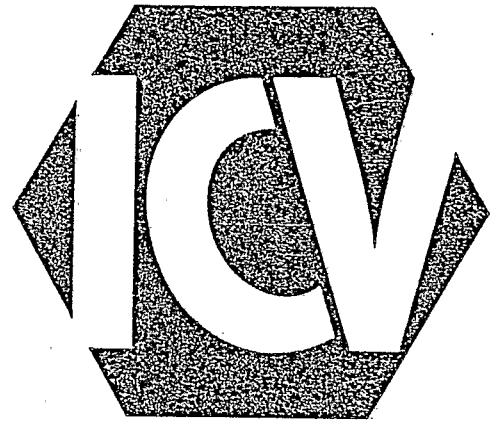


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VIIIth INTERNATIONAL CONGRESS OF VIROLOGY

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BERLIN
August 26 - 31, 1990


in affiliation with the
IUMS-SYMPOSIUM
ON NEW DEVELOPMENTS IN DIAGNOSIS
AND CONTROL OF INFECTIOUS DISEASES

An Interdivisional Meeting of the International Union of
Microbiological Societies (IUMS)

August 24 - 26, 1990

ABSTRACTS

Fonds Documentaire ORSTOM



010014584

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P70-007

THE GLYCOPROTEIN OF MARBURG VIRUS IS N- AND O-GLYCOSYLATED
C. Will, H. Feldmann, M. Schikore, W. Slenczka, and Klenk, H.-D., Institut für Virologie, Philipps-Universität, Marburg, Germany
Marburg virus is an enveloped, negative-stranded RNA virus and constitutes together with Ebola virus a new virus family, the Filoviridae. Both viruses are highly pathogenic for humans and cause a severe hemorrhagic disease. The glycoprotein (GP) is a highly glycosylated membrane protein with a molecular weight of at least 150 kd. The sugar part of the protein amounts to about 30% of the total

P70-008

ANTIGENIC CHARACTERIZATION OF HANTAVIRUSES ISOLATED IN YUGOSLAVIA

T. Avšič¹, A. Gligić², G. van der Groen³, J.W. Le Duc⁴

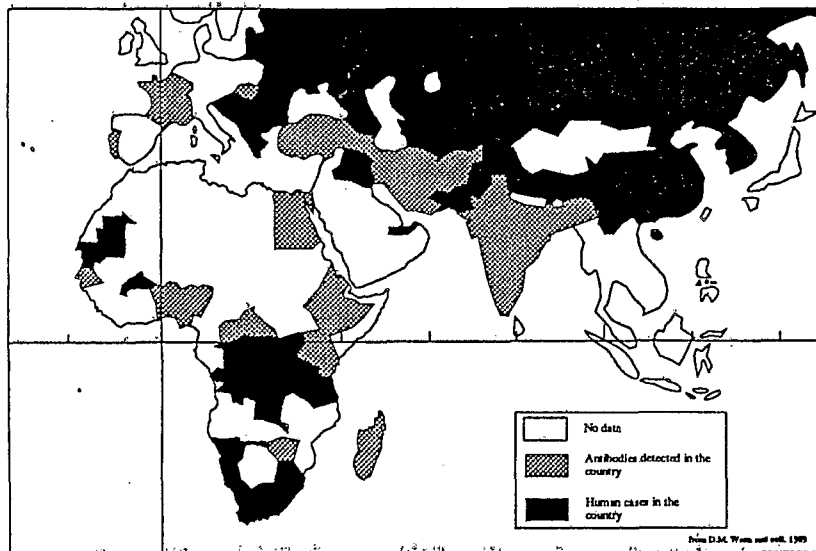
- 1 Institute of Microbiology, Ljubljana, Yugoslavia
- 2 Institute of Virology, Beograd, Yugoslavia
- 3 Institute of Tropical Med., Antwerpen, Belgium
- 4 USAMRIID, Fort Detrick, Frederick, MD, USA

Two viruses have been isolated from yellow-neck field mice (*Apodemus flavicollis*) and bank vole (*Clethrionomys glareolus*), captured in endemic areas of HFRS in Yugoslavia. Viruses were isolated directly in Vero E-6 cells and partially characterized based upon the sero-

Developing experimental models of Crimean-Congo haemorrhagic fever virus (CCHFV) using strains and hosts from West Africa

