematodes in the soil is shown in Fig. 4. The percentage of larvae parasitized by nematodes in relation to depth

Vol. 20, n° 5 - 1997 475

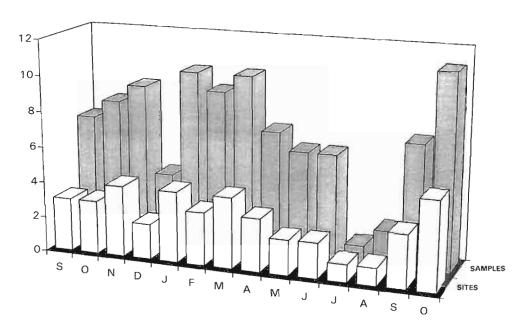


Fig. 2. Number of sites and number of samples containing entomopathogenic nematodes during the 14 months of the survey.

was calculated from all the samples taken during the four seasons of the study in those fields where nematodes were present. We assumed that the number of infected larvae found by sampling was related to the actual number of nematodes that were present in the soil (Mráček, 1982). The results relating to the two species of nematodes found (H. bacteriophora and S. feltiae) are presented as two separate graphs, as their dispersive behaviour and persistence are different, which causes different vertical distributions. Although for both species the greatest numbers of nematodes were detected between 5 and 10 cm depth, in the aggregate they have a significantly different vertical distribution ($\chi^2 = 13.10$, d.f. = 5, P = 0.0224). H. bacteriophora is present in greater numbers at greater depths and in smaller numbers (6% of the Galleria larvae parasitized) near the surface (0-5 cm depth) (Fig. 4). On the contrary, S. feltiae is mostly present in the upper zones and there are less nematodes at greater depths: 9% of parasitized larvae at 20-25 cm and 5% at 25-30 cm depth (Fig. 4). Fig. 5 shows the vertical distribution of the entomopathogenic nematodes, measured as the percentage of Galleria larvae parasitized by nematodes at each depth, for every season and for every field studied. Separate statistical analyses were made for sites with entomopathogenic nematodes from the family Heterorhabditidae and for site P3 with the steinernematid nematode S. feltiae. In both cases, there was a statistical difference in the

vertical distributions during the different seasons $(\chi^2 = 154.8, d.f. = 15, P \le 0.0001 \text{ for } H. bacteriophora$ and $\chi^2 = 342.4, d.f. = 15, P \le 0.0001 \text{ for } S. feltiae)$.

Discussion

There are very few studies on the temporal development of natural populations of entomopathogenic nematodes. In the present study, in some sites the nematodes were regularly detected in the soil throughout the entire period of the study (with the exception of the summer months), while in other sites their occurrence was more variable. Besides seasonal effects, this variability can be attributed to various intrinsic and extrinsic factors, and it may also depend on the abundance of the nematodes. The most significant extrinsic factors were abiotic factors, such as temperature, humidity, structure, and organochemical characteristics of the soil (Kaya, 1990), and biotic factors, such as cultivation cycle of the host and presence of antagonists (Poinar & Jansson, 1986 a,b; Epsky et al., 1988; Kim et al., 1988; Gilmore & Potter, 1993). Intrinsic factors may also play a major role in the observed variability in the occurrence of entomopathogenic nematodes. Various authors have shown that infective juveniles of the nematodes go through non-infective stages in order to stagger their infectiousness as a survival strategy (Bednarek &

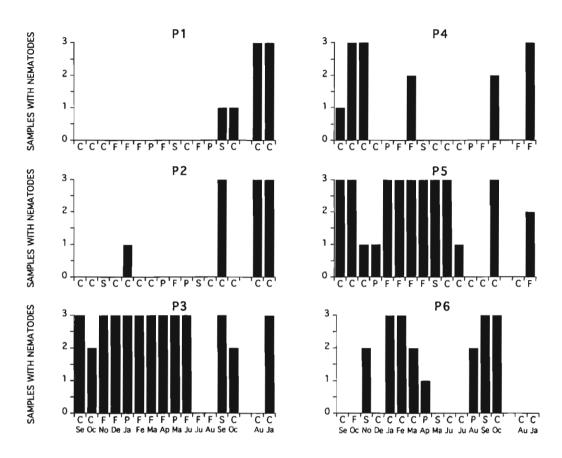


Fig. 3. Presence of nematodes in each of the sites during the study, and current agricultural practices at the sites: (C) crop present, (F) fallow, without crop, (P) ploughed, (S) sowed. (n = 3 per sampling date).

