

CRYOPRESERVATION OF SEEDS FOR LONG-TERM CONSERVATION OF COFFEE GERMPLASM AND ELITE VARIETIES : SUCCESSFUL APPLICATION AT CATIE

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Fonds Documentaire IRD

Cote : Bx 22617 Ex : 1

Introduction

Though *C. arabica* seeds can withstand desiccation down to 0.06-0.08 g H₂O.g⁻¹ dw (Becwar *et al.* 1983 ; Ellis *et al.* 1990), they cannot be considered orthodox because they remain cold-sensitive and desiccation does not improve their longevity (Van der Vossen 1977; Ellis *et al.* 1990). *C. arabica* seeds are also characterized by their very short lifespan in the hydrated state (Couturon 1980).

Whatever their water content, *C. arabica* seeds do not withstand direct immersion in liquid nitrogen (Becwar *et al.* 1983). However, successful cryopreservation of zygotic embryos extracted from mature seeds has been achieved with *C. liberica* (Normah and Vengadasalam 1992), *C. arabica* (Abdelnour-Esquivel *et al.* 1992 ; Florin *et al.* 1993), *C. canephora* and the interspecific hybrid arabusta (Abdelnour-Esquivel *et al.* 1992). With all species tested, partial dehydration of excised embryos to 0.2 g H₂O/g dw was sufficient to obtain high survival after their direct immersion in LN.

Even if cryopreservation of excised zygotic embryos represents an interesting alternative strategy for long-term preservation of *C. arabica* genetic resources, this technique presents some drawbacks for routine use in coffee genebanks: i) in the case of coffee seeds, embryo extraction is very time consuming and labour intensive ; ii) low reproducibility was observed when desiccation zygotic embryos using classical desiccation methods (air-flow or silica gel); iii) all stages of the cryopreservation procedure have to be performed under aseptic conditions, which does not allow to avoid using *in vitro* culture techniques. Cryopreservation of whole seeds, instead of zygotic embryos, would allow to eliminate these drawbacks.

In this aim, the effects of several parameters of the cryopreservation protocol (desiccation, cooling, thawing and post-treatment) were investigated with *C. arabica* seeds to define conditions which would ensure survival of both the endosperm and the embryo (Dussert *et al.* 1997, 1998). It was shown that: i) the optimal water content for cryopreserving whole coffee seeds was 0.2 g H₂O/g dw; ii) a two-step freezing procedure including precooling at 1°C/min to -50°C was imperatively required to recover normal seedlings after cryopreservation; and iii) there was no effect of the thawing rate on survival of cryopreserved seeds. Under these conditions, the maximal percentage of normal seedlings produced after cryopreservation was about 30% (Dussert *et al.* 1997). In addition, it was observed that, whatever the cooling process, the survival rate of zygotic embryos extracted from cryopreserved seeds after thawing was always very high (80-90%). In this study, the reproducibility of seedling recovery was investigated by carrying out several cryopreservation experiments using the same optimal conditions. In addition, the effect of various post-thawing treatments was studied in order to improve the production of normal seedlings after cryopreservation of whole seeds.

CATIE coffee (*C. arabica*) collection is one of the most important in the world. This collection has been well characterized using molecular markers and agronomic traits. One of the objectives of CATIE is now to rationalize this collection in order to improve its long-



term conservation. Besides, excellent cryopreservation facilities are available in CATIE. Therefore, a collaborative project between CATIE and IRD, supported by IPGRI, was initiated in 1998 for the development of coffee seed cryopreservation techniques at CATIE. The aim of this project was to transfer to CATIE the methodologies for cryopreservation of coffee germplasm developed by IRD, to adapt them to local conditions and to test them on several accessions of the collection.

Materials and methods

Plant material, desiccation and cryopreservation

Fresh mature seeds of *C. arabica* var. Typica were employed. After the testa was removed, seeds were desiccated to 0.2 g H₂O/g dw by equilibration for 3 weeks under 78% RH obtained using a saturated NH₄Cl solution (Dussert *et al.* 1997). The different cryopreservation procedures (rapid cooling, two-step cooling, rapid rewarming) were carried out following the protocols described by Dussert *et al.* (1997, 1998).

Osmo-conditioning

After thawing, some seeds were osmo-conditioned for two weeks at 27°C in the dark using PEG solutions. Osmo-conditioning was carried out by placing batches of 10 seeds in Petri dishes sealed with Parafilm Ribbon, on a thin layer of cotton wool imbibed with 20 ml of aqueous PEG 6000 solution. PEG concentrations were calculated to achieve osmotic potentials of -1, -2 and -4 MPa at 27°C using the equation given by Michel and Kaufmann (1973).

Culture conditions

After thawing and/or osmo-conditioning, seeds were cultured *in vitro* for survival assessment. Disinfection and *in vitro* culture were performed as described by Dussert *et al.* (1997). Extraction of zygotic embryos and *in vitro* culture were carried out according to the method of Bertrand-Desbrunais and Charrier (1989).