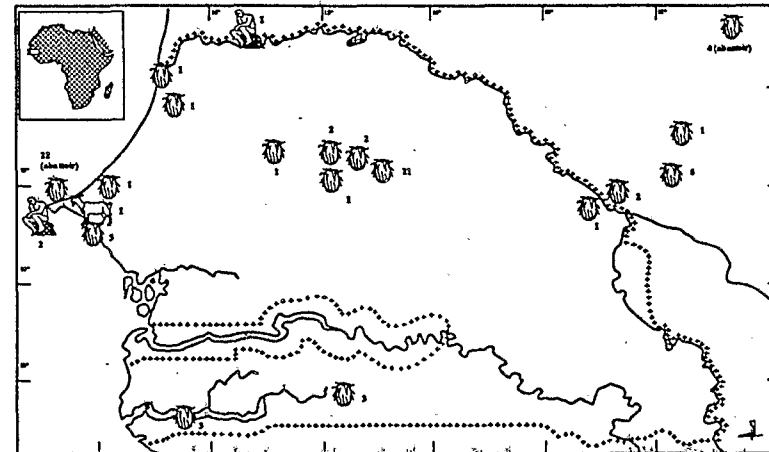
J.P. Gonzalez^{1,3}, M.L. Wilson^{2,3}, J.P. Cornet¹, F. Adam^{1,3}, B. Le Guenno³, H. Zeller³ & J.L. Camicas¹

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Geographical distribution of Crimean Congo Hemorrhagic Fever

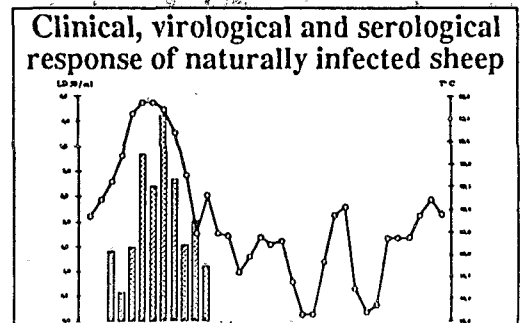
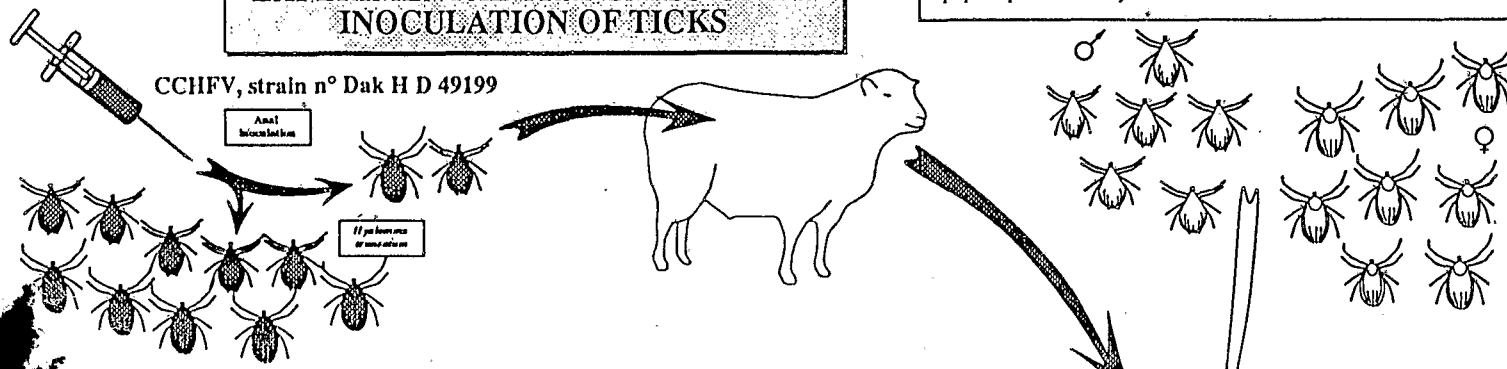
Acknowledgments :
This research is supported by grant DAMD 17-87-G-7003 from US Army Medical Research Institute of Infectious Diseases.