Nowicki, 1991; Fan & Hominick, 1991; Kaya & Gaugler, 1993; Bohan & Hominick, 1996 *a,b*).

The seasonal fluctuations observed in the populations of entomopathogenic nematodes in the Mediterranean region consist in a decrease of the population levels during the summer months, which is largely due to the higher temperatures. Hominick and Briscoe (1990) found no evidence of a seasonal component in the occurrence of the nematode S. bibionis (= S. feltiae), in a survey carried out over 28 months in Southern England, where the temperatures during July and August vary between 14.5 and 22 °C with a mean temperature of 17 °C. By contrast, in our study the mean temperature was 26.5 °C and the maximum temperature was 30.5 °C during the same period. This absence of seasonality in the occurrence of nematodes in the northern European countries has also been observed by Sturhan (1996) in Germany, who found the highest number of infective juveniles in

June and August, which is quite the opposite of our results. These various observations suggest that the presence or absence of a seasonal fluctuation in the population levels of entomopathogenic nematodes, and the nature of such a seasonal fluctuation if it exists, depend on the climatic conditions of the locality where the nematodes are present. As various authors have already observed (Burrman & Pye, 1980 a,b; Molyneux, 1985; Kung & Gaugler, 1991), temperature has an effect on the speed of depletion of nutritive reserves, therefore on the persistence and survival of infective juveniles in the soil. Moreover, temperature affects other environmental factors such as humidity, and biotic factors such as presence and population levels of predators and pathogens of the infective juveniles, which leads to a lesser survival of these nematodes in the soil.

In agricultural soils, cultural practices periodically destroy the soil structure and the plant cover. The

Vol. 20, n° 5 - 1997

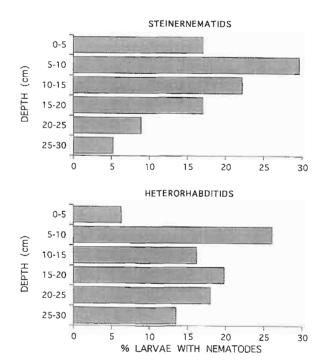


Fig. 4. Presence of steinernematid and heterorhabditid nematodes at six soil depths in all the samples taken during the whole study.

presence of a crop in the field can affect the presence of entomopathogenic nematodes in the soil, either directly by the action of the roots, that play an important role in the formation of the soil structure by giving it greater humidity, porosity and aeration, or indirectly because of regular watering that prevents excessive desiccation of the soil, regular supply of insects that can become hosts, etc. Several authors (Blackshaw, 1988; Vänninem et al., 1989) have suggested that these agricultural practices can have an effect on the presence of entomopathogenic nematodes. However, Sturhan (1996) and our results do not seem to support this theory. The analysis of our results (e.g., P3, Fig. 3) leads us to conclude that agricultural practices apparently have no effect on the presence of entomopathogenic nematodes, as these nematodes have been regularly observed both in sites under permanent cultivation and in sites without crops for a long period. This suggests the existence of an adaptability that allows these natural populations of entomopathogenic nematodes to survive under the fluctuating and adverse conditions generated by agricultural practices. Such phenomena as diapause and quiescence (Ishibashi & Kondo, 1986) and anhydrobiosis (Womersley, 1990, 1993) can explain the persistence of the entomopathogenic nematodes in these agricultural soils, as well as their reappearance after the summer period.

