





trate reductase A is demonstrable in anaerobic culture with  $\text{NO}_3^-$ ; nitrate reductase B is demonstrable in aerobic culture grown in the absence of  $\text{NO}_3^-$  (6). Extracts of cells cultivated anaerobically in the presence of  $\text{NO}_3^-$ ,  $\text{NO}_2^-$ , or  $\text{N}_2\text{O}$  yield extremely little or no tetramethyl-*p*-phenylenediamine-nitrite reductase activity. Tetrathionate reductase is produced in anaerobiosis in a medium containing  $\text{S}_4\text{O}_6^{2-}$ .

Growth and denitrification of  $\text{N}_2\text{O}$  are obtained in complex media with yeast extract or in minimal media containing acetate, butyrate, succinate, DL-lactate, or L-malate. The dinitrogen produced in stoichiometric amounts from the reduction of  $\text{N}_2\text{O}$  in the presence of succinate by cell suspensions has been measured and identified by gas chromatography (3).

**DNA base composition.** The deoxyribonucleic acid (DNA) has a buoyant density in CsCl of  $1.721 \text{ g/cm}^3$ , indicating a base composition of 62.2 mol% guanine plus cytosine (G+C) (5, 8).

**Taxonomy.** The bacterium clearly is a member of the genus *Pseudomonas*. The species known to denitrify are *P. aeruginosa*, *P. fluorescens*, *P. stutzeri* (including *P. stanieri*), *P. mendocina*, *P. pseudomallei*, *P. mallei*, *P. solanacearum*, *P. caryophylli*, and *P. pickettii*. All of these are nutritionally omnivorous and assimilate a broad range of carbohydrates and amino acids (2, 9). Physiologically, the bacterium we have encountered and described bears greatest resemblance to *P. lemoignei*. The latter assimilates only acetate, butyrate, valerate, pyruvate, succinate, and DL-3-hydroxybutyrate, and accumulates poly- $\beta$ -hydroxybutyrate as a reserve substance. It also is characterized by equally small colonies. However, *P. lemoignei* differs in growing at  $41^\circ\text{C}$ , in producing colonies that are very coherent and adherent to agar, in not assimilating DL-lactate, propionate, L-malate, and  $\alpha$ -ketoglutarate, in failing to denitrify, and in being

Is this organism a new species or is it a denitrifying variety of *P. lemoignei*? Only by studying new strains can we determine its taxonomic position. Unfortunately there are serious difficulties in isolating such strains because the culture conditions used during enrichment are not selective. They allow the growth of other denitrifying bacteria such as *P. stutzeri* and *Alcaligenes denitrificans*. It should be remembered that only one strain of *P. lemoignei* has been isolated and described (1).

We have deposited the holotype in the collection of the Pasteur Institute (number CIP 301-75).

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#### REPRINT REQUESTS

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#### LITERATURE CITED

1. Delafield, F. P., M. Doudoroff, N. J. Palleroni, C. J. Lusty, and R. Contopoulos. 1965. Decomposition of poly- $\beta$ -hydroxybutyrate by pseudomonads. *J. Bacteriol.* **90**:1455-1466.
2. Doudoroff, M., and N. J. Palleroni. 1974. *Genus I. Pseudomonas Migula 1894, 237 Nom. cons.* Opin. 5, *Jud. Comm.* 1952, 121, p. 217-243. *In* R. E. Buchanan and N. E. Gibbons (ed.), *Bergey's manual of determinative bacteriology*, 8th ed., The Williams & Wilkins Co., Baltimore.
3. Garcia, J.-L. 1974. Réduction de l'oxyde nitreux dans les sols de rizières du Sénégal: mesure de l'activité dénitrifiante. *Soil Biol. Biochem.* **6**:79-84.
4. Law, J. H., and R. A. Slepecky. 1961. Assay of poly- $\beta$ -hydroxybutyric acid. *J. Bacteriol.* **82**:33-36.
5. Mandel, M., C. L. Schildkraut, and J. Marmur. 1968. Use of CsCl density gradient analysis for determining the guanine plus cytosine content of DNA, p. 184-195. *In* L. Grossman and K. Moldave (ed.), *Methods in*