

Genetic Variability and Microdistribution of *Triatoma infestans* Genotypes and *Trypanosoma cruzi* Clones in Arequipa Region (Peru)

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The genetic variability of Triatoma infestans and Trypanosoma cruzi populations was studied by isoenzyme analysis in two distinct areas of Arequipa province (Peru); one, Santa Rita de Sigüas, being an endemic area for Chagas' disease, the second, Arequipa, recently infected. Analysis of T. infestans genetic variability indicates, (i) temporal stability of genotypes found in Santa Rita de Sigüas, (ii) high genetic differences between Arequipa and Santa Rita de Sigüas populations suggesting minor contact between them, (iii) multiple origin of the T. infestans population in Arequipa, and (iv) poor dispersal capacity of T. infestans: the panmictic unit could be reduced to a house. Parasite isoenzyme analysis was performed in 29 Peruvian stocks of T. cruzi, mainly isolated from bugs taken in a single locality, Santa Rita de Sigüas. The results show, (i) a high genetic polymorphism, (ii) nine different multilocus genotypes were detected and clustered in two different clades, (iii) most of the parasite isolates pertained to one of the clade and were genetically similar to those analyzed 12 years before. This sample allowed the study of the mating system of T. cruzi in strict sympatric conditions and gave more strength to the hypothesis of the clonal structure of T. cruzi populations.

Key words: population genetics - *Triatoma infestans* - *Trypanosoma cruzi* - Chagas' disease - clones

The extent of genetic variation and population structure in *Triatoma infestans*, the principal vectors of Chagas' disease in South America, were studied in Bolivian, Peruvian and Brazilian populations by isoenzyme analysis (Dujardin & Tibayrenc 1985 a,b, Dujardin et al. 1987). The results show: (i) a relatively low rate of variability when compared with most other insects (3 variable loci among 19); (ii) an apparent absence of speciation between geographically separate populations and between domestic and sylvatic ones; (iii) free gene flow within *T. infestans* from a given locality and sometimes between neighboring localities. In Peru, genetic of *T. infestans* populations is poorly documented.

Trypanosoma cruzi, the causative agent of Chagas' disease, is present in nature as populations of "natural clones" as demonstrated by popula-

tion genetic studies; these natural clones appear to be evolving over time and dispersing over geographic locations without genetic exchange between organisms, thus maintaining genetically similar populations (Tibayrenc & Ayala 1988). Statistical tests exploring the reproductive system of population (segregation within a locus and recombination among loci) were developed in order to standardize the genetic analysis (Tibayrenc et al. 1990). Lack of evidence for segregation and recombination, indicate that genetic exchanges are limited and therefore that the population could present a clonal structure. The goals of this work were firstly, to study the epidemiological implications of the genetic variability of the *T. infestans* populations, second to analyze the genetic variability and mating system of *T. cruzi* populations in sympatric conditions, finally, to compare the present results with those obtained 12 years before (Brenière et al. 1985) and analyze the micro distribution of *T. infestans* genotypes and *T. cruzi* clones in a single locality.

MATERIALS AND METHODS

Origins of T. infestans insects and T. cruzi stocks - Table I summarizes the numbers and origin of vectors and stocks of parasites. All of the insects were identified as *T. infestans* according to

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Lent and Wygodzinsky (1979). Eighty-three bugs were captured from 8 different houses in Santa Rita de Sigüas locality, 85 km from Arequipa town (populations A to E). Population A grouped the insects from 4 different houses because of the low number of specimens. The other populations are from single houses and the maximum distance between houses is 1.6 km. Populations F and G are from the neighborhood of Arequipa town, while population H is from laboratory colonies which originated from Arequipa. Twenty-three *T. cruzi* stocks were isolated from *T. infestans* taken in Santa Rita de Sigüas, two other stocks were isolated from two different chagasic patients of Arequipa town by xenodiagnosis, and two more stocks originated from *T. infestans* taken in Moquegua department, Matalaque.