Moyle and Kaya (1981) have shown that certain species of Steinernema prefer to search for hosts close to the surface, and Georgis and Poinar (1983) and later Choo et al. (1989) have established that species of Heterorhabditis are more suited to searching for hosts at a greater depth. The present data confirms these observations in that S. feltiae is generally present in greater numbers in the upper layers (0-20 cm) of the section of soil analysed, while H. bacteriophora is more abundant in the lower layers (as deep as 30 cm). Villani and Wright (1990) observed that many soilinhabiting animals undergo vertical migrations to avoid the damaging effects of temperature or lack of humidity. Although there are many laboratory studies of vertical distribution of entomopathogenic nematodes and of their ability to find a host at different depths (Schroder & Beavers, 1987; Gaugler et al., 1989 a,b; Choo et al., 1989; Westerman & Godthelp, 1990; Nguyen & Smart, 1990; Shapiro et al., 1993), the importance of this avoidance behaviour for the survival of entomopathogenic nematodes has not yet been studied. Our results do not show any clear trend, perhaps because of the interference of the agricultural practices carried out in these soils. However, it can be observed that in autumn, i.e., after the nematode populations had been subjected to the harmful effects of high summer temperatures at the soil surface, a greater number of nematodes (with steinernematid and heterorhabditid nematodes in equal numbers) were present in the deepest layers of the ground. In winter, when the surface temperature presented no danger to their survival, there was a greater number of nematodes in the surface layers of the soil, as Glazer et al. (1996) have also observed. These results could indicate that entomopathogenic nematodes display an avoidance behaviour from adverse conditions through migration to greater depths.

To conclude, the present results confirm that natural populations of entomopathogenic nematodes are capable of persisting and surviving for a long period in the soil. These nematodes are better adapted to the fluctuating and adverse conditions characteristic of their environment, and they survive in greater numbers, than populations of entomopathogenic nematodes artificially introduced. This issue should be considered when selecting the entomopathogenic nematodes the most suitable for use in biological pest control programmes.

Acknowledgments

The authors thank Dr. Javier Retana for his technical support in the preparation of the figures, and Suzzan Bush for the English correction of the manuscript.

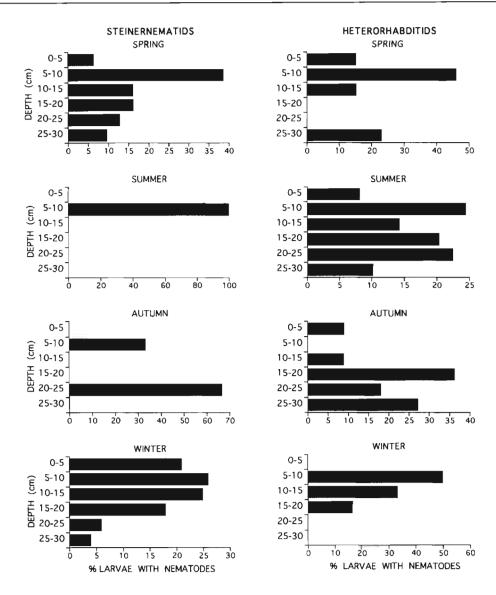


Fig. 5. Presence of steinernematid and heterorhabditid nematodes at six soil depths during the various seasons studied. (ten samples per season).

References

BEDDING, R. A. & AKHURST, R. J. (1975). A simple technique for the detection of insect parasitic rhabditid nematodes in soil. *Nematologica*, 21: 109-110.

BEDNAREK, A. & NOWICKI, T. (1991). New estimation method for the density of entomogenous nematodes (Rhabditida: Steinernematidae) in the soil. *Revue Nématol.*, 14: 638-639.

BLACKSHAW, R. P. (1988). A survey of insect parasitic nematodes in Northern Ireland. *Ann. appl. Biol.*, 113: 561-565.

BOHAN, D. A. & HOMINICK, W. H. (1996 a). Examination of the Steinernema feltiae (Site 76 strain) infection intera-

tion with the Galleria mellonella host, using an infection model. Parasitology, 111: 617-625.

