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Evidence of pollen transport by the Sanaga River on the Cameroon shelf

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Abstract. Thirty-eight samples were taken from the banks of the Sanaga River and its main tributaries draining different plant ecosystems, but also from rivers in the coastal basins of Cameroon. This study aims to characterize the origins of pollen and the dominant mode of transport of pollen inputs to the continental shelf. The classical method of pollen spectra analysis and multivariate statistical analysis revealed three groups of samples corresponding to the three main ecosystems in which the samples were taken. Pollens typical of the northern savannas are found in the spectra of samples taken further downstream from the Sanaga River in forest areas and not in samples from rivers draining mainly the forested coastal basins. The level of similarity between groups and the spatial evolution of spectra from upstream to downstream are related to the fluvial transport of pollen.

Keywords. Palynology, Pollen, Savanna, Forest, River transport, Sediments, Sanaga, Cameroon.

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1. Introduction

There are many past and recent studies on pollen fluxes in current deposits in Africa. Some have been the subject of attempts to model paleoenvironments and paleoclimates by various authors [Elenga et al., 2000, Jolly et al., 1998a, Lebamba et al., 2009b, Peyron et al., 2006], current [Bengo, 1996, Frédoux and Maley, 1996, Hooghiemstra et al., 1986], and fossils

[Bengo and Maley, 1991, Dupont and Agwu, 1991, Elenga et al., 2004, Giresse et al., 2009, Lebamba et al., 2009a, Lézine et al., 2009, Ngomanda et al., 2008, Reynaud and Maley, 1994]; and others, have shown the importance of wind contributions to the Sahara [Nguestop et al., 2004], and Sahel winds blowing north and which contributed to the loess deposits in Europe [Haerserts, 1985].

But what about the origin of the terrigenous deposits of the southern part of the Gulf of Guinea which we know that the winds of the Harmattan blowing towards the ocean, pass over the north-

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ern limit of the Gulf, and hardly exceeding the Forest/Savannah limit in Cameroon?

Thus, the research questions that this work must answer are:

- Are the pollen flows covering the bottom of the Cameroonian basin brought back by the Harmattan?
- Otherwise, how can we prove that they are part of the procession of fluvial contributions?

The choice of the Sanaga basin is justified by the fact that the Sanaga River, through its main tributaries, successively crosses large plant units (savannah, dense forest and coastal forest). This study thus aims to characterize the origins of pollens and the dominant mode of transport of pollen inputs to the continental shelf. Since the continental winds are weak, the hypothesis of wind inputs can be excluded, or else these inputs are negligible. The importance of fluvial inputs remains to be demonstrated, for example if specific savanna pollens could be found in samples taken downstream in forest areas.

2. Material and methods

2.1. Study site

2.1.1. Location and climate

The study area occupies the central region of Cameroon (Figure 1), including the large Sanaga Basin and the coastal basins facing the Atlantic Ocean. The Sanaga Basin, with an area of 133,000 km², extends between latitudes 3-32' N-7-22' N and longitudes 9-45' E-14-57' E [Laclavère, 1979]. It is bounded to the west by the Cameroonian volcanic Dorsale, and to the north, by the Adamaoua Plateau. The Sanaga Basin is subject to the moist monsoon air masses that flow from the ocean to the mainland, and incidentally in its northern part, to the winds of the Harmattan direction NE that blow from the continent to the ocean [Leroux, 1983, Piton, 1987, Suchel, 1988].

2.1.2. Soils and vegetation

Plant units have a zonality in relation to soils [Martin, 1966, Segalen, 1967] mainly ferralitics related to the quality of substrate [Guillet *et al.*, 1996, Nzila *et al.*, 2018], and the more or less important

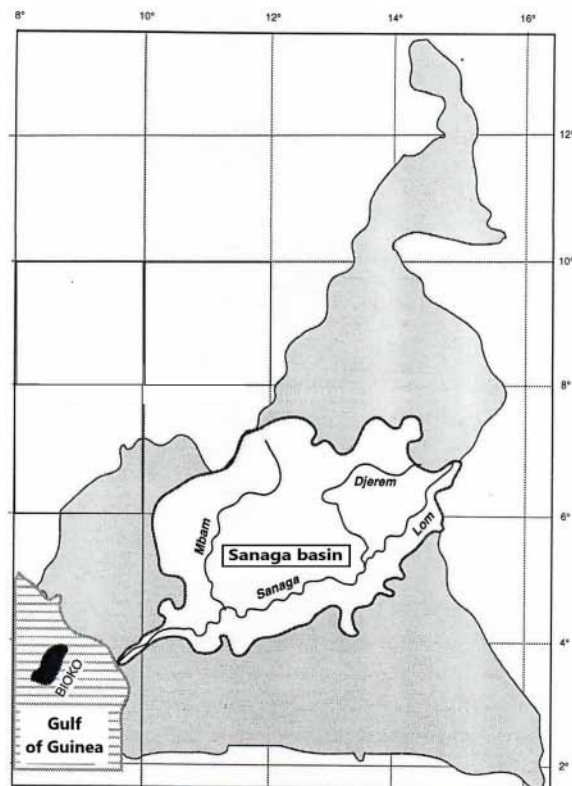


Figure 1. Location of the Sanaga basin in Cameroon.