Isoenzyme analysis - For parasites and vectors, we used electrophoretic separation of isoenzymes on cellulose acetate plates (Helena, Beaumont, TX). The vector isoenzyme study was performed from thoracic extracts of each individual (adult or nymphs) according to Dujardin et al. (1985a) procedures with slight modifications. A total of 10 enzyme systems were tested on a sample of 32 individuals: alfa - glycerophosphate dehydrogenase (α GPD, EC 1.1.1.8), glutamate oxaloacetic transaminase (GOT, EC 2.6.1.1), glucosephosphate isomerase, (GPI, EC 5.3.1.9), leucine aminopeptidase (LAP, EC 3.4.11.1), malate dehydrogenase (MDH, EC 1.1.1.37), malic enzyme (ME, EC 1.1.1.40), peptidases (substrates: L-leucyl-leucine-leucine and L-leucyl-L-alanine) (PEP1 and 2, EC 3.4.11.11 and EC 3.4.11.13), 6-phosphogluconate dehydrogenase (6PGD, EC 1.1.1.44), and phosphoglucomutase (PGM, EC 2.7.5.1). The three enzyme systems previously described as variable in *T. infestans* (Dujardin et al. 1985a) were resolved from each individual (PGM, 6PGD and α GDH).

Parasites were isolated according to Tibayrenc et al. (1982) and cultured in LIT liquid medium. Parasites were harvested by centrifugation and pellets stored at -70°C . Before electrophoresis, pellets were mixed with an equal volume of enzyme stabilizer (Godfrey & Kilgour 1976) and left on ice for 20 min. They were then centrifuged at 11,000 g at 4°C for 10 min. Supernatants were aliquoted and stored at -70°C or used immediately. Two reference stocks named So34 cl4 and Mn, pertaining to natural clones 20 and 39 (see Tibayrenc & Ayala 1988) were selected as controls. Electrophoresis was performed according to Ben Abderrazak et al. (1993) with slight modifications. Twelve different enzyme systems were surveyed: glutamate dehydrogenase NAD (GD1, EC 1.4.1.2), glutamate dehydrogenase NADP⁺ (GD2, EC 1.4.1.4) glutamate oxaloacetic transaminase (GOT,

EC 2.6.1.1), glucose 6-phosphate dehydrogenase (G6PD, EC 1.1.1.49), glucosephosphate isomerase (GPI, EC 5.3.1.9), isocitrate dehydrogenase (IDH, EC 1.1.1.42), malate dehydrogenase (MDH, EC 1.1.1.37), malic enzyme (ME, EC 1.1.1.40) peptidases (substrates: L-leucyl-leucine-leucine and L-leucyl-L-alanine) (PEP1 and 2, EC 3.4.11.11 and EC 3.4.11.13), 6-phosphogluconate dehydrogenase (6PGD, EC 1.1.1.44) and phosphoglucomutase (PGM, EC 2.7.5.1).

Genetic variability - In vectors - To determine whether genetic exchange has been occurring in the studied populations, we used the (F) fixation index (Nei 1987) according to the formula: $F = (4n_{11}n_{22} - n_{12}^2) / (2n_{11} + n_{12})(2n_{22} + n_{12})$ with n_{11} and n_{22} the number of homozygotes, and n_{12} the number of heterozygotes. The deviation of F from 0 was tested by $X^2 = nF^2$ with one degree of freedom, and $n = n_{11} + n_{12} + n_{22}$ (Nei 1987). We applied the test in the entire population and in subpopulations.

Analysis of spatial subdivision, was conducted by Nei's F statistics analysis (Nei 1987). The significance of the Fst (gene frequencies differences among populations) and Fis (fixation indices of individuals relative of subpopulations) values were calculated according to Bilton (1992). This test is not applicable when only two sub populations are compared and the Workman and Niswander (1970) statistic was applied to test the homogeneity in allelic frequencies (*Pgm* and *6Pgdh* loci) between two populations.

