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Histeridae (Staphyliniformia: Coleoptera) from Brazil: an overview

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ABSTRACT. Histerid beetles represent a group that has historically received little attention from entomologists, especially in Brazil, which makes it difficult not only to understand biodiversity and its conservation, but also to develop research using these beetles in applied areas. Here we present an overview of the biodiversity of Histeridae in Brazil, focusing on the following information i) historical data and the current richness known for Brazil, including a list of species; ii) the main authors who described the Histeridae biodiversity recorded in the Brazilian territory, and in relevant entomological collections; iii) the literature to identify Histerids; iv) the known biological associations and collection methods; v) and the conservation of Brazilian species; in addition, to proposing goals for future studies. Most of the data presented comes from the Taxonomic Catalog of the Brazilian Fauna (TCBF), part Histeridae, a Brazilian initiative to generate knowledge about the faunal biodiversity present in the country. Our data indicate that to date six subfamilies, 16 tribes, 132 genera, 607 species and one subspecies are known in Brazil. We believe that the development of taxonomic tools that contribute to the knowledge of biodiversity, as well as the increase in taxonomic revisions, constitute the guiding principle to boost research in Histeridae.

KEY WORDS. Biodiversity, clown beetle, conservation, South America, Neotropical.

INTRODUCTION

The Histeridae can be recognized by their compact and retractile body, short elytra that leave the last two abdominal tergites visible (propygidium – 6th tergite and pygidium – 7th tergite) and the geniculate antennae with compact apical club (Fig. 1).

Histerid beetles are mostly predators, mainly of dipterous larvae. They are morphologically very diverse in view of their adaptations to different environments and associations with other animals (Kovarik and Caterino 2001, Leivas et al. 2013, Kovarik and Caterino 2016, Leonel and Leivas 2023, Dégallier et al. 2020).

These beetles are relevant to Applied Sciences such as Forensic Entomology (Mise et al. 2007, 2010, Aballay et al. 2013); Archaeoentomology (Kirgis 2023); Biological Conservation (Brustel 2004, Gomy and Millarakis 2012, Vieira et al. 2018); harmful to beekeeping (Krüger et al. 2017) and meliponiculture (Coletto-Silva and Freire 2006, Silva-Neto et al. 2019); pest control in pastures (Kovarik and Caterino 2001, 2016), banana planting (Mesquita 2003, Domínguez et al. 2018), agave planting (Salcedo-Delgado et al. 2018), palm planting (Mazur 2009, Passos et al. 2019), stored food (Hinton 1945), and forestry (bark beetles) (Camara et al. 2003, Shepherd and Goyer 2005).

Despite their importance to applied sciences, basic information (e.g., distribution, ecology, natural history and conservation) about Histerid beetles from the Neotropical region is scarce. By extension, their diversity in conservation areas is mostly unknown (Ganho and Marinoni 2005, Leivas

ZOOLOGIA
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Figure 1. *Oxysternus maximus* (Linnaeus, 1767), dorsal view. This is the largest species of Histerid beetles in the world, and of all the species known in Brazilian territory, this was the first to be described. Scale bar: 1.0 cm.

et al. 2013, Rafael et al. 2017) and it may be for this reason that they were never included/used in the management plans of Conservation Units (CUs) in Brazil.

Following traditional classification of Histerid beetles, all around the world, the family has 4,252 species belonging to 391 genera, 17 tribes and 11 subfamilies (Mazur 2011). In the Neotropical region, 139 genera and 1,047 species were registered (respectively 85 and 352 in Brazil) by Costa (2000) and among these, 120 genera and 449 species are registered for Brazil according to the global catalog of Mazur (2011).

Herein we present an overview about the Histerid beetles biodiversity from Brazil. More specifically, this contribution provides: i) the historical data and the current richness known for Brazil, including a list of species; ii) the main authors who described the Histeridae biodiversity recorded in Brazilian territory, and the most relevant entomological collections for studies with Brazilian fauna; iii) the literature to identify histerids registered in Brazilian territory; iv) the known biological associations and collection methods; v) and the conservation of Brazilian species; in addition, to proposing goals for future studies.