annual distribution of precipitation and temperatures (Figure 2). Wet dense forests in the south, savannahs in the centre and north of the country [Aubréville, 1948, Letouzey, 1985, White, 1986]. Dense humid forests are made up of semi-deciduous forest, evergreen biao forest, and evergreen littoral forest, each of which is characterized by specific taxa [Puig, 2001]. Savannahs are of two types on the way to the Adamoua Plateau in the north: the grassy and shrub-rich peri-forest savannahs (*Bridelia ferruginea*, *Terminalia glaucescens*, ...) and the Sudan-Guinean savannahs that are enriched by new species (*Combretum molle*, *Daniellia oliveri*, *Entada abyssinica*, *Syzygium macrocarpum*, ...). Plant associations linked to specific edaphic or climatic conditions are present, such as: near the coast, mangroves at *Rhizophora mangle*, *Avicennia africana* [Boyé *et al.*, 1975, Letouzey, 1985, Din, 1991]; and at altitude, the mountain formations at *Podocarpus*, *Olea*, *Rapanea* [Maley, 1987]. Moreover, anthro-

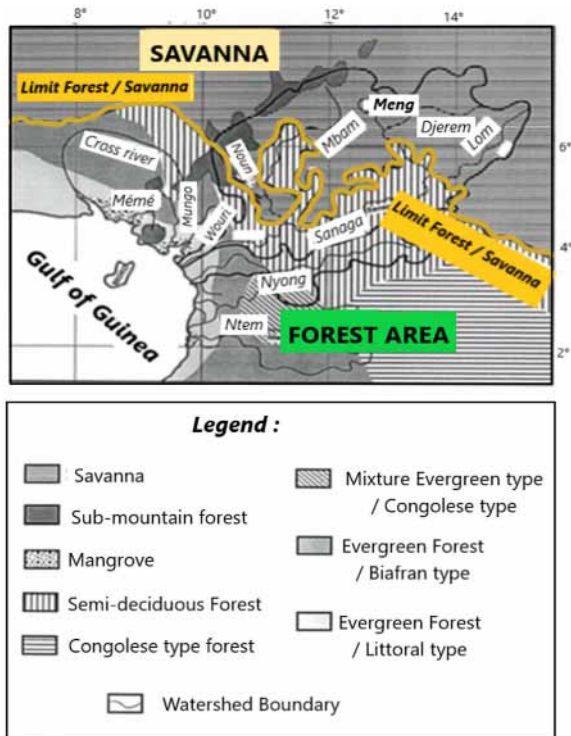


Figure 2. Main types of vegetation in the Sanaga basin.

pogenic actions related to logging, industrial and food crops do not prevent the regeneration of the forest, which is currently advancing on the savannah [Letouzey, 1985, Maley, 1990, Youta Happi, 1998].

2.1.3. Fluvial inputs of the Sanaga River

Continental erosion depends mainly on soil condition and vegetation cover. On the Edea station, the average turbidity, corresponding to all the suspended solid matter of the Sanaga basin transported to the ocean, was estimated annually at 6,000,000 tons of silt and clay, of which 2,500,000 from the Mbam by the importance of agriculture in The Bamiléké [Nouvelot, 1972, Olivry, 1977].

2.2. Sampling

The Sanaga River is fed along its course by several rivers that drain small basins covering various ecological and floristic areas. Mineral and organic particles of different sizes are deposited on the riverbed or on the banks. In order to be likely to find pollen

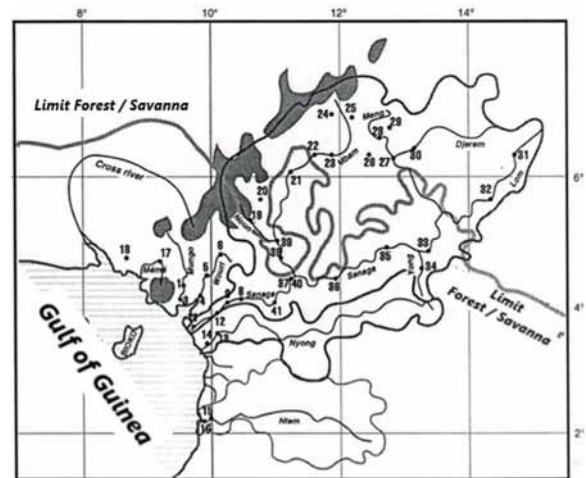


Figure 3. Location of samples on rivers.