BOHAN, D. A. & HOMINICK, W. H. (1996 b). Investigations on the presence of an infectious proportion among populations of *Steinernema feltiae* (Site 76 strain) infective stages. *Parasitology*, 112: 113-118.

BURMAN, M. & PYE, A. E.. (1980 a). Neoaplectana carpocapsae: movements of nematode populations on a thermal gradient. Exp. Parasit., 49: 258-265.

BURMAN, M. & PYE, A. E.. (1980 b). Neoaplectana carpocapsae: respiration of infective juveniles. Nematologica, 26: 214-219.

CHOO, H. Y., KAYA, H. K., BURLANDO, T. M. & GAU-GLER, R. (1989). Entomopathogenic nematodes: host-

Vol. 20, n° 5 - 1997

- finding ability in the presence of plant roots. *Envir. Ent.*, 18: 1136-1140.
- EPSKY, N. D., WALTER, D. E. & CAPINERA, J. L. (1988). Potential role of nematophagous microarthropods as biotic mortality factors of entomogenous nematodes (Rhabditida: Steinernematidae, Heterorhabditidae). J. econ. Ent., 81: 821-825.
- FAN, X. & HOMINICK, W. M. (1991). Effects of low storage temperature on survival and infectivity of two *Steinernema* species (Nematoda: Steinernematidae). *Revue Nématol.*, 14: 407-412.
- GARCÍA DEL PINO, F. & PALOMO, A. (1996). Natural occurrence of entomopathogenic nematodes (Rhabditida: Steinernematidae and Heterorhabditidae) in Spanish soils. J. Invert. Pathol., 68: 84-90.
- GAUGLER, R., CAMPBELL, J. F. & MCGUIRE, T. R. (1989 b). Selection for host-finding in Steinernema feltiae. J. Invert. Pathol., 54: 363-372.
- GAUGLER, R., MCGUIRE, T. R. & CAMPBELL, J. F. (1989 a). Genetic variability among strains of the entomopathogenic nematode Steinernema feltiae. J. Nematol., 21: 247-253.
- GEORGIS, R. & POINAR, G. O., JR (1983). Vertical migration of Heterorhabditis bacteriophora and H. heliothidis (Nematoda: Heterorhabditidae) in sandy loam soil. J. Nematol., 15: 652-654.
- GILMORE, S. K. & POTTER, D. A. (1993). Potential role of collembola as biotic mortality agents for entomopathogenic nematodes. *Pedobiologia*, 37: 30-38.
- GLAZER, I., KOZODOI, E., SALAME, L. & NESTEL, D. (1996). Spatial and temporal occurrence of natural populations of *Heterorhabditis* spp. (Nematoda: Rhabditida) in a semiarid region. *Biol. Control*, 6: 130-136.
- HOMINICK, W. M. & BRISCOE, B. R. (1990). Survey of 15 sites over 28 months for entomopathogenic nematodes (Rhabditida: Steinernematidae). *Parasitology*, 100: 289-294.
- HOMINICK, W. M. & REID, A. P. (1990). Perspectives on entomopathogenic nematology. In: Gaugler, R. & Kaya H. K. (Eds). Entomopathogenic nematodes in biological control. Boca Raton, FL, USA, CRC Press: 327-345.
- ISHIBASHI, N. & KONDO, E. (1986). A possible quiescence of the applied entomogenous nematode, *Steinernema feltiae*, in soil. Jap. J. Nematol., 16: 66-67.
- KAYA, H. K. (1990). Soil ecology. In: Gaugler, R. & Kaya H. K. (Eds). Entomopathogenic nematodes in biological control. Boca Raton, FL, USA, CRC Press: 93-115.
- KAYA, H. K. & GAUGLER, R. (1993). Entomopathogenic nematodes. A. Rev. Ent.., 38: 181-206.
- KIM, H. K., JEONG, M. J., CHOO, H. Y. & PARK, C. S. (1988). Decrease of nematode population by introduction of nematophagous fungi into the soil as affected by inoculum concentration and temperature in vitro. *Korean J.* appl. Ent., 27: 159-164.
- KUNG, S. P. & GAUGLER, R. (1991). Effects of soil temperature, moisture, and relative humidity on entomopathogenic nematode persistence. J. Invert. Pathol., 57: 242-249.

