

International Journal for Parasitology 29 (1999) 415-418



Research note

Schistosoma bovis: vaccine effects of a recombinant homologous glutathione S-transferase in sheep

Denis Boulanger^{a, *}, Dominique/Schneider^a, Jean-Philippe/Chippaux^a, Bertrand/Sellin^a, André Capron^b

^a Centre de Recherche sur les Méningites et Schistosomoses (CERMES/OCCGE/ORSTOM), WHO Collaborating Centre for the Control of Schistosomosis, BP 10887, Niamey, Niger ^bInstitut Pasteur, Lille, France

Received 13 July 1998; received in revised form 16 November 1998; accepted 16 November 1998

Abstract

The economic importance of the trematode *Schistosoma bovis* in African livestock has justified the development of a specific vaccine. Administered preventively to sheep, rSb28GST—the only molecule cloned from *S. bovis* which has demonstrated vaccine potentialities in goats and cattle—reduced the mean worm burden in vaccinated animals and improved their health status compared with that of non-vaccinated controls. As in goats, but not in bovines, the fecundity of the settled worm pairs was not modified. Therefore, rSb28GST can be proposed as a universal tool for the prevention of clinical disorders engendered by the main schistosome species affecting domestic ruminants in the African continent. © 1999 Australian Society for Parasitology. Published by Elsevier Science Ltd. All rights reserved.

Keywords: Schistosoma bovis; Vaccination; Sheep; Recombinant protein; Glutathione S-transferase

Schistosoma bovis is responsible for a helminthiasis affecting livestock in Africa, with prevalences reaching up to 90% in calves [1]. Its severe pathological impact, also demonstrated in small ruminants [2], has led to attempts to control the disease through vaccination. Irradiated larvae have been efficacious [3], but their large-scale utilisation remains logistically difficult. Recently, the preventive administration of a recombinant S. bovisderived glutathione S-transferase (rSb28GST) resulted in dissociated effects according to the

* Corresponding author. Tel: +227-75-20-45; fax: +227-75-31-80; e-mail: boulange@niamey.orstom.ne. definitive host species: in cattle, it markedly diminished the fecundity of paired adult worms [4] whereas, in goats, it significantly reduced worm burden [5]. Because the ovine commercial market is of crucial importance in many sub-Saharan countries, it was considered important to check whether or not the rSb28GST was protective in sheep and, if so, what was the expression of the vaccine-induced immunity.

The same schedule of immunisations/infection as in the caprine experiments [5] was used. Briefly, eight male Sahelian sheep (14–26 kg at the start of the experiment) received two s.c. injections, 6 weeks apart, of 100 μ g of rSb28GST [6] in PBS, with Freund's complete, then incomplete, adjuvant v/v.

0020-7519/99/\$-see front matter © 1999 Australian Society for Parasitology. Published by Elsevier Science Ltd. All rights reserved. PII: S0020-7519(98)00222-7





Table 1

Clinical and parasitological data in sheep twice-immunised with the recombinant Sb28GST and in controls receiving adjuvant alone^a

	rSb28GST	Controls	\mathbf{p}^{b}	Р
Clinical status				
PCV (%)°	-8.2 ± 3.7	-13.8 ± 4.2	-41%	< 0.05
Body weight (kg) ^c	-2.9 ± 1.3	-3.8 ± 1.2	-23%	< 0.05
Faecal egg excretion				
Per 24 h	2511 ± 1216	4786±2892	-48%	< 0.05
Per 24 h per female	33.8 ± 18.9	36.2±23.6	-7%	NS
Adult worms				
Males	84.4 <u>+</u> 28.3	142.1 <u>+</u> 44.8	-41%	< 0.05
Females	86.0 <u>+</u> 23.9	136.6 <u>+</u> 41.8	-37%	< 0.05
Total	170.4 <u>+</u> 39.5	278.6 <u>+</u> 79.3	-39%	< 0.05
Tissue eggs				
Liver (%)	16 <u>+</u> 14	28 ± 30		
Small intestine (%)	39 <u>+</u> 26	46±29		
Large intestine (%)	46 ± 28	27 ± 23		
Total ($\times 10^3$)	434±368	527 <u>+</u> 332	-18%	NS
Per female	4571 ± 2688	3875 <u>+</u> 2199	+18%	NS
Intra-uterine eggs	16.1 ± 4.3	19.1 ± 3.4	-16%	NS

^aData are expressed as mean \pm S.D.