In parasites - Population structure was explored by two set of statistical tests. The tests estimate deviations from random mating (segregation of alleles at given loci) and linkage disequilibrium (recombination of genotypes among loci). Departure from Hardy-Weinberg equilibrium (segregation within a locus) was tested by X^2 test for expected values of > 2 . The recombination tests have been detailed elsewhere (Tibayrenc et al. 1990, 1991), together with the possible biases brought by geographical distance and natural selection. Test d1 relies upon a combinatory analysis, and gives the probability of sampling the most frequent genotype as many or more times than actually observed in a population. Test d2 measure the probability of observing as many or more individuals of any genotype as actually observed of the most common genotype. Test e gives the probability of observing as few or fewer different genotypes as found in the sample. Test f estimates the probability of observing a linkage disequilibrium as high as actually found.

For estimation of phenetic divergence and clustering procedure, Jaccard's distance (Jaccard 1908) was used. It is estimated as follows:

$$D_j = 1 - (C / 2N - C)$$

where, C = number of bands common to both genotypes to be compared; N = total number of observed bands in the two compared genotypes. The UPGMA method (unweighted pair-group method with arithmetic averages, Sneath & Sokal 1963) was used to cluster the zymodemes together according to their Jaccard's distances. Dendrogram was computed by using the software kindly communicated by Serres and Roux (1986).

The polymorphism rate P, was determined by p/n with p = number of variable loci and n = number of studied loci.

We measure the clonal diversity in different populations by Whittam index (1989):

$$d = n(1 - \sum Xi^2) / (n-1)$$

with, Xi = frequency of each zymodeme and n = number of individuals.

RESULTS

Genetic variability of vectors - Enzymatic polymorphism: among 10 enzyme systems tested, 2 (ME and PEP1) made it possible to infer the activity of two different enzyme loci, consequently, the 10 enzyme systems corresponded to 12 putative loci. In the entire sample, only two loci appeared polymorphic, *Pgm* and *6Pgdh* exhibiting patterns reported before (Dujardin et al. 1985 a,b). The proportion of polymorphic loci was 0.16. Eight popu-

lations were analyzed, 5 from a single locality (Santa Rita de Sigwas), 2 from Arequipa town, and 1 from laboratory. Genotypes of the different populations are presented in Table I. The 6PGD enzyme system was variable only in bugs from Arequipa town, in natural populations and laboratory rearing.

Genetic structure of populations: departure from Hardy-Weinberg equilibrium was tested by the fixation index (F) of Nei and the results are presented in Table II. Genotype frequencies (*Pgm* and *6Pgdh* loci) of triatomines in the entire population, in Santa Rita de Sigwas and in Arequipa ones were not significantly different from those expected for random mating population. Results of F-statistic (Table II) are suggestive of inbreeding and spatial partition among the seven natural populations from Arequipa and Santa Rita de Sigwas (significant Fst). We also detected spatial partition among Santa Rita de Sigwas populations which are from different houses (populations A, B, C, D, and E). When the population B, composed of bugs taken in house no. 3, is discarded from the analysis of Santa Rita de Sigwas populations, spatial structuring disappeared. We observed differences in genotypic frequency between both natural populations from Arequipa (populations F and G) for only *Pgm* locus (Workman and Niswander test, $p < 0.01$).

Genetic variability of parasites - Among the 12 enzyme systems tested, 1 (ME) made it pos-

TABLE I
Origin of vectors and parasites from Arequipa province (Peru) and vector genotype

