

Complete genome sequences of two *Klebsiella pneumoniae* phages from Dakar, Senegal

Issa Ndiaye,^{1,2} Laurent Debarbieux,³ Moussa Moise Diagne,⁴ Ousmane Sow,¹ Abdoulaye Cissé,¹ Bissoume Sambe Ba,⁵ Cheikh Fall,¹ Yakhya Dieye,¹ Ndongo Dia,⁴ Abdoulaye Seck,^{1,2} Guillaume Constantin de Magny^{6,7}

AUTHOR AFFILIATIONS See affiliation list on p. 3.

ABSTRACT Two bacteriophages (phages) of *Klebsiella pneumoniae* were isolated from sewage water collected from Dakar, Senegal. Phage vKpIN17 belongs to the *Przondovirus* genus within the *Autographiviridae* family, with double-stranded DNA genomes, whereas vKpIN18 belongs to the *Webevirus* genus of the *Drexelviridae* family.

KEYWORDS bacteriophages, *Klebsiella pneumoniae*, bacteriophage therapy, Healthcare associated infections

Using a clinical strain of *Klebsiella pneumoniae* KP26 isolated from healthcare-associated infection in a Children's Hospital Center Albert Royer of Fann in Dakar, two lytic bacteriophages were isolated from two community sewage water (14.695121–17.455776; 14.685763–17.46923) in Dakar, Senegal. Twenty milliliters of sewage sample was centrifuged at 5,000 *g* for 10 min, and then the supernatants were filtered through a 0.22 μ m pore size membrane filter. We assessed the filtered supernatants individually for phage presence using double layer agar method (1). Phage plaques were processed by three rounds of purification and amplified as previously described (2). The two phages produced clear plaques with a large halo (Fig. 1).

Genomic DNA was isolated from high titer stocks ($>10^9$ PFU/mL). Briefly, 1 mL of phage lysate was treated with 10 μ L of DNase I (20 U) and 4 μ L of RNase A (20 mg/mL), incubated for 30 min at 37°C, followed by DNA extraction using phenol-chloroform method (2). Library preparations were performed using 1 ng of DNA and Nextera XT DNA library preparation kits (Illumina, San Diego, CA, USA) and executed according to the manufacturer's protocol. Whole-genome sequencing was performed on Illumina iSeq100 sequencers utilizing the 300-cycle i1 Reagent V2 Kit (Illumina, San Diego, CA, USA).

A total of 299,580 and 323,244 (2×150 bp) paired-end reads were generated, respectively, for vKpIN17 and vKpIN18, and quality control was performed with FastQC v0.12.1 (3). Reads were trimmed using trim-galore v0.6.10 (4). The *de novo* assembly was performed using SPAdes v3.15.5 (5) with careful parameters. Contig coverage and assembly validation were performed with BMap v 35.85 (6). Furthermore, reads were sorted and indexed using Samtools v1.18 (7) and were submitted to assembly error corrections using Pilon v1.24 (8). Phage termini were identified with PhageTerm (9). Predicted coding sequences (CDS) were annotated using Pharokka v1.3.0 (10). For taxonomic classification, closely related genomes were obtained from the NCBI database.

The vKpIN17 and vKpIN18 genome sizes were 40,702 and 48,639 bp, respectively, with GC contents of 53.35% and 50.4%, and mean coverage of 1,674 \times and 1,481 \times . Both phages' genomes are permuted and feature redundant ends. There are, respectively, 54 and 84 predicted CDS of which 24 (44%) and 48 (57%) are hypothetical proteins for vKpIN17 and vKpIN18. CDS with homology to other known genes encode, among others, structural elements, DNA, RNA and nucleotide metabolism, and host lysis. No genes associated with lysogeny (e.g., integrases), virulence, toxin, transfer RNAs, clustered

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Department of Biology, New York, USA

Address correspondence to Issa Ndiaye,
seydina.ndiaye14@gmail.com.

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FIG 1 Plaque morphology of phages (A) vKpIN17 and (B) vKpIN18.

regularly interspaced short palindromic repeats, or antibiotic resistance were detected within their genomes.