Survival assessment

Both germination *sensu stricto* and development of normal seedlings were used to assess seed survival. Emergence of the hypocotyl and radicle was used as the criterion for estimating the germination rate. Seedlings which stood upright on the medium were considered normal. Excised embryos were considered viable when they stood upright on the culture medium and when their first pair of leaves was developed. The time to reach half of the final proportion (P_f) of normal seedlings, T_{50} , was estimated using the least square regression and the following model where P is the proportion of normal seedlings, T the time in days and A a treatment-dependant variable describing the synchronisation of seedlings development: $P = P_f / (1 + \exp(A(T - T_{50})))$.

Test at CATIE of cryopreservation procedures set up at IRD

In order to verify the reproducibility of results obtained at IRD (France), two cryopreservation procedures resulting from the experiments described above were performed at CATIE using *C. arabica* var. Typica seeds. In both procedures, seeds were desiccated to 0.2 g H₂O.g⁻¹ dw by equilibration for 2 weeks over a NH₄Cl saturated salt solution. In procedure 1, seeds were cooled rapidly (direct immersion into LN) and thawed rapidly in a 40°C water-bath for 3 min before zygotic embryos were extracted from cryopreserved seeds and germinated *in vitro*. In procedure 2, seeds were cooled slowly by precooling them at 1°C/min to -50°C prior to immersion into LN, thawed rapidly and germinated *in vitro*.

Table 1. Number of experiments (n) and percentage (mean, minimal and maximal values) of normal seedlings recovered from whole seeds or extracted zygotic embryos after desiccation of seeds to 0.2 g H₂O/g dw, followed by rapid cooling (direct immersion in LN; 200°C/min) or slow cooling (1°C/min to -50°C prior to immersion in LN).

	Whole seeds		Zygotic embryos	
	n	Seedlings (%) (min-max)	n	Seedlings (%) (min-max)
Rapid cooling	4	0	3	91 (83-98)
Slow cooling	7	17 (6-29)	1	70

Results

Effects of slow and rapid cooling

When cooled rapidly (200°C/min) by direct immersion in LN, none of the cryopreserved seeds produced normal seedlings. By contrast, viability of zygotic embryos extracted from rapidly cooled seeds was always very high. If seeds were precooled to -50°C at 1°C/min before immersion in LN (slow cooling), over 7 repeats, a mean value of 17% of cryopreserved seeds developed into normal seedlings. Under these conditions, some normal seedlings were always recovered after germination of whole seeds, but high variability in the final survival rate was observed (6 to 29%).

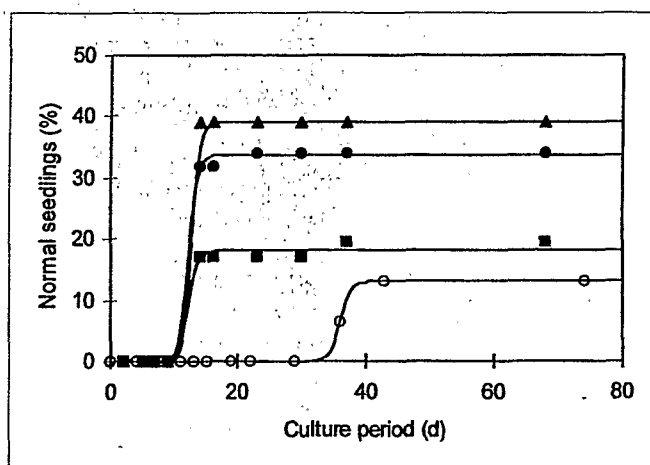


Figure 1. Evolution with time in culture under germination conditions of the percentage of normal seedlings recovered from cryopreserved seeds after a 2 weeks osmo-conditioning treatment on PEG solutions at -1 (▲), -2 (●) and -4 (■) MPa or without osmo-conditioning treatment (○).

Effect of osmo-conditioning of seeds after rewarming

Osmo-conditioning of seeds after rewarming improved the final proportion of normal seedlings recovered from cryopreserved seeds but the gain in survival decreased in line

with decreasing osmotic potential of the PEG solution used for osmo-conditioning treatment (Fig. 1). Production of normal seedlings under optimal osmo-conditioning conditions (-1 MPa) was three-fold higher than that of non osmo-conditioned cryopreserved seeds. Post-thawing seed osmo-conditioning drastically reduced T_{50} , the time to reach half of the final proportion of seedlings: when cryopreserved seeds were placed under germination conditions directly after thawing, T_{50} value was 36 d, while with osmo-conditioned seeds, it was about 12-13 d, independently of the osmotic potential of the osmo-conditioning solution.

Test at CATIE of cryopreservation procedures set up at IRD

Desiccation to 0.2 g H₂O.g⁻¹ dw by equilibration over a saturated NH₄Cl solution had no detrimental effect on seed viability since 100% of desiccated seeds developed into normal seedlings.