MATERIAL AND METHODS

The most recent taxonomic data presented herein are from the Taxonomic Catalog of the Brazilian Fauna – TCBF (in portuguese Catálogo Taxonômico da Fauna do Brasil – CTFB), a Brazilian initiative to generate knowledge about the biodiversity of the fauna present in the country (Boeger et al. 2024). More specifically, the data are from the Histeridae part from TCBF that are being performed by the authors since 2015 until December 31, 2023 (Bicho et al. 2023). Therefore, this publication is not intended to have precise and completely established data, but rather an approximation of the status of knowledge of the family's biodiversity in the country, as this data is constantly being updated.

During the construction of the catalog of Bicho et al. (2023), the first list of taxa was initially proposed based on the catalog by Mazur (2011) and updated with subsequent publications that are cited for each taxon on the TCBF homepage. At this moment, information from biological collections is not being considered, although in the future this information will be used to expand the TCBF database.

The data about the main literature that can be useful to identify Histeridae biodiversity recorded in Brazilian territory; information about known biological associations and collecting methods; and information about conservation of the Brazilian species come from literature, and part of this information is already available in the TCBF.

RESULTS AND DISCUSSION

Historical data and known current biodiversity from Brazil

A first list of Histeridae from Brazil was drawn up by Schmidt (1896) (Table 1), with a list rich in taxa, but still restricted to the Southeast region of the country, with species records

only for the state of Rio de Janeiro (Petrópolis, city of Rio de Janeiro, "Boa Sorta" [currently a municipality of Cantagalo, state of Rio de Janeiro] and "Santa Rita" [current location uncertain, but probably within the state of Rio de Janeiro].

Costa Lima (1952), in his classic work on the Brazil insects, described the general aspects of the family's morphology, presented their habits and some lifestyles. Finally, he addressed the classification, citing more than 3,000 species throughout the world and little more than 1,000 species for the Neotropical region.

The second compilation record on the fauna of Histeridae from Brazil was presented by the Brazilian Jacintho Guérin (1953). He mentioned a few species occurring in Brazil, in a lower richness than presented by Schmidt (1896), but pointing species distribution to the North, Center and South of the country (Table 1).

Table 1. History of knowledge of Histeridae biodiversity in Brazil, according to global, national and regional data. (NC) Not cited.

	Subfamilies	Tribes	Genera	Species
Schmidt (1896)	NC	NC	16	64
Costa Lima (1952)	NC	NC	6	4
Guérin (1953)	NC	NC	6	25
Blackwelder (1957)	8	5	85	352*
Costa (2000)	NC	NC	85	352
Mazur (2011)	7	12	120	449*
Casari and Ide (2012)	NC	NC	NC	352
Kovarik and Caterino (2016)	7	13	126	530
Gonçalves and Leivas (2017)	7	11	66	157
Leonel and Leivas (2023)	7	11	66	164
Casari et al. (2024)	NC	NC	NC	501
Present publication	6	16	132	607

*Not accounting for the species cited for "South America" which may be present in Brazil.

For six decades, these lists of Histeridae species occurring in Brazil were not updated, and just much time later from Guérin, a regional list of species occurring in southern Brazil (states of Paraná, Santa Catarina and Rio Grande do Sul) was published, being the first most complete regionalized list of species for the country (Gonçalves and Leivas 2017). Recently, information on taxa occurring in southern Brazil was substantially improved by the work of Leonel and Leivas (2023), in which authors presented an update of the regional list of Histeridae species, a list of species that are inside Conservation Units in southern Brazil, and biotic (biome, ecoregion and associations/affinities) and abiotic information (altitude and month of occurrence) for each species.