Population code	Locality	Capture place (house code)	<i>Triatoma infestans</i>						<i>Trypanosoma cruzi</i>		
			Number	Infected (%)	Genotypes			Number of stocks			
					PGM		6PGD				
1/1	1/2	2/2	1/1	1/2	2/2						
A	Santa Rita de Sigwas	4 houses (no. 1, 2, 4, 5)	10	60.0	1	4	5	10	0	0	4
B		1 house (no. 3)	41	95.1	0	3	38	41	0	0	15
C		1 house (no. 6)	7	14.3	0	3	4	7	0	0	1
D		1 house (no. 7)	16	43.7	2	8	6	16	0	0	3
E		1 house (no. 9)	9	11.1	0	5	4	9	0	0	0
Total			83	65.0	3	23	57	83	0	0	23
F	Arequipa	Simon Bolivar	11	-	3	5	3	8	3	0	0
G		Mariano Melgar	5	-	0	0	5	1	3	0	0
H		Laboratory rearing	14	-	11	3	0	2	5	6	2 xenodiagnosis
Total			30		14	8	8	11	11	6	
Entire population			113		17	31	65	94	11	6	27 ^a

^a: two additional *T. cruzi* stocks have been isolated from *T. infestans* captured in Matalaque locality of Moquegua province.

TABLE II
Genetic variability of *Triatoma infestans* natural population

Compared populations			Fixation index		F-Statistic					
Origin	Code	Number	PGM	6PGD	PGM			6PGD		
					Fis	Fit	Fst	Fis	Fit	Fst
Santa Rita de Siguan and Arequipa	A, B, C, D, E, F, G	7	1.41	0.05	-0.04	0.14	0.17 ^b	-0.4 ^b	-0.004	0.28 ^b
Santa Rita de Siguan	A, B, C, D, E,	5	0.12	-	-0.1	0.004	0.1 ^a	-	-	-
	A, C, D, E,	4	0.54	-	-0.1	-0.04	0.05	-	-	-
Workman and Niswander test X ²										
Arequipa	F, G	2	1.51	0.93		7.62 ^a			2.08	

a: p < 0.01; b: p < 0.001

sible to infer the activity of two different enzyme loci. The locus which zone of activity had the fastest electrophoretic migration was coded: "1", while the other one was coded "2". The 12 enzyme systems hence made it therefore possible to evidence the activity of 13 different genetic loci. Among them, 12 loci proved to be variable (polymorphism rate = 0.92). Genetic variability was interpreted when possible in terms of alleles, according to the hypothesis that *T. cruzi* is a diploid organism (Lanar et al. 1981, Tibayrenc et al. 1981). Nevertheless, at *Pgm* locus the patterns were too complex to be interpreted in terms of alleles. In this case, each distinct and reproducible pattern was considered as a distinct genotype, which precise allelic composition remained unknown (Fig. 1).

Nine distinct multilocus genotypes, or zymodemes, were detected of the 27 Peruvian *T. cruzi* stocks examined (Table III, Fig. 2). Two stocks evidenced at some loci a superposition of

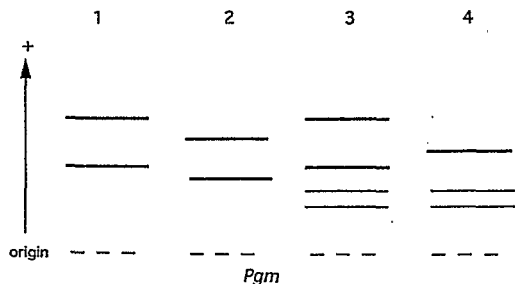


Fig. 1: non-allelic genotypes recorded at the *Pgm* locus. Slot 1: 1-4 phenotype, slot 2: 2-5 phenotype, slot 3: 1-4-6-7 phenotype, slot 4: 3-6-7 phenotype. The bold bands are intense bands.