grains, sampling was done in fine leashes or deposits of the lutite class, namely silts and clays. These leashes, considered surface deposits, can contain both river-drained inputs and the fallout from atmospheric dust laden with local or even allochthon pollens. Samples were taken throughout the basin, along the Sanaga River and on a dozen of its tributaries. A total of thirty-eight samples (Table 1 and Figure 3) were collected, including twenty-seven from upstream and downstream of the Sanaga Basin, and another 11 samples were taken near the offshore outlets of the main coastal rivers draining only watersheds located in dense forest areas, in order to reassure ourselves of the assumption of a dominant mode of transport.

2.3. Pollen identification

After the treatment of samples using conventional methods [Cour, 1974, Faegri and Inversen, 1975], the identification of pollens was made at the level of the Family, Gender and Species by referring to the blades of the Reference Collection of current pollens of the Montpellier Laboratory and monographs of the region [Bonnefille and Rioulet, 1980, Caratini *et al.*, 1974, Salard-Chebouldaef, 1980, 1981, 1982, 1983].

2.4. Statistical analysis of data

Data placed in a contingency array matrix [Benzecri and Benzecri, 1980, Volle, 1993] is preparing for mul-

Table 1. Distribution of the samples taken in the different rivers and under different floristic facies

River	Savanna	Forest	Mangrove	Total
Sanaga	1	9		10
Mbam	6			6
Djerem	5			5
Lom	2			2
Noun	2	1		3
Yong		1		1
Mémé and Lobé		1	1	2
Wouri et Mungo	4	1		5
Nyong		2		2
Ntem		1	1	2
Total	16	19	3	38

tivariate analyses, such as Factor Correspondence Analysis (FCA) and Principal Component Analysis (PCA). These analyses are supplemented by a Dendrogram hierarchical classification (Clusters analyses and Neighbour joining), so as to define the relationships and nuclei of affinities between the samples [Roux, 1985]. Data processing, graphic representation of results, and dendrogram of hierarchical classification were made with MacMul, GraphMu and MacDendro software [Thioulouse, 1990] and PAST software [Hammer et al., 2001]. In the study region, this type of statistical analysis has recently been adopted on current pollen [Jolly et al., 1996] and fossil [Reynaud-Farrera, 1995].

2.5. Pollen spectra

Pollen spectra are analysed using two transects from upstream to downstream (mouth), i.e. from savannah areas to dense forest areas:

- a Mbam transect with 10 samples from the Mbam Basin;
- and a Sanaga transect consisting of 12 samples taken upstream of the Sanaga/Mbam confluence.

From the downstream confluence of the Sanaga and the Mbam to the mouth, five samples complete the two transects mentioned above. These allow tracking the variation in samples along the route,

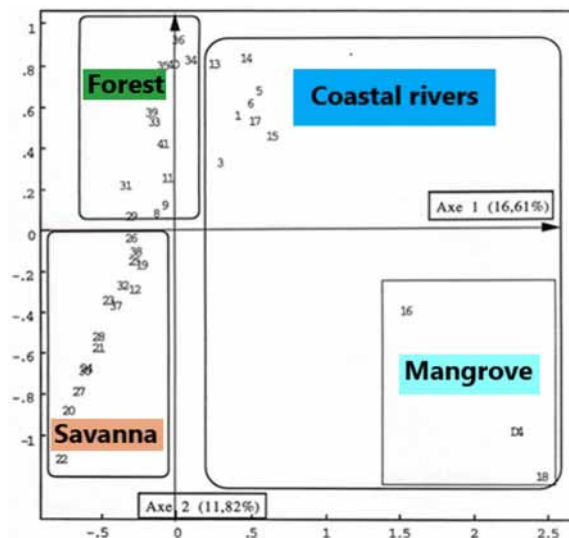


Figure 4. With all samples (38) and taxa (237).

from their area of origin, appearance or abundance in the samples to the mouth, of certain characteristic taxa revealed by the statistical processing of the data.

3. Results

3.1. Pollen analysis

The pollen analysis of the 38 samples resulted in the identification of 237 taxa, the most represented of which are carried out in Table 2.

3.2. Multivariate analyses

3.2.1. Factor Correspondence Analysis (FCA)

Factor analysis of the matches was applied to two separate matrixes, the initial matrix containing all the analysed samples (Figure 4); the other includes only samples from the Sanaga Basin (Figures 5 and 6).