Other regional compilations for Histeridae, although on a smaller scale, were limited to the work of Leivas et al. (2013) (about the species present in the vegetation formation of "Campos Gerais" from Paraná), Rafael et al. (2017) (on species present in Conservation Units under the Caatinga biome) and Leivas et al. (2024) (the first list of Histeridae from the state of Acre, Brazil)

Our data (Bicho et al. 2023) indicate that so far six subfamilies, 16 tribes, 132 genera and 607 species and one subspecies are known to Brazil (Table 1). These numbers represent an increase of 37% of genera and 43% of species in relation to Costa (2000).

Checklist of known current biodiversity of Histeridae from Brazil

Recent works by Zhou et al. (2020) and Lackner et al. (2023) suggested considerable changes in the internal classification of Histeridae, bringing a modern proposal for definitions of subfamilies and tribes, which we adopt in the present publication.

The Appendix 1 presents the complete list of Histeridae taxa recorded in Brazil until December 31, 2023 (Bicho et al. 2023). The most diverse genera in Brazil are the Exosternini *Phelister* (69 spp.) and *Operclipygus* (57 spp.) but several genera are known by just one species.

Regarding knowledge of the biodiversity of Histeridae in Brazilian states, for all Brazilian states, excepted for Alagoas and Sergipe, there is at least one record of the family. The southern region of Brazil has better knowledge of species, with Paraná having the largest number of species recorded among the States, while the northeast has the greatest deficiency: Acre (16 spp., including data from Leivas et al. 2024), Amapá (9 spp.), Amazonas (18 spp.), Bahia (7 spp.), Ceará (3 spp.), Distrito Federal (5 spp.), Espírito Santo (7 spp.), Goiás (5 spp.), Maranhão (4 spp.), Mato Grosso (29 spp.), Mato Grosso do Sul (5 spp.), Minas Gerais (15 spp.), Pará (35 spp.), Paraíba (1 sp.), Paraná (75 spp.), Pernambuco (2 spp.), Piauí (2 spp.), Rio de Janeiro (17 spp.), Rio Grande do Norte (3 spp.), Rio Grande do Sul (10 spp.), Rondônia (7 spp.), Roraima (1 sp.), Santa Catarina (48 spp.), São Paulo (10 spp.) and Tocantins (3 spp.).

Authors of Brazilian taxa, Collections, and depositories of type specimens

Forty one authors contributed describing species that occur in Brazilian territory (Table 2). In the 19th century, the most significant researcher was the French Abbé S.-A. De Marseul (118 described species) contributing enormously

to the subfamilies Abraeinae (Acritini, Plegaderini and Trypanaeini), Dendrophilinae (Paromalini), Haeteriinae (Haeteriini), Histerinae (Exosternini, Histerini, Hololeptini and Omalodini) and Saprininae (Euspilotini and Saprinini).

During the transition from the 19th century to the 20th century, we highlight the British G. Lewis (83 described species) contributing to the description of many Abraeinae (Teretriini and Trypanaeini), Dendrophilinae (Bacaniini and Paromalini), Haeteriinae (Haeteriini and Nymphisterini), Histerinae (Exosternini, Hololeptini, Omalodini and Platysomatini), Saprininae (Euspilotini) and Tribalinae.

In the 20th century, A. Reichensperger made an important contribution (71 described species) to the myrmecophiles of Haeteriinae (Haeteriini and Nymphisterini) and a few Histerinae (Exosternini).

In the 21st century, Belarusian A.K. Tishechkin is in the spotlight (124 described species) from Haeteriinae (Haeteriini and Nymphisterini), Histerinae (Exosternini), this is the researcher who most described species of Histeridae occurring in Brazil; the North American M. S. Caterino (110 described species), mainly with Histerinae (Exosternini and Histerini); and the French N. Dégallier (33 described species) including Dendrophilinae, Haeteriinae (Haeteriini and Nymphisterini), Histerinae (Exosternini) and Tribalinae.