One hundred percents of embryos extracted from cryopreserved seeds according to the Procedure 1 (desiccation to 0.2 g H₂O.g⁻¹ dw, rapid cooling and rapid thawing) developed into normal seedlings. These seedlings showed a normal development after transfer under greenhouse conditions.

All seeds cooled slowly and placed under germination conditions directly after rewarming (Procedure 2) showed hypocotyl extrusion and radicle growth. Further development into normal seedlings occurred in 24% of cryopreserved seeds. This final percentage of normal seedlings is very similar to those observed at IRD over 7 repeats (Table 1).

Discussion

When seeds of *C. arabica* at 0.2 g H₂O/g dw were cooled rapidly (200°C/min), none of them developed into normal seedlings. This result is consistent with those of Becwar *et al.* (1983) who showed that *C. arabica* seeds did not survive after immersion in LN, even if all freezable water had been removed from the seeds. By contrast, when seeds were slowly precooled to -50°C at 1°C/min before immersion in LN, an average value of 17% (over 7 repeats) of cryopreserved seeds developed into normal seedlings. It is thus clear that slow precooling of *C. arabica* seeds had a dramatic effect on their survival and their capacity to develop normally (Dussert *et al.* 1998). However, in view of the high variability observed for the survival rate over 7 experiments, improvement to the method appeared necessary before routine use and various post-thawing treatments were investigated.

It was shown for the first time that seed osmo-conditioning (osmo-conditioning) carried out after thawing had a dramatic beneficial effect on the proportion of normal seedlings recovered after cryopreservation and on seedling growth rate: under optimal osmo-conditioning conditions (-1 Mpa for 2 weeks), the percentage of cryopreserved coffee seeds which developed into normal seedlings was three-fold that of non osmo-conditioned cryopreserved seeds (39 % vs 13%) and the time to reach half of final percentage of normal seedlings (T_{50}) was about three-fold lower (13 versus 36 d). To our knowledge, the effect of seed osmo-conditioning after cryopreservation has been investigated previously in one study on celery seeds only (Gonzales-Benito *et al.* 1995). In this study, no effect of osmo-conditioning could be found since cryopreservation did not affect germination rates and T_{50} values. Seed osmo-conditioning was first employed to improve their rate and uniformity of germination (Heydecker *et al.* 1975). Since then, a beneficial effect of osmo-conditioning after seed ageing has been observed both on germination percentage and germination rate with numerous species (Bewley and Black 1994; Bray 1995).

Thus, even if the percentage of seeds which developed into normal seedlings remained relatively low in comparison with that obtained from excised zygotic embryos, a combination of slow cooling and osmo-conditioning treatment could represent a simple and

efficient complementary option to field conservation for genebanks which cannot afford *in vitro* culture facilities. Moreover, this method might be simplified by using a -80°C freezer for precooling seeds to -50°C and could thus become more easily employed routinely in a larger number of genebanks maintaining coffee genetic resources. However, additional research should be undertaken to optimize osmo-conditioning conditions and to carry out direct germination tests under greenhouse or nursery conditions.

In cases where very high survival rates are required for routine use, we propose a second alternative approach based on the extraction of zygotic embryos after rewarming of seeds, which would allow to avoid most of the drawbacks of cryopreservation protocols developed for zygotic embryos. Equilibrating coffee seeds under 78% RH allowed seeds to reach optimal water content for cryopreservation in a very easy and reproducible manner. This method also allows the processing of large amounts of seeds at the same time. Moreover, aseptic culture conditions are requested after thawing only.

Both procedures proposed in this study were successfully applied at CATIE. Optimization of post-thawing osmo-conditioning conditions and testing of the two procedures on 14 accessions of *C. arabica* are currently under investigation at CATIE. Some preliminary results indicate that up to 70% of normal seedlings could be recovered from cryopreserved seeds (without zygotic embryo extraction) under some osmo-conditioning conditions. Testing of optimized protocols on 14 accessions of *C. arabica* will provide preliminary information on the feasibility of establishing a *C. arabica* germplasm cryobank at CATIE.

Abbreviations

CATIE: Centro Agronomico Tropical de Investigacion y Enseñanza; dw: dry weight; LN: liquid nitrogen; IRD: Institut de recherche pour le développement; IPGRI: International Plant Genetic Resources Institute.

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Association Scientifique
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18°

COLÓQUIO CIENTÍFICO
INTERNACIONAL SOBRE
EL CAFÉ

Helsinki, 2-6 août 1999

ISBN 2-900212-17-0

**DIX-HUITIEME COLLOQUE SCIENTIFIQUE
INTERNATIONAL SUR LE CAFE**

Helsinki, 2 - 8 août 1999

Association Scientifique Internationale du Café (ASIC)
Siège social : 18, rue de la Pépinière, 75008 PARIS

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