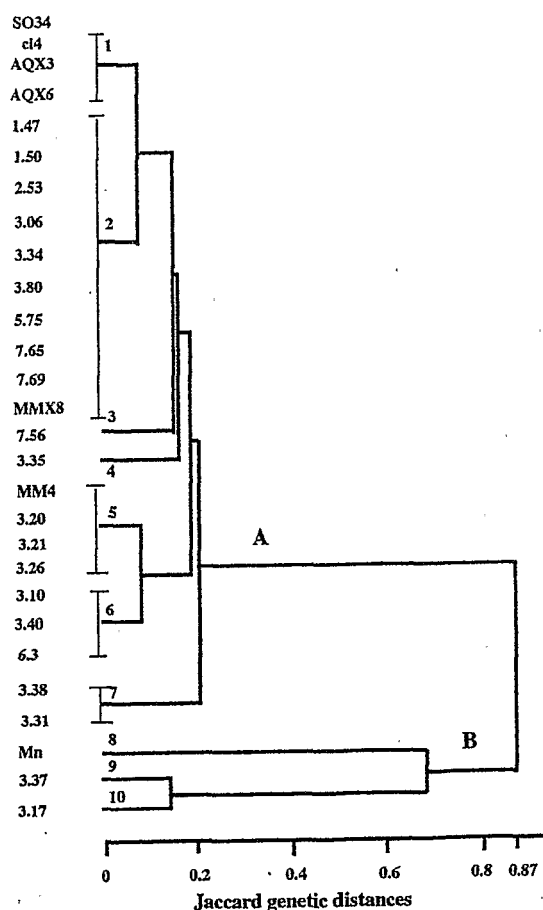


Fig. 2: UPGMA (unweighted pair group method of analysis) dendrogram derived from genetic Jaccard distances obtained from multilocus enzyme electrophoresis (MLEE) of 25 *Trypanosoma cruzi* Peruvian stocks and two reference ones.

analysis)
obtained
25 Try-
mes.

0.8 0.87

B

0.28^b

Fst

TABLE III

Multilocus genotypes of 27 *Trypanosoma cruzi* Peruvian stocks and two reference stock

Stock ^a	G6pd	Gpi	Gd nad	Gd nad +	Idh	Mdh	Mel	Me2	Pep1	Pep2	Pgm	6Pgd	Got	Zymodemes
1.47, 1.50, 2.53 3.06, 3.34, 3.80, 5.75, 7.65, 7.69, MMX8 ^b	2/2	3/3	3/3	2/2	1/1	1/1	2/2	1/1	2/2	1/1	1-4	2/4	2/2	2
3.10, 6.3, 3.40	2/2	3/3	3/3	1/1	1/1	1/1	2/2	1/1	2/2	1/1	1-4	2/4	2/2	6
3.20, 3.21, 3.26 MM4	2/2	3/3	3/3	1/1	1/1	1/1	2/2	1/1	2/2	2/2	1-4	4/4	2/2	5
3.35	2/2	3/3	3/3	2/2	1/1	1/1	2/2	1/1	2/2	2/2	1-4	2/4	2/2	4
3.31, 3.38 ^c	2/2	3/3	3/3	2/2	1/1	1/1	2/2	1/1	1/1	1/1	1-4	2/4	2/2	7
7.56	1/1	3/3	3/3	2/2	1/1	1/1	2/2	1/1	2/2	1/1	1-4	2/4	2/2	3
So34 cl4, AQX3, AQX6	2/2	3/3	3/3	2/2	1/1	1/1	2/2	1/1	2/2	1/1	1-4	4/4	2/2	1
3.30, mixture	2/2	3/3	3/3	1/1	1/1+2/2	1/1	2/2	1/1	2/2+3/3	1/1	1-4	2/4	1/1+2/2	6+(9 or 10)
3.32, mixture	2/2	3/3	1/1+3/3	1/1	1/1	1/1	2/2	1/1	2/2+3/3	1/1	1-4-6-7	2/4	1/1+2/2	6+(9 or 10)
3.37	2/2	2/2	1/1	1/1	2/2	1/1	1/1	2/2	3/3	2/2	3-6-7	4/4	1/1	9
3.17	2/2	2/2	1/1	2/2	2/2	1/1	1/1	2/2	3/3	2/2	3-6-7	4/4	1/1	10
Mn	1/1	1/2	2/2	2/2	2/2	1/1	2/2	2/2	3/3	2/2	2-5	1/3	1/1	8

^a: the number stock are from Santa Rita de Siguan, the first number indicate the code of the house, both others the triatomine number; AQX3 and AQX6 are from two patients; MMX8 and MM4 are from *Triatoma infestans* of Moquegua department. So34 cl4 and Mn are reference stocks pertaining to clone 20 and 39 respectively (see Tibayrenc & Ayala 1988); ^b: the Gd NAD genotype not determined; ^c: the 6Pgd genotype not determined.

patterns compatible with mixture of zymodemes as previously reported by Tibayrenc et al. (1986) and were excluded from genetic analysis when necessary.