^b Protection (%) = $(A - B)/B \times 100$, where A is the rSb28GST-immunised group average and B the control group average.

°Expressed as the mean difference between the start and the end of the experiment.

Control animals received adjuvant alone. Four weeks after the boost, sheep were anaesthetised then infected percutaneously with a single dose of 2000 cercariae of a local strain of S. *bovis*. Perfusion took place 20 weeks later. Statistical comparisons were made by the non-parametric Mann–Whitney test.

Clinical and parasitological data are summarised in Table 1. One animal died before the end of the experiment in each group (weeks 16 and 18, respectively). Packed cell volume values were negatively correlated with the intensities of faecal egg excretion. Decreases of body weights and of PCV were significantly lower in the vaccinated group. Mean worm burdens and faecal egg excretion were significantly reduced. In contrast, the mean number of eggs trapped in the tissues or counted in the uterus of each female schistosome on the day of perfusion were not affected by the immunisation schedule. Thus, the mean numbers of faecal or tissue eggs per female were comparable in both groups. Individual levels of protection were somewhat heterogeneous. Figure 1 shows that, in terms



Fig. 1. Individual levels of reduction of faecal egg excretion in rSb28GST-immunised sheep compared to the control average.

of faecal egg excretion, six vaccinated sheep had protection levels ranging from 47% to 68%, whereas two animals displayed levels of egg excretion close to the control group average.

The dual goal of this experiment was first to

	Goats ^a	Sheep
Female worm burden	46% ^b	-37% ^b
Tissue eggs	35% (NS)	-18% (NS)
Tissue eggs per female	7% (NS)	+18% (NS)
Faecal egg excretion 24 h	30% (NS)	-48% ^b
Faecal egg excretion per 24 h per female	+ 29% (NS)	-7% (NS)

Table 2 Compared protective effects of the rSb28GST in two species of small ruminants

^a Data from Boulanger et al. [5].

^bP < 0.05; NS: not significant.

check in sheep the vaccine capacity of the rSb28GST already demonstrated in goats [5], and second to assess the expression of vaccine-induced immunity. Table 2 compares the protective effects on parasitological parameters obtained in both species. At a somewhat lower level, a similar worm burden reduction was obtained in sheep. In contrast, a sharp effect on faecal egg excretion was obtained in sheep, whereas it did not reach significance in goats. Taken as a whole, one can assume that both species reacted roughly in the same manner. The immunological peculiarities of goats and sheep when facing parasites [7] required this confirmation.

The lack of modification of faecal, tissue and intra-uterine eggs per female schistosome is a strong argument in favour of an immune target being the worm capacity to establish an adult population in its definitive host, rather than its egg-laying productivity. Both male and female worms were equally affected, suggesting that paired schistosomes, not migrating larvae, are likely to be the real targets. These data confirm that small ruminants do not develop the same protective mechanisms as other animal models of schistosomosis [8]. For instance, vaccination with the Schistosoma mansoni-derived 28GST of Patas monkeys challenged with Schistosoma haematobium clearly targeted worm fecundity [9]. The contrast is especially striking with the results of a trial performed on calves [4], showing that vaccination with native SbGST induced significant reductions in faecal egg counts and in tissue egg densities, although adult worm counts were not modified. However, rSb28GST was able to affect both parameters when calves were sub-

Ş

jected to natural infection with *Schistosoma matthei* [10]. It should also been kept in mind that, unlike cattle [11], the development of a naturally acquired resistance does not seem to occur in sheep following repeated exposures to the parasite in the field [12], suggesting that the two host species do not develop the same mechanisms when subjected to natural schistosome infection. Basic studies on their immune responses to experimental challenge are under investigation and would be of great interest in the context of comparative immunology.