The contribution of Brazilian researchers to the description of the biodiversity of Histeridae started a little over 10 years ago, so it is still not very significant in the country, being represented by only three researchers: D.P. Moura (three species described) L.M. Almeida (two species described), and F.W.T. Leivas (two species described). It is worth noting that, before these researchers, no Brazilian taxonomists had ever dedicated themselves to studying the family.

As a reflection of the history of the contribution of foreign researchers (Marseul, Lewis, Reichensperger, Schmidt, Erichson and Bickhardt) much of the type material of Brazilian species of Histeridae is deposited in foreign collections (Horn et al. 1990), the main ones being: Muséum National d'Histoire Naturelle, Paris (MNHN), Museum für Naturkunde, Berlin (ZMHB) and British Museum, London (BMNH).

In relation with general collections around the world, a good representation of specimens of Neotropical Histeridae, including species known for Brazilian territory, can be accessed in the Histeridae collection at The Field Museum of Chicago (FMNH), being an important collection for any study on the Neotropical fauna of Histeridae. This collection was extensively worked on by R.L. Wenzel (1916–2006) (Gomy 2014a, 2014b), and is the largest Histeridae dataset in the world, with more than 40 thousand specimens (GBIF 2023).

Table 2. Authors who contributed to the description of the Histeridae biodiversity recorded in Brazilian territory according to Bicho et al. (2023) and the respective number of species described by each of them, in descending order. *Although the number of species described by each author is presented individually, many of them published new species in collaboration, such as many species described by Caterino, Dégallier and Tishechkin. This means that the sum of the number of species described by each author is much greater than the number of species recorded in Brazil.

Author	Number of described species*
Tishechkin	124
Marseul†	118
Caterino	110
Lewis†	83
Reichensperger†	71
Schmidt†	50
Dégallier	33
Erichson†	28
Bickhardt†	22
Wenzel†	13
Mazur	11
Bruch†	10
Hinton†	9
Desbordes†	7
Borgmeier†	6
Fabricius†	5
Helava	5
Paykull†	5
Arriagada	4
Dybas†	4
Gomy	4
Kovarik	3
Moura	3
Almeida	2
Dillon†	2
Lackner	2
Leivas	2
Mann†	2
Blanchard†	1
Boheman†	1
Fairmaire†	1
Kanaar†	1
Kirby†	1
Kirsch†	1
Le Conte†	1
Linnaeus†	1
Olivier†	1
Sahlberg†	1
Steinheil†	1
Tarsia in Curia†	1
Westwood†	1

† Deceased researcher.

Regarding Brazilian collections, few biological collections have their Histeridae collections digitized (Fig. 2), which makes it difficult to track the main repositories that serve as a basis for the development of any area of Biological Sciences. However, important collections of Histeridae can be found in the following regions of Brazil: North: Instituto Nacional de Pesquisas da Amazônia, Manaus (INPA) and Museu Paraense Emílio Goeldi, Belém (MPEG); Northeast: Coleção Entomológica da Universidade Federal Rural de Pernambuco, Recife (CERPE); Center West: Coleção Entomológica de Mato Grosso Eurides Furtado da Universidade Federal de Mato Grosso, Cuiabá (CEMT); Southeast: Museu de Zoologia da Universidade de São Paulo, São Paulo (MZUSP); South: Coleção Entomológica Pe. J.S. Moure, Universidade Federal do Paraná, Curitiba (DZUP), and the largest collection in Brazil Coleção Entomológica do Setor Palotina, Universidade Federal do Paraná, Palotina (CESP).

Taxonomic revisions and ability to identify species from Brazil

To date there is no identification key that covers the recognition of suprageneric taxa occurring in Brazil, and information for species identification is also scattered. In order to meet an immediate demand, we present in Appendix 2 a review of the main works that may be useful to identify Histeridae taxa occurring in Brazil. Clearly, when compared to Appendix 1, the urgent need for taxonomic revisions of the genera occurring in Brazil is noticeable, as only 45 genera (26% of the total) present some more robust taxonomic information to identify the species, although several of them need updating.