3.2.2. The Principal Component Analysis (PCA)

The point cloud on the foreground factor (Figure 7) from a main component analysis (PCA) on the Sanaga Basin samples confirms FCA 2. Almost all taxa are grouped in the centre, except those with high contributions ending up towards the periphery of the ellipsoid of correlations (Graminae, Spores, *Alchornea* and *Uapaca guineensis*); as is the case on the

Table 2. Initial matrix of frequent taxa in various plant ecosystems

TAXONS / ECOSYSTEMS	Ech 1	Ech 3	Ech 4	Ech 5	Ech 6	Ech 9	Ech 11	Ech 12	Ech 13	Ech 14	Ech 15	Ech 16	Ech 17	Ech 18	Ech 19	Ech 20	Ech 21	Ech 22	Ech 23	Ech 24	Ech 25	Ech 26	Ech 27	Ech 28	Ech 29	Ech 30	Ech 31	Ech 32	Ech 33	Ech 34	Ech 35	Ech 36	Ech 37	Ech 38	Ech 39	Ech 40	Ech 41	
1 <i>GRAMINEAE</i>	GRAM	2.44	10.04	4.56	3.01	0.00	20.08	17.48	17.06	37.14	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
2 <i>Scieros</i>	Spe	33.17	40.83	8.26	12.51	2.27	14.13	12.36	24.76	24.11	10.82	13.53	14.53	16.13	12.67	29.53	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
3 <i>Alchornea</i>	Alch	3.81	4.92	2.70	2.71	10.23	9.81	8.53	2.86	20.37	11.27	6.58	2.77	7.53	4.14	3.63	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
4 <i>Uapaca ligustralis</i>	Uap2	0.00	0.43	0.00	1.30	4.55	10.46	10.23	12.94	3.81	1.85	1.40	7.49	9.49	6.99	3.51	0.52	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	
5 <i>Indelermia</i>	Inde	14.83	4.68	1.35	2.41	8.82	14.83	15.59	3.24	2.86	11.89	7.41	8.49	6.99	3.51	0.52	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	
6 <i>Uapaca guineensis</i>	Uap1	0.00	2.44	2.98	55.27	0.60	1.41	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
7 <i>Rhizophora</i>	Rhiz	0.00	1.70	0.54	1.51	2.27	2.19	3.28	2.06	2.86	6.48	1.72	6.68	0.69	1.28	0.49	3.31	1.77	4.22	3.16	7.43	2.59	3.14	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
8 <i>CYPREPACIAE</i>	CYPE	4.88	0.85	1.27	2.11	1.14	4.18	3.84	2.94	0.00	3.70	4.27	2.77	4.30	2.29	1.55	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
9 <i>COMBRETACEAE</i>	COMB	2.44	0.64	0.54	0.00	0.00	0.42	0.85	0.88	0.00	2.78	2.47	0.00	1.04	3.61	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
10 <i>Myrsineae / Maki</i>	MYR	0.00	1.60	0.41	0.60	0.00	1.28	2.13	1.76	1.90	0.60	1.90	0.45	0.00	1.41	0.49	0.52	0.31	0.00	0.00	1.00	0.82	0.74	0.78	1.24	0.85	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	
11 <i>Elaeagnaceae</i>	ELA	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
12 <i>CASALPINACEAE</i>	CASP	0.00	0.00	0.00	0.00	0.00	0.42	0.00	0.00	1.90	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
13 <i>Platanaceae</i>	PLAT	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
14 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
15 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
16 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
17 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
18 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
19 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
20 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
21 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
22 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
23 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
24 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
25 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
26 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
27 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
28 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
29 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
30 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
31 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
32 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
33 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
34 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.00	1.26	0.00	1.47	0.95	0.00	0.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
35 <i>Myrsineae</i>	MYR	0.00	0.43	0.27	1.51	0.																																

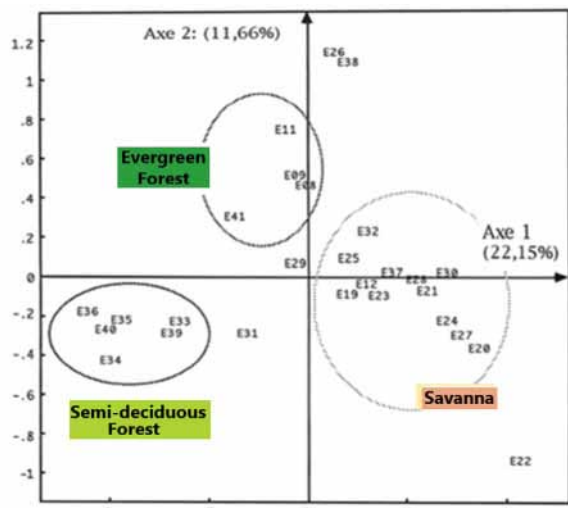


Figure 5. All samples, without *Rhizophora*.

diagonal (Figure 8) formed by the factor plane of the PCA where axis 1 is placed on the axis of the abscises and on the axis of the ordinated.