Taken together with the results derived from experiments carried out in cattle against *S. bovis* and against *S. matthei*, the results presented here suggest that the recombinant molecule rSb28GST can be proposed as a relevant tool to control the important economic impact of schistosomes affecting ruminants in Africa. In the more general framework of trematodosis, they confirm previous observations showing that preventive immunisation of sheep and cattle with *Fasciola hepatica*derived GST also results in significant worm burden reductions [13].

Acknowledgements

This work was supported by the Science and Technology for Development (STD3) Programme of the Commission of the European Communities, contract No. TS3-CT91-0030, by the Institut Français de Recherche Scientifique pour le Développement en Coopération (ORSTOM), by Pasteur Institute in Lille (France) and by the Organisation de Coordination et de Coopération pour la lutte contre les Grandes Endémies (O.C.C.G.E.). The authors would like to acknowledge François Trottein and Claude Godin for providing the recombinant antigen. The technical assistance of D. Couret, A. Islamane, S. Kadri, I. Mahamadou, A. Sidiki, F. Sidikou and A. Yacouba is greatly appreciated.

References

- Majid AA, Marshall TF de C, Hussein MF, et al. Observations on cattle schistosomiasis in the Sudan, a study in comparative medicine. I. Epizootiological observations on S. bovis in the White Nile Province. Am J Trop Med Hyg 1980;29:435-441.
- [2] Kassuku A, Christensen NO, Nansen P, Monrad J. Clinical pathology of *Schistosoma bovis* infection in goats. Res Vet Sci 1986;40:44–47.
- [3] Majid AA, Bushara HO, Saad AM, et al. Observations on cattle schistosomiasis in the Sudan, a study in comparative medicine. III. Field testing of an irradiated S. *bovis* vaccine. Am J Trop Med Hyg 1980;29:452–455.
- [4] Bushara HO, Bashir MEN, Malik KHE, et al. Suppression of *Schistosoma bovis* egg production in cattle by vaccination with either glutathione S-transferase or keyhole limpet haemocyanin. Parasite Immunol 1993;15:383–390.
- [5] Boulanger D, Trottein F, Mauny F, et al. Vaccination of goats against the trematode *Schistosoma bovis* with a

recombinant homologous schistosome-derived glutathione S-transferase. Parasite Immunol 1994;16:399–406.

- [6] Trottein F, Godin C, Pierce RJ, et al. Inter-species variation of schistosome 28-kDa glutathione S-transferases. Mol Biochem Parasitol 1992;54:63–72.
- [7] Huntley JF, Patterson M, Mackellar A, et al. A comparison of the mast cell and eosinophil responses of sheep and goats to gastrointestinal nematode infections. Res Vet Sci 1995;58:5–10.

Ĺ

- [8] Capron A, Riveau G, Grzych JM, et al. Development of a vaccine strategy against human and bovine schistosomiasis. Background and update. Mem Inst Oswaldo Cruz 1995;90:235-240.
- [9] Boulanger D, Warter A, Trottein F, et al. Vaccination of patas monkeys experimentally infected with *Schistosoma haematobium* using a recombinant glutathione S-transferase cloned from S. mansoni. Parasite Immunol 1995;17:361-369.
- [10] De Bont J, Vercruysse J, Grzych JM, et al. Potential of a recombinant *Schistosoma bovis*-derived glutathione *S*transferase to protect cattle against experimental and natural *S. mattheei* infection. Parasitology 1997;115:249–255.
- [11] Bushara HO, Majid AA, Saad AM, et al. Observations on cattle schistosomiasis in the Sudan, a study in comparative medicine. II. Experimental demonstration of naturally acquired resistance to S. bovis. Am J Trop Med Hyg 1980;29:442–451.
- [12] Majid AA, Hussein MF, Taylor MG. Age specific prevalence and intensity of *S. bovis* infection in sudanese desert sheep in the White Nile Province. Res Vet Sci 1983;35:120– 121.
- [13] Spithill TW, Dalton JP. Progress in development of liver fluke vaccines. Parasitol Today 1998;14:224-228.