Biological associations known and collecting methods

The breadth of environments in which the Histeridae occur, as well as the wide geographical distribution of the family, demonstrate that they are very successful predatory carnivorous beetles.

Many groups are associated with nest or burrows of vertebrates, abandoned or live nests of several bird families, ant nests (~400 spp. of Abraeinae, Chlamydopsinae [not registered for Brazil], Dendrophilinae, Haeteriinae, Histerinae, Onthophilinae [not registered for Brazil], Saprininae, and Tribalinae), termites nest (decaying trees infested by termites, living colonies of arboreal and subterranean nests, abandoned nests, as well as detritus piles produced by termites), hymenopteran nests (stingless bees and Africanized honeybees) and egg sacs of theraphosid spiders (Dégallier et al. 2020, Iannuzzi et al. 2021).

However, we still know little about the affinities and ecological associations for the species in the family in Neotropical region. Leonel and Leivas (2023) presented the first compilation of ecological associations/affinities of species of Histeridae with animal carcass, association with dung, with live plants, decaying fruit, decaying tree, litter, ants and termites. Papers like this are important, as it is rich in information to feed the TCBF. A compilation of the Histeridae associated with the carcass can be accessed in Correa et al. (2020a, 2020b).

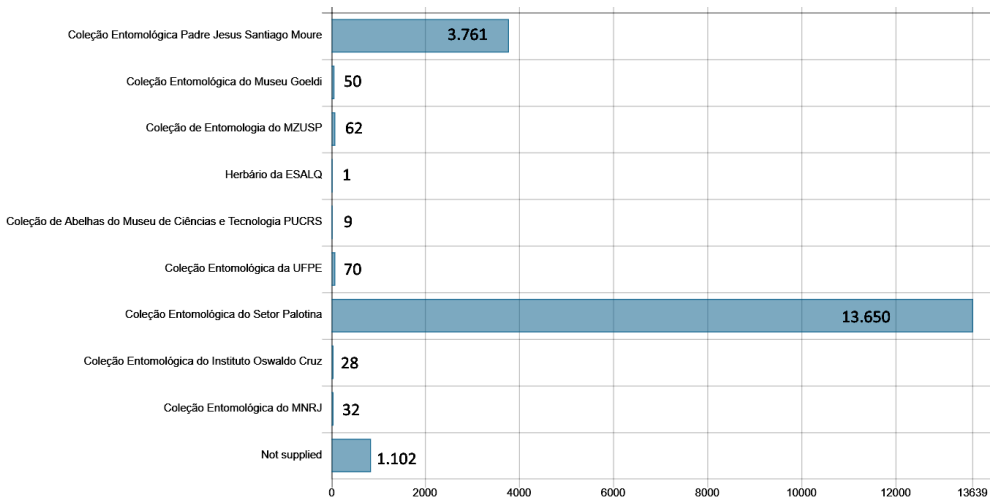

Figure 2. Number of Histeridae specimens (X axis) present in Brazilian collections (Y axis) that have their data digitized and available on the platform of Brazilian Biodiversity Information System (SIBBr 2023), accessed in December 2023.

Regarding how to collect Histeridae in the face of a large occupation of environments, several active and passive collection methods can be applied, and the choice of the most appropriate method is related to prior knowledge of the natural history of the group. Certainly, for ecological work, pitfall and flight interception traps can be great choices, as they allow the capture of specific groups (in the case of pitfall, depending on the attraction used) or a random diversity of species that are in flight (in the case of FIT), in addition to both enabling standardized field efforts. More information about these capture methods can be accessed in the specialized works of Dégallier and Gomy (1983), Dégallier (2004), Leivas et al. (2013) and Iannuzzi et al. (2021), or general bibliography of Almeida et al. (1998, 2012, 2024).