3.3. Pollen spectra

Recurrent taxa, characteristic of different plant ecosystems and highlighted by analysis of statistical data, are shown in Figure 11 showing the evolution in space of pollen spectrum through the Sanaga and Mbam transects.

4. Discussion

4.1. Conformity of the image of the local plant cover in the samples of river banks

Generally for many palynological studies, the authors base their interpretations based solely on analyses of pollen diagrams. Here, the important results of this work and the analysis of pollen spectra were easily supported by the prior use of a multivariate statistical treatment.

The FCA 1 carried out on all 38 samples allowed to obtain on the factor plane the first two axes bringing together the bulk of inertia, a cloud of points around axis 1 of the abscises that discriminates on either side of the origin the samples Sanaga basin from those in coastal basins, particularly samples (4, 16 and 18).

The eccentricity of these three samples is due to the over-representation of pollens from *Rhizophora*, a major taxon of mangroves. Undeniably, the Sanaga group is characterized by the absence of *Rhizophora* pollens. On the other hand, the identity of the coastal basin sample group, opposed to the Sanaga group, is defined not only by the presence of *Rhizophora*, but also by other taxa characteristic of the coastal evergreen forest.

By examining only the 27 samples of the Sanaga Basin in FCA 2 this time, the effect of The Over-representation of *Rhizophora* is resolved. According to Figure 5 and in relation to axis 1, the samples are arranged on either side of the origin: on the right, the samples of the savannah, and on the left those of the forest. On the other hand, on the forest samples side, axis 2 discriminates above the samples of the evergreen forest, and at the bottom the samples of the semi-deciduous forest. Simultaneous projection or projection on different graphs of point clouds of samples or taxa from the same FCA, in this case FCA 2, shows the taxa that play a great role in the disposal of samples (Figure 6). Such taxa have also been revealed by the PCA (Figure 7), almost all taxa are grouped in the centre, except those with high contributions that are located towards the periphery of the ellipse of correlations (Graminae, spores, *Alchornea* and *Uapaca guineensis*). The arrangement of these taxa on the same side of the positive values of the axis shows a “size effect”, since they are the most abundant. These same taxa are aligned on the diagonal formed by axis 1 (Figure 8) that could be considered an ecological indicator.

The Dendrogram analyses applied to the initial matrix and the reduced matrix to the Sanaga basin samples complements the factor analyses in the same direction. According to Figure 9 from a “Clusters analysis” shows that the aggregation levels and the proximity of the samples are not random, they rather come from the affinities provided by the absence, presence or abundance of the associated taxa. Thus, Figure 10 from a “Neighbour joining” correlates the taxa in the disposition of the samples in a Dendrogram hierarchical classification.

From what is done, the factor analyses and the “Cluster analyses” have revealed different ecosystems, namely the savannah, the semi-deciduous forest, the evergreen forest and the mangroves. In summary, savannah samples contain high propor-