The polymorphism rates (Table IV) in the entire Peruvian sample and in Santa Rita de Sigüas were very high and similar. Surprisingly, we observed a very important polymorphism in the sample limited to the single house no. 3. The clonal diversity measured by the index of Whittam, is important and maintained in the population originated from a single house; the probability to sample two different zymodemes in the population of house no. 3 is 0.92 (Table IV).

Population genetic analysis of parasites - Severe general deficit of heterozygotes is apparent in the entire sample and in the set of stocks from Santa Rita de Sigüas. Departures from Hardy-Weinberg expectations (segregation tests) were highly significant ($p < 0.001$) for glutamate dehydrogenase NADP+ locus. The H-W test was not applicable for other loci. Linkage disequilibrium statistics (recombination tests; see Tibayrenc et al. 1990) were highly significant in the entire sample and in the set of *T. cruzi* stocks from Santa Rita de Sigüas (Table IV). In the population composed of the *T. cruzi* stocks isolated from bugs taken in the house no. 3, we also observed significant tests of recombination (Table IV). In this population, under the null hypothesis of lack of departure from panmictic expectations, the probability of recording the most abundant genotype with a size as high or higher than actually observed (4 stocks) was < 0.05 (test d1); the probability of observing any genotype with a size as high or higher than the dominant genotype's one was < 0.05 (test d2); the probability of observing as few or fewer different genotypes than actually observed (7 zymodemes) was $< 10^{-4}$ (test e); the probability of observing as high a linkage disequilibrium as actually observed

was close to 0 (test f).

Clustering - The UPGMA dendrogram computed from Jaccard's distance matrix (Fig. 2) shown two clades (A and B) each composed of Peruvian isolates. Most of the Peruvian stocks were clustered together with SO34 cl4 reference stock (clone 20) and pertained to the clade A. Only two stocks, namely 3.37 and 3.17 were genetically very different, more related to MN reference stock (clone 39) and clustered in clade B; they were only sampled in the house no. 3.

Spatial distribution of clones - Genotype 2 is the most frequently sampled in Santa Rita de Sigüas. We studied its distribution among capture sites (houses) by X^2 . This genotype was significantly more abundant in the house no. 3 than in others of Santa Rita de Sigüas ($X^2 = 7.71$, $ddl = 1$, $p < 0.01$).

DISCUSSION

Genetic variability of vectors - This study realized in the province of Arequipa, Peru, allowed us to find two variable loci in *T. infestans* triatomines: *Pgm* and *6Pgdh*, whereas three different polymorphic loci were previously described in *T. infestans* species (Dujardin & Tibayrenc 1985 a,b, Dujardin et al. 1987). *T. infestans* triatomines from Arequipa and Santa Rita de Sigüas showed high genetic differences: while both *Pgm* and *6Pgdh* loci were variable in Arequipa only *Pgm* locus was variable in Santa Rita de Sigüas. This last result is consistent with a previous study realized 12 years before in the same area of Santa Rita de Sigüas (Dujardin 1990). The presence of *T. infestans* triatomines in Arequipa, described for the first time in this study, is recently and confirmed by the local authorities of the Ministry of Health and the scientists of the University of Arequipa. The origin of *T. infestans* in Arequipa could derived in part from Santa Rita de Sigüas, but the identification of allele 2 of *6Pgd*