About the conservation of the species

Sixty-two species (10.1% of the total) are recorded inside of CUS from Brazil: Área Especial de Interesse Turístico (AEIT) do Marumbi was the richest (25 spp.), followed by Parque Estadual de Guartelá (14 spp.), Parque Nacional de Sete Cidades (11 spp.), Parque Estadual Mata São Francisco (10 spp.), Area de Proteção Ambiental Serra de São José (9 spp.), Refúgio da Vida Silvestre Libélulas da Serra de São José (9 spp.), Adolpho Ducke Forest Reserve (5 spp.), Reserva Particular de Patrimônio Natural Fazenda (5 spp.), Almas Parque Nacional de Ubajara (4 spp.). Estação Ecológica da Ilha do Mel, Parque Ecológico Samuel Klabin, Parque Estadual do Marumbi, Parque Estadual Turvo, Parque Mata dos Godoy, Reserva Estadual de Vila Rica and Parque da Ferradura had only one species (Appendix 3).

Until now, the Atlantic Forest is the best-known biome (148 spp., Vieira et al. 2018, Leonel and Leivas 2023), followed by Cerrado (21 spp., Vieira et al. 2018, Leonel and Leivas 2023), Amazon (17 spp., Mise et al. 2010, Leivas et al. 2012, Leivas et al. 2024), Caatinga (16 species, Santos et al. 2014, Rafaela et al. 2017), Pampa (14 spp., Leonel and Leivas 2023) and no information for Pantanal (Fig. 3). Certainly, this is not the real diversity of Histeridae present in Brazilian biomes. The underestimated data, especially for the Pantanal, Cerrado, Caatinga and Amazon, reflects the lack of compilation of regional data.

Future studies

To improve the family's knowledge in Brazil, on aspects of taxonomy, ecology and conservation, we suggest some goals for the next five years:

Development of taxonomic tools, such as identification keys, to improve the ability to recognize Brazilian biodiversity and define new taxa to be described. These tools should be designed for use not only by histeridologists, but also by other entomologists who can advance in other areas of study without dependence on a specialist taxonomist;

Development of taxonomic revisions to improve the ability to identify species and describe new species. An inestimable number of Histeridae species are yet to be described, and only taxonomic revisions can provide certainty for these descriptions;

Preparation of regional lists of species rich in information on geographic distribution, biotic (biome, ecoregion and associations/affinities) and abiotic information (altitude and month of occurrence) for each species. These lists are essential for updating and expanding information in the Brazilian Fauna Taxonomic Catalog, help to improve knowledge of biodiversity in Brazilian biomes and it is fundamental for discussing aspects of species conservation. Studies should be prioritized for regions with little information on the Histeridae fauna, such as the north, northeast and central-west regions of Brazil. Regarding biomes, priority must be given to studies aimed at improving knowledge in the Pantanal, Amazon, Pampa and Caatinga;

Preparation of lists of species that are inside Conservation Units in Brazil. This information is essential to understand the effectiveness of Conservation Units in conserving biodiversity, and in addition to being a fundamental source for improving CUs management plans and developing public conservation policies;

Improve the digitization of Histeridae collections in Brazil and provide free access. The open access of data from scientific collections is essential for the development of research in the most diverse areas of science, from basic to applied. Using this data, we were able to define the best groups for taxonomic revisions, even understanding the geographic distribution of a taxon of applied importance;

Training of human resources specialized in taxonomy, ecology, and conservation of Histeridae. Historically, Brazil is a country lacking people interested in research on Histeridae, which makes it difficult to advance knowledge in the national territory. We also understand that, as important as training human resources, is the placement of these future researchers in public or private research institutions.

This is the first most complete scenario of information on the Histeridae family in Brazil. We believe that the first two items of this list (to develop taxonomic tools for identification of the biodiversity and to perform taxonomic revisions) are priority research with the family. We hope that in five years we will make important progress in the objectives listed above.