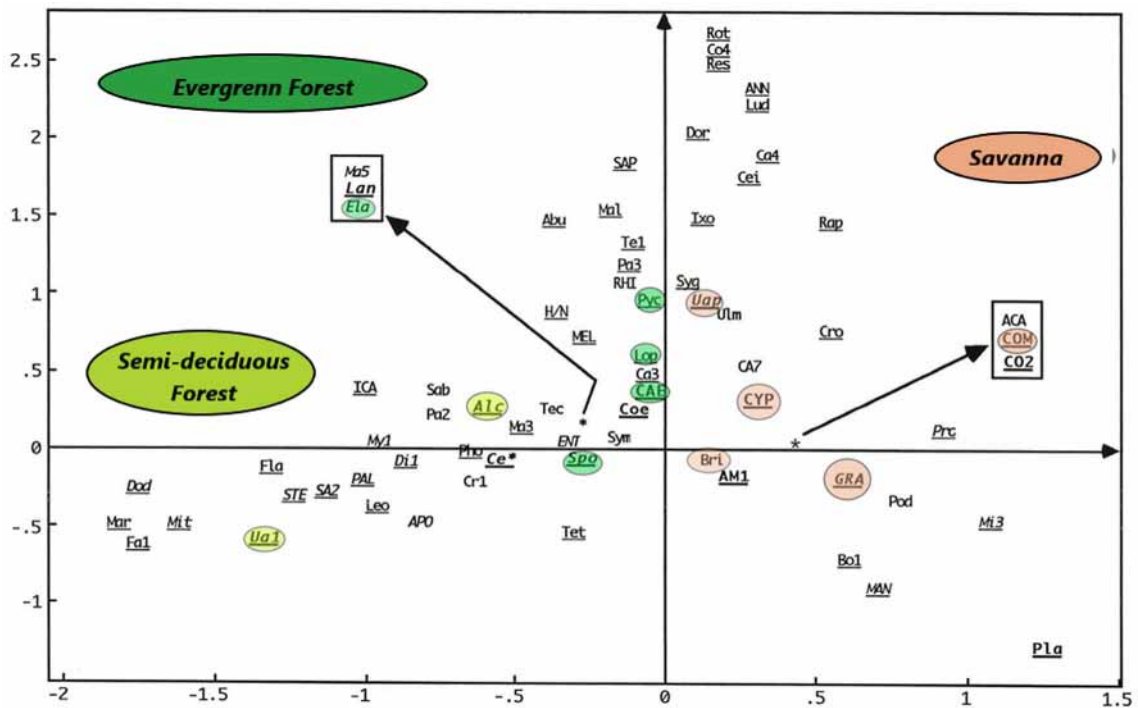


Figure 6. AFC 3: Point cloud of taxa, including those with high contributions and inertias are located on either side of the origin, and correlate the distribution of the samples on axis 1: positive values for savannah, and negative values for forests.

tions of Poaceae (Graminae), the omnipresence of Asteraceae (Compositae), Cyperaceae, *Bridelia*; the semi-deciduous forest is characterized *Uapaca guineensis*, *Elaeis*, *Myrianthus*, the Ulmaceae and the Sapotaceae; the evergreen forest characterized by the taxa of Caesalpiniaceae, *Pycnanthus*, *Lophira* and *Syzygium*; and mangroves by *Rhizophora*, and *Pandanus*. We note the presence of some ubiquitous taxa such as *Alchornea* and Spores. If the high pollen values of *Alchornea* result from the fact that it is a pioneer shrub of forest recruits, while the high proportions of spores are related to the Pteridophytes and Bryophytes that grow abundantly in wetlands [Letouzey, 1968].

These multivariate analyses provide further evidence to the numerous palynological studies that link pollen analyses to botanical harvests in the different ecosystems of the surrounding vegetation cover [Bengo, 1996, Kimpouni et al., 2014, Lebamba et al., 2009a, Vincens et al., 2000, Youta Happei, 1998].

4.2. Characterization of pollen transport from the continent to the ocean (Guttman effect)

The FCA 2 cloud has a parabolic form that generally indicates that the factors of axis 1 and 2, as a matter of linear independent principle, are in fact related to one or more other axes [Benzecri and Benzecri, 1980]. For interpreting this unique factor, a projection of the different dots is made on a straight line joining the two ends of the cloud (Figure 12). Samples (8, 9, 11, and 41) of the evergreen forest downstream of the Sanaga are placed in a middle or intermediate position between the savannah sample group and the semi-deciduous forest group. If factor 1 had instead been defined as an ecological indicator of samples, on the other hand axis 2 would be the single factor that would bring together some savannah and semi-deciduous forest taxa in samples further downstream, and this factor could be transport, a phenomenon also observed with the hierarchical classification where the evergreen forest samples are closer to those of the savannah, then geographically it is rather

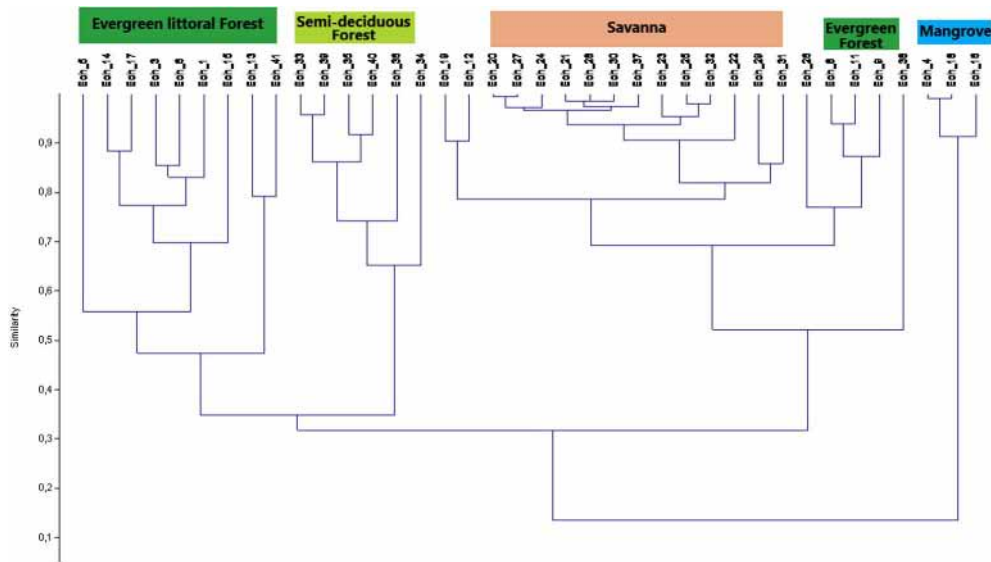


Figure 9. Clusters analyses_Mode Paired group/Correlation.