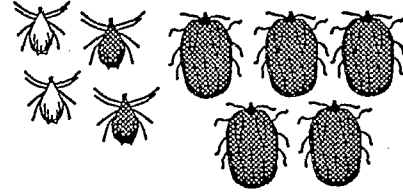
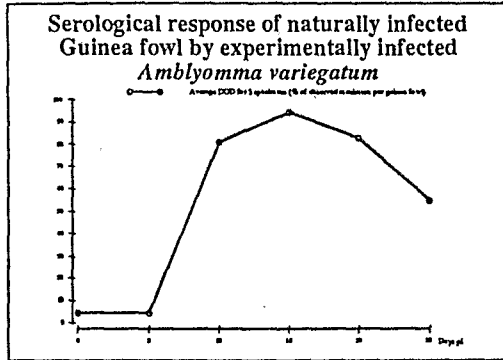
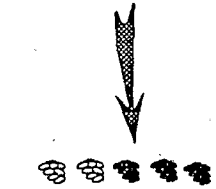
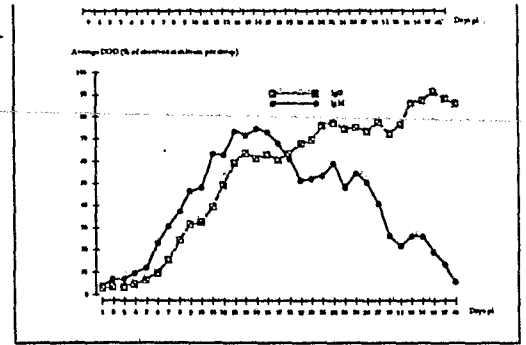
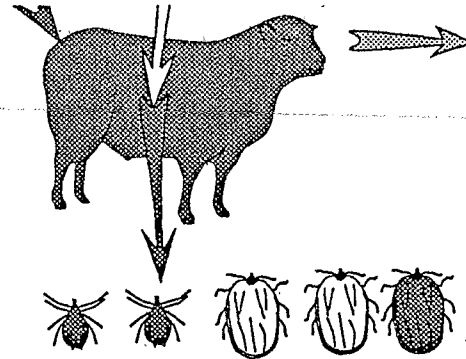
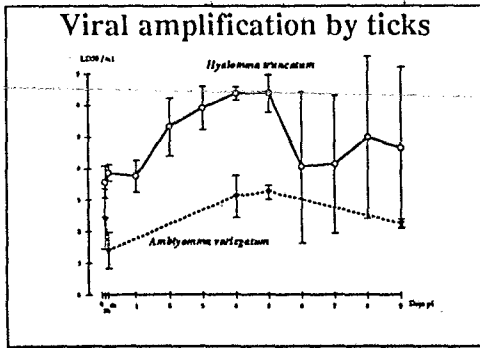
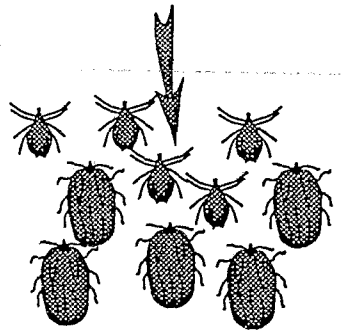


Number of CCHF virus strains isolated in Senegal and Mauritania
(data from CRORA 1990)

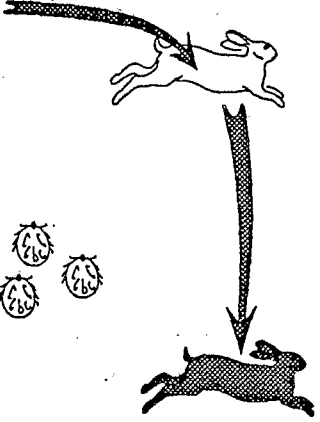
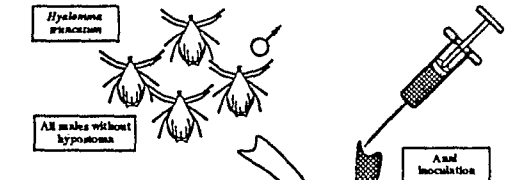
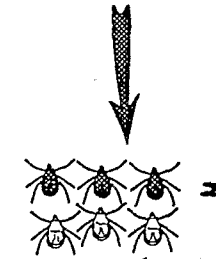
Ticks (*Hyalomma truncatum*, *Amblyomma variegatum*) naturally and experimentally infected with CCHFV have been monitored for virus replication throughout stadia development. Sheep were infected with CCHFV either by infestation of experimentally infected ticks or by inoculation. Viremia, including reisolation using suckling mice and serological response (IgM capture & IgG Elisa) have been monitored over 6 months. Pathogenic and immunological responses were observed in pregnant ewes and their offspring. Experimental transmission to laboratory rabbits by naturally infected ticks has been carried out. Wild rodents (*Mastomys erythroleucus* and *Arvicanthus niloticus*) as well as hedgehog (*Erinaceus albiventris*), guinea fowl, domestic chicken and laboratory mice were studied. The intensity and pattern of the serological response varied with hosts and inoculation route. Virus reisolation occurred in some cases (*Mastomys*, rabbit, guinea pig, mouse). Certain domestic and peridomestic animals that were naturally infected appeared to replicate the virus occasionally at a low titer and develop variable immunity. Pathogenicity seems to be limited, with no major clinical involvement in any of the species studied. These results combined with field observations (serosurveys and virus isolations) are used to propose a putative natural cycle of CCHFV in the west african environment.

EXPERIMENTAL TRANSMISSION BY INOCULATION OF TICKS





SEXUAL AND TRANSOVARIAL TRANSMISSION BETWEEN TICKS



LEGEND

Ticks:

- ♀ (female tick)
- ♂ (male tick)
- ♀ (infected female tick)
- ♂ (infected male tick)

Egg

Nymph

Virus:

- Present (shaded box)
- Absent (white box)