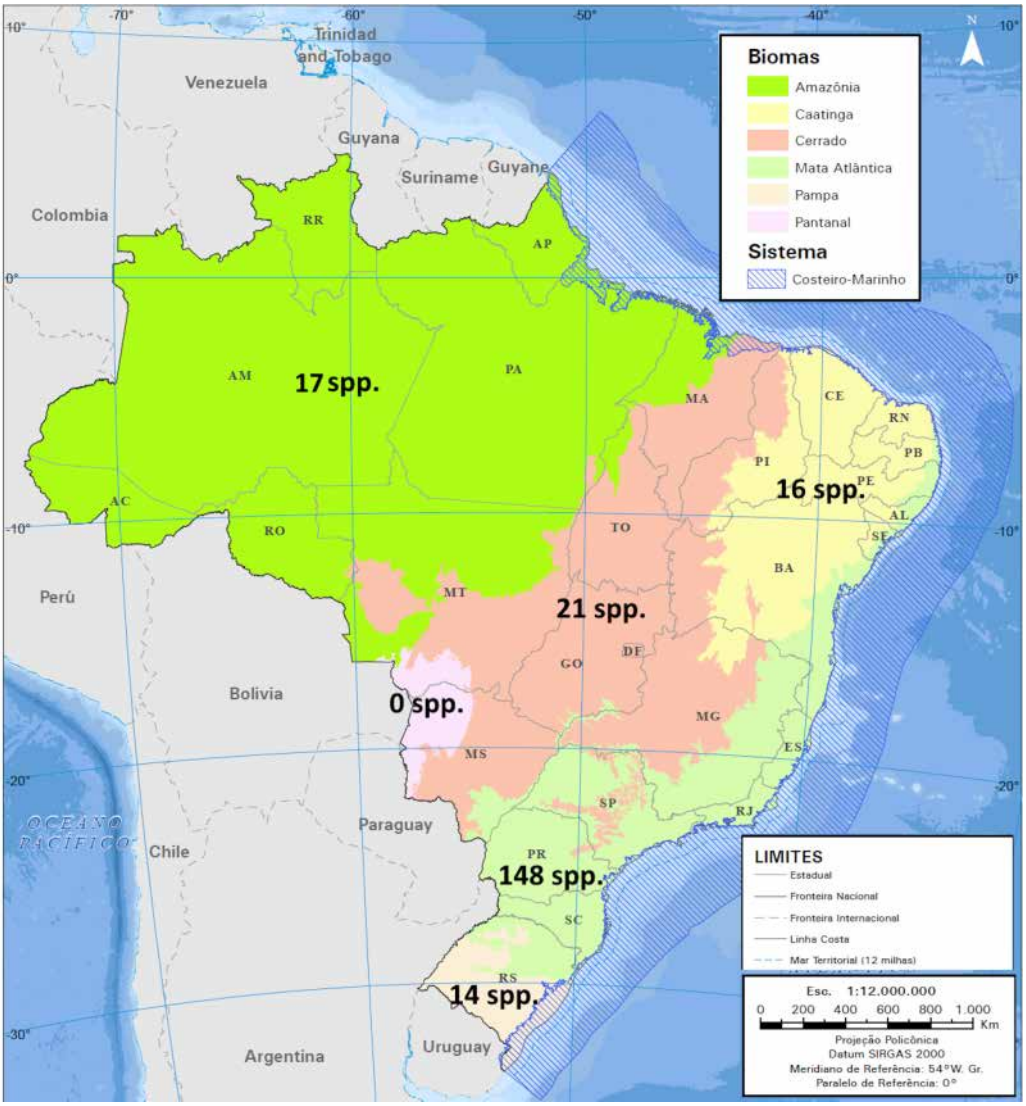

Figure 3. Species richness of Histeridae species known in Brazilian biomes. Adapted from IBGE (2024).