The closest phages of vKpIN17 and vKpIN18 were *Klebsiella* phage K11 (accession: [NC_011043.1](#)) (95.99%) and *Klebsiella* phage Kp8 (accession: [NC_048700.1](#)) (95.70%), respectively. The phage K11 belongs to the *Przondovirus K11* species, *Przondovirus* genus, and *Autographiviridae* family, while Kp8 belongs to the *Webervirus KLPPOU149* species, *Webervirus* genus, and *Drexelviriidae* family.

We have identified two phages, vKpIN17 and vKpIN18, that show significant promise as candidates for therapeutic applications. However, further characterization is needed to explore their putative therapeutic value.

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I.N. conceptualized the study, performed sampling, curated the data, performed formal analysis and investigation, designed the methodology, helped with software, wrote the original draft, and reviewed and edited the manuscript. L.D. reviewed and edited the manuscript and supervised the study. M.M.D. designed the methodology and reviewed and edited the manuscript. B.S.B. reviewed and edited the manuscript. O.S. helped with sampling and reviewed and edited the manuscript. A.C., C.F., Y.D., and N.D. reviewed and edited the manuscript. A.S. conceptualized the study, administrated the project, reviewed and edited the manuscript, and supervised the study. G.C.d.M. conceptualized the study, administrated the project, reviewed and edited the manuscript, acquired funding, and supervised the study.

AUTHOR AFFILIATIONS

¹Pole de Microbiologie, Institut Pasteur de Dakar, Dakar, Senegal

²Faculté de Médecine, Pharmacie et Odontostomatologie, Université Cheikh Anta Diop, Dakar, Senegal

³Laboratoire de Bactériophage, Bactérie, Hôte, Département de Microbiologie, Institut Pasteur Paris, Paris, France

⁴Département de Virologie, Institut Pasteur de Dakar, Dakar, Senegal

⁵WCARO, World Health Organization, Dakar, Senegal

⁶MIVEGEC, Univ. Montpellier, CNRS, IRD, Montpellier, France

⁷MEEDiN, Montpellier Ecology and Evolution of Disease Network, Montpellier, France

AUTHOR ORCID*s*

Issa Ndiaye  <http://orcid.org/0000-0001-8918-187X>

Laurent Debarbieux  <https://orcid.org/0000-0001-6875-5758>

Moussa Moise Diagne  <http://orcid.org/0000-0001-5461-5623>

Ousmane Sow  <http://orcid.org/0000-0001-9338-6544>

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AUTHOR CONTRIBUTIONS

Issa Ndiaye, Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Software, Writing – original draft, Writing – review and editing | Laurent Debarbieux, Supervision, Writing – review and editing | Moussa Moise Diagne, Methodology, Writing – review and editing | Ousmane Sow, Writing – review and editing | Abdoulaye Cissé, Writing – review and editing | Bissoume Sambe Ba, Writing – review and editing | Cheikh Fall, Writing – review and editing | Yakhya Dieye, Writing – review and editing | Ndongo Dia, resources, Writing – review and editing | Abdoulaye Seck, Conceptualization, Project administration, Supervision, Writing – review and editing | Guillaume Constantin de Magny, Conceptualization, Funding acquisition, Project administration, Supervision, Writing – review and editing

DATA AVAILABILITY

The data for the two phages are available in the European Nucleotide Archive (ENA) under the project number [PRJEB68306](https://www.ebi.ac.uk/ena/record/PRJEB68306) with accession numbers [ERP153263](https://www.ebi.ac.uk/ena/record/ERP153263) (vKpIN17) and [ERP153263](https://www.ebi.ac.uk/ena/record/ERP153263) (vKpIN18) for raw sequences reads and accession numbers [GCA_963669465](https://www.ebi.ac.uk/ena/record/GCA_963669465) (vKpIN17) and [GCA_963669475](https://www.ebi.ac.uk/ena/record/GCA_963669475) (vKpIN18) for genome sequences.

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