TABLE IV
Population genetic analysis of *Trypanosoma cruzi* stocks

<i>T. cruzi</i> populations	Sample size ^a	P rate	Number of observed genotypes	Whittam ^b index	Effective of the most common genotypes	Recombination tests ^c (p)			
						d1	d2	e	f
Entire Peruvian	29	0.92	9	0.86	10	0.015	10-3	<10-4	0
Santa Rita de Sigüas	25	0.92	8	0.83	9	0.002	<10-4	<10-4	0
Stocks from population B (house no. 3)	17	0.83	7	0.92	4	0.018	0.018	<10-4	0

P: polymorphism rate; a: the mixed stocks have been counted as two stocks each; b: probability to sample two different genotypes; c: mixed stocks have been excluded.

locus, found only in Arequipa triatomines, indicate a second origin; this allele 2 is also described in Bolivian populations (Dujardin 1990) and in southern Peruvian populations (unpublished data).

Genetic variability of parasites and clonality - Among the Peruvian *T. cruzi* stocks under study we observed 9 different zymodemes clearly grouped in two principal clades A and B (Fig. 2). The 25 stocks pertaining to clade A were genetically related to those characterized 12 years before (Brenière et al. 1985, 1991) by similar isoenzyme analysis using the same reference stocks as control, clone 20 and clone 39 (Tibayrenc & Ayala 1988). Two other stocks pertaining to genotypes 9 and 10 were never described before in this region and were clustered in clade B.

Criteria of clonality have been extensively described by Tibayrenc et al. (1990). The present results agree with the hypothesis of clonality of *T. cruzi* population.

Two stocks isolated from *T. infestans* in Moquegua departement present the same genotype (2 and 5) than several stocks isolated in Santa Rita de Sigwas. The fact that these areas are geographically distant confirms the ubiquitous character of *T. cruzi* zymodemes, one of the criteria of clonality (Tibayrenc et al. 1990).

The genetic segregation and recombination tests were mostly significant in the entire population and in the population of Santa Rita de Sigwas. Significant segregation and recombination tests are also criteria for clonal structure of populations (Tibayrenc et al. 1990). Even though, reliability of H-W test, depends on the diploid hypothesis of *T. cruzi* and is limited by small sample size, recombinant tests, independent of diploid hypothesis, give more unquestionable interpretation of the results.

Geographic partition of population (absence of sympatric conditions) is a potential source of significant tests even in sexual species. That can explain the significativity of the tests when populations of Santa Rita de Sigwas and Arequipa are tested together and when the populations of the different houses in Santa Rita de Sigwas are analyzed. However, the *T. cruzi* population isolated from a single house (no. 3) is sympatric and the recombination test are still significant.

The stocks isolated from house no. 3 are clustered in two separate clades (A and B, Fig. 2) and the recombinant tests performed on clade A (data not shown) were not significant. Therefore, the presence of two cryptic species cannot be excluded. The absence of significativity of the recombination tests could be related to the presence of two cryptic species either to bias due to the application of the tests to sub phylogenic division where ge-

netic diversity is low.

Micro distribution of T. infestans genotypes and T. cruzi clones - The micro distribution of *T. infestans* genotypes and *T. cruzi* clones within individual houses of Santa Rita de Sigwas indicate that house no. 3 differs from others for: (a) a higher density of insects in this house; (b) a greater infection rate of the triatomines; (c) a different *T. cruzi* clone distribution; and (d) differences in allelic frequencies of *T. infestans*.

Therefore, the dispersal capacity of *T. infestans* is very reduced and the panmictic unit appears to be smallest than the whole locality as previously proposed by Dujardin et al. (1985). The polymorphism rate evidenced by isoenzyme in *T. infestans* taxon is very low and limits the genetic interpretation of genetic flux between populations. New markers (eg. RAPD and microsatellite DNA) has to be developed to reach more resolutive approach. The absence of homogeneity in *T. cruzi* clone distribution between house no. 3 and the others confirm the separation of the insect populations.

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