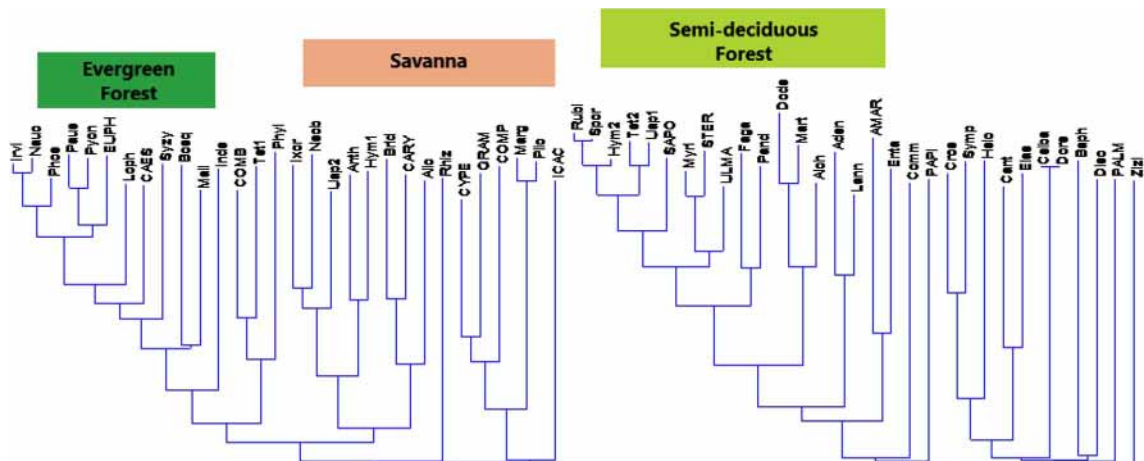


Figure 10. Neighbour joining_Mode Correlation/Final branch on the matrix reduced to 27 samples from the Sanaga basin.

After the deduction of pollen transport by the Guttman effect following the correspondence factor analysis, based on the analysis of pollen spectra of transects, it can therefore be inferred that a good part of the pollen that arrives on the platform of Cameroon is transported by the Sanaga River. This was also observed in dredging samples on the Cameroonian platform where the large proportions of pollen processions were located in the axis of river deposits [Bengo, 1996]. Similarly, studies similar to

ours have also shown the more or less important role in pollen transport towards the mouth of the Senegal River [Lézine and Edoth, 1991], and Trinity River in America [Traverse, 1990].

The isotopic study of carbon 13 on the same samples as ours [Bird *et al.*, 1994] gave average values of -19 to -17‰ for upstream Sanaga samples and -27‰ for forest samples. Thus, the values of -25 to -24‰ near the mouth of the Sanaga could therefore be explained by a mixture of organic matter coming

