Comparison of thick blood smear and saponin haemolysis for the detection of *Loa loa* and *Mansonella perstans* infections

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Summary

In an endemic area, *Loa loa* and *Mansonella perstans* microfilariae were detected by the examination of 5 ml blood in respectively 7.4% and 26.2% of subjects who would have been erroneously considered amicrofilaraemic by the conventional method of two thick blood smears (40 µl blood). Correction factors to be applied to the results obtained with thick blood films in order to approach the true parasitological prevalences were 1.5 for *L. loa* and 1.6 for *M. perstans*. In addition, the analysis of a large volume of blood provides a better estimation of microfilarial load of the parasitized population.

Introduction

The epidemiological evaluation of filarial infections relies on clinical, parasitological, immunodiagnostic and entomological methods (WHO 1984). In *Loa loa* and *Mansonella perstans* filariases, parasitological data take into consideration the prevalence and intensity of microfilaraemia and measure different aspects of the endemicity of the parasites in the population (Noireau et al. 1989). In the field, the search for microfilariae (mf) is usually carried out by examination of thick blood smears prepared from specimens of capillary blood (Sasa 1967). However, a greater volume of blood, obtained by venipuncture, is necessary to detect low-density carriers. The concentration of mf by sedimentation of haemolysed blood was first described by Knott (1939). The most commonly used enriching techniques are saponin haemolysis (Franks & Stoll 1945), membrane filtration (Bell 1967; Dennis & Kean 1971) and density gradient centrifugation (Jones et al. 1975). Comparative studies of parasitological techniques conducted in a region where onchocerciasis is endemic are scarce (Gordon & Weber 1955; Richard-Lenoble et al. 1980) although it has been demonstrated in foci of lymphatic filariasis that the use of a concentration technique increases three- to five-fold the chances of detecting mf (Ramachandran 1975). We estimated the mf prevalence rate and microfilarial density in a population sample living in an area endemic for *Loa loa* and *Mansonella perstans* by examining thick blood films, and the results were compared with data obtained with an enrichment technique such as haemolysis/centrifugation. Haemolysis was chosen as the reference test rather than filtration since it was found to be more sensitive in that a greater volume of blood could be examined (Richard-Lenoble et al. 1980).

Materials and methods

Capillary and venous blood samples were taken from 201 adult Bantus (mean age: 35.9 ± 13.2 years; M:F sex ratio: 1.2:1) between 1000 and 1400 h. Two 20-µl thick blood films were prepared from each sample of capillary blood. After Giemsa staining, the mf were identified and counted. Citrated venous blood (5.0 ml) was lysed with 2% saponin. The mf in the sediment after centrifugation were identified and counted after Giemsa staining.
Table. Loa loa and Mansonella perstans: comparison of two parasitological techniques for the estimation of prevalences of microfilariae carriers and microfilarial densities

<table>
<thead>
<tr>
<th>Filariae identified</th>
<th>No. samples</th>
<th>Thick smear</th>
<th>Haemolysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>L. loa</td>
<td>201</td>
<td>+ 26 12.9</td>
<td>+ 39 19.4</td>
</tr>
<tr>
<td>M. perstans</td>
<td>201</td>
<td>+ 60 29.9</td>
<td>+ 97 48.3</td>
</tr>
</tbody>
</table>

MfD, Geometric mean of microfilarial densities per ml of blood.
Thick smear, 40 µl of capillary blood.
Haemolysis, 5 ml of venous blood.

The value of thick blood smears in the detection of both species of mf was studied by determining its sensitivity (or rate of co-positivity) and its negative predictive value. The correction factors to be applied to the results obtained with thick blood films in order to approach the true parasitological prevalences were calculated by dividing the number of positives observed in the examination of 5 ml of blood by the number of positive cases to be found by examination of 40 µl (Sasa 1967).

For both types of samples, the microfilarial load of the parasitized population was expressed as the geometric mean of microfilarial densities (MfD) per millilitre of blood.

Results and discussion
The results are shown in Table I. Twenty-six subjects showed L. loa mf in the thick blood films and 39 in the haemolysis test (60 vs 97 for M. perstans). The rate of Co-positivity was 0.67 for L. loa and 0.62 for M. perstans whereas the negative predictive values were respectively 0.93 and 0.74. All negative results obtained with haemolysis were also negative with the thick smears. The correction factors to be applied to the number of cases positive with the thick smears were 1.5 for L. loa and 1.6 for M. perstans.

Above the detection threshold with thick blood film (> one mf per 40 µl of blood), the quantitative distributions of L. loa and M. perstans microfilaraemia were similar with both techniques. For L. loa, the average microfilarial density calculated by thick blood films was 4.1 times that observed after haemolysis, which included low-density carriers (550.0 vs 135.5 mf ml⁻¹). For mansonellosis, the average microfilarial densities were 106.9 mf ml⁻¹ (thick smear) and 42.2 mf ml⁻¹ (haemolysis).

The evaluation of the prevalence of mf carriers is the basis of studies on human filariases (Sasa 1967), particularly filariasis due to L. loa and M. perstans (Kershaw 1950). For infection with M. perstans the pathogenicity of which is not clearly established (Janssens 1964; Wiseman 1967), there are no reliable clinical manifestations which can be taken into consideration. However, for loiasis, the prevalence of a pathognomonic symptom such as subconjunctival migration of the adult worm is an interesting epidemiological indicator which complements the estimation based on mf carriers (Noireau et al. 1990). The immunodiagnosis of L. loa filariasis enables the immunity of the amicrofilaraemic population to be assessed (Pinder 1988).
fact be applied since it has been confirmed that there is no significant difference between the number of mf in the same volume of venous and capillary blood (Gordon & Weber 1955).

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References