- MOLYNEUX, A. S. (1985). Survival of infective juveniles of *Heterorhabditis* spp., and *Steinernema* spp. (Nematoda: Rhabditida) at various temperatures and their subsequent infectivity for insects. *Revue Nématol.*, 8: 165-170.
- MOYLE, P. L. & KAYA, H. K. (1981). Dispersal and infectivity of the entomogenous nematode, *Neoaplectana carpocapsae* Weiser (Rhabditida: Steinernematidae), in sand. *J. Nematol.*, 13: 295-300.
- MRAČEK, Z. (1982). Estimate of the number of infective larvae of *Neoaplectana carpocapsae* (Nematoda: Steinernematidae) in a soil sample. *Nematologica*, 28: 303-306.
- NGUYEN, K. B. & SMART G. C. (1990). Vertical dispersal of Steinernema scapterisci. J. Nematol., 22: 574-578.
- POINAR, G. O., JR (1990). Taxonomy and biology of Steinernematidae and Heterorhabditidae. *In:* Gaugler, R. & Kaya H. K. (Eds). *Entomopathogenic nematodes in biological* control. Boca Raton, FL, USA, CRC Press: 23-61.
- POINAR, G. O., JR & JANSSON, H. B. (1986a). Infection of Neoaplectana and Heterorhabditis (Rhabditida: Nematoda) with the predatory fungi, Monacrosporium ellipsosporum and Arthrobotrys oligospora (Moniliales: Deuteromycetes). Revue Nématol., 9: 241-244.
- POINAR, G. O., JR & JANSSON, H. B. (1986b). Susceptibity of Neoaplectana spp. and Heterorhabditis heliothidis to the endoparasitic fungus Drechmeria coniospora. J. Nematol., 18: 225-230.
- SCHROEDER, W. J. & BEAVERS, J. B. (1987). Movement of the entomogenous nematodes of the families Heterorhabditidae and Steinernematidae in soil. J. Nematol., 19: 257-259.
- SHAPIRO, D. I., BERRY, E. C. & LEWIS, L. C. (1993). Interactions between nematodes and earthworms: enhanced dispersal of Steinernema carpocapsae. J. Nematol., 25: 189-192.
- STURHAN, D. (1996). Seasonal occurrence, horizontal and vertical dispersal of entomopathogenic nematodes in a field. *Mitt. biol. BundAnst. Ld-u. Forstw.*, 317: 35-45.
- VÄNNINEN, I., HUSBERG, G. B., & HOKKANEN, H. M. T. (1989). Occurrence of entomopathogenic fungi and entomoparasitic nematodes in cultivated soils in Finland. *Acta* ent. fennica, 53: 65-71.
- VILLANI, M. G. & WRIGHT, R. J. (1990). Environmental influences on soil macroarthropod behavior in agricultural systems. A. Rev. Ent., 35: 249-269.
- WESTERMAN, P. R. & GODTHELP, J. M. (1990). The host-searching ability of the insect parasitic nematode *Hete-rorhabditis* sp. in sand columns. *Meded. Fac. LandbWet. Rijksuniv. Gent*, 55: 691-698.
- WOMERSLEY, C. Z. (1990). Dehydration survival and anhydrobiotic potential. In: Gaugler, R. & Kaya, H.K. (Eds). Entomopathogenic nematodes in biological control. Boca Raton, FL, USA, CRC Press: 117-137.
- WOMERSLEY, C. Z. (1993). Factors affecting physiological fitness and modes of survival employed by dauer juveniles and their relationship to pathogenicity. *In:* Bedding, R. A., Akhurst, R. J. & Kaya, H. (Eds). *Nematodes and the biological control of insect pests.* East Melbourne, Australia, CSIRO: 79-88.