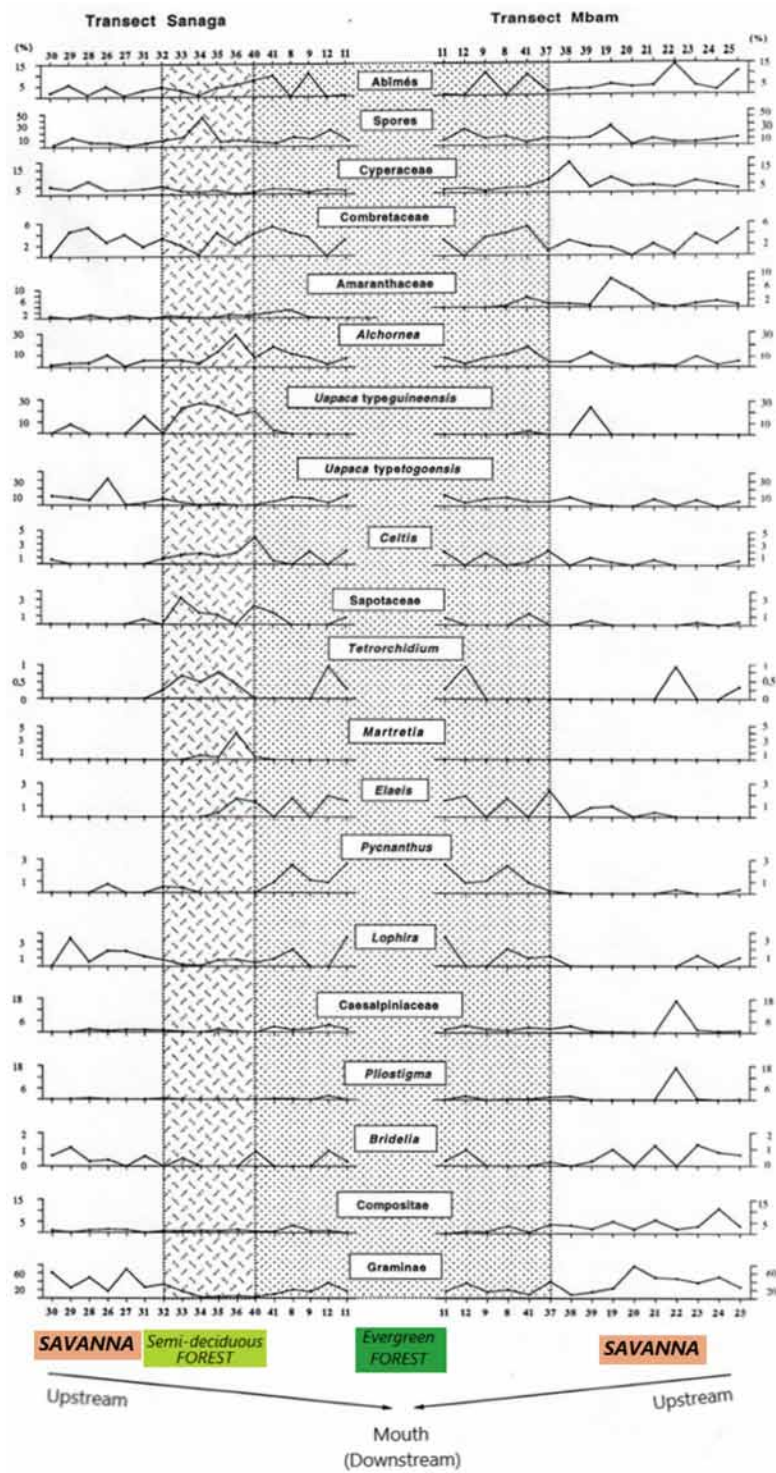


Figure 11. Evolution spectra of taxa characteristic of different ecosystems following the Sanaga and Mbam transects.

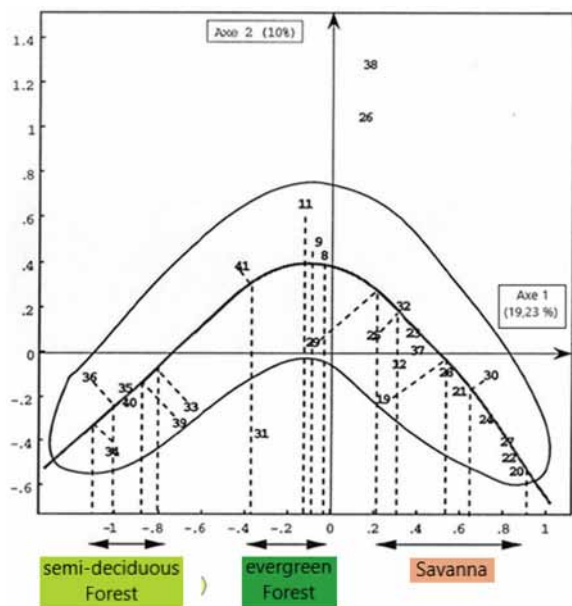


Figure 12. AFC 2: Guttman effect.

from the savannahs by the Mbam and the Sanaga. Once again, the results of the isotopic study converge completely with those of palynology.

5. Conclusions

Pollen trapped in the bank silt quite clearly records the image of the surrounding vegetation cover. The methodology and analyses adopted for this palynological study finally led to highlight the terrigenous origin of sedimentary inputs from the Cameroon continental shelf and the preponderance of fluvial transport. Multivariate analyses allowed to corroborate the pollen identification, to differentiate and group the samples according to the plant ecosystems where they were taken. The Guttman effect and the Dendrogram hierarchical classification showed that the matching of the different groups of samples was related to a single factor which is pollen transport. Compared to the regional atmospheric circulation, wind transport was found to be negligible, whether it is the Harmattan winds blowing towards the ocean or the trade winds and westerly monsoon winds that are directed towards the mainland. The absence of typical forest taxa downstream in the savanna samples, and the presence of typical savanna taxa upstream

in the downstream samples, leads to the conclusion that there is fluvial transport of Graminae and others towards the mouth. Thus, it can be noted that, on the West African coasts, pollen transport by wind is important; whereas, on the southern coasts of the Gulf of Guinea, it is essentially fluvial through the drainage of large basins. These results of pollen dynamics could be transposed to a basin other than that of the Sanaga.

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