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Appendix 1. Complete list of Histeridae taxa recorded in Brazil until December 31, 2023, according to Bicho et al. (2023).

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
Abraeinae	Acritini	*Acritus*	*Acritus*	*tuberculatus*	Wenzel and Dybas, 1941
		Acritus	*Pycnacritus*	*acinus*	Marseul, 1853
		Aeletes	*Acritinus*	*floridae*	(Marseul, 1853)
		Aeletes	*Aeletes*	*nicolasi*	Leivas, 2012
		Aeletes	*Aeletes*	*troglodytes*	Wenzel, 1944
	Plegaderini	*Plegaderus*	*Plegaderus*	*monachus*[a]	Marseul, 1870
	Teretriini	*Teretriosoma*		*festivum*	Lewis, 1879
		Teretriosoma		*prasinum*	Lewis, 1902
		Teretriosoma		*unicorne*	Lewis, 1901
		Teretriosoma		*viridicatum*	Lewis, 1891
		Teretrius	*Neotepetrius*	*cingulus*	(Lewis, 1891)
		Teretrius	*Neotepetrius*	*gigas*	(Mazur, 1978)
		Teretrius	*Neotepetrius*	*grouvellei*	(Lewis, 1891)
		Teretrius	*Neotepetrius*	*tuberculifrons*	(Mazur, 1978)
		Trypolister		*capucinus*	Bickhardt, 1916
	Trypanaeini	*Coptotrophis*		*bellator*	(Lewis, 1902)
		Coptotrophis		*deyrolii*	(Marseul, 1856)
		Coptotrophis		*ornatulus*	Lewis, 1910
		Coptotrophis		*proboscideus*	(Fabricius, 1801)
		Coptotrophis		*resectus*	(Marseul, 1870)
		Trypanaeus		*bipustulatus*	(Fabricius, 1801)
		Trypanaeus		*bisulcifrons*	Marseul, 1870
		Trypanaeus		*carinirostris*	Marseul, 1856
		Trypanaeus		*catharinensis*	Bickhardt, 1920
		Trypanaeus		*cornifrons*	Lewis, 1902
		Trypanaeus		*ensifer*	Marseul, 1856
		Trypanaeus		*fasciatus*	Lewis, 1891
		Trypanaeus		*montivagus*	Lewis, 1888
		Trypanaeus		*nasicornis*	Marseul, 1870
		Trypanaeus		*obesus*	Schmidt, 1896
		Trypanaeus		*petropolitanus*	Schmidt, 1896
		Trypanaeus		*plagiatus*	Lewis, 1891
		Trypanaeus		*quadricollis*	Marseul, 1856
		Trypanaeus		*singularis*	Lewis, 1894
		Trypanaeus		*spinifrons*	Bickhardt, 1916
		Trypanaeus		*sulcipygus*	Marseul, 1870
		Trypanaeus		*thoracicus*	(Fabricius, 1801)
		Trypanaeus		*torpedo*	Lewis, 1885
		Trypanaeus		*volvulus*	Erichson, 1834
		Xylonaeus		*aculeatus*	(Lewis, 1888)
		Xylonaeus		*breviatus*	Bickhardt, 1916
		Xylonaeus		*fallax*	(Marseul, 1856)
		Xylonaeus		*forficula*	Bickhardt, 1916
		Xylonaeus		*immarginatus*	Lewis, 1902
		Xylonaeus		*lamellicauda*	Bickhardt, 1916
		Xylonaeus		*praecutus*	Bickhardt, 1916
Dendrophilinae	Bacaniini	*Bacanius*	*Bacanius*	*convergens*	Schmidt, 1896
		Bacanius	*Bacanius*	*scalptus*	Lewis, 1888
		Dégallierister		*antoinei*	Dégallier, 2015
		Dégallierister		*bihamatus*	Dégallier, 2015
		Dégallierister		*diquis*	Gomy, 2013
		Dégallierister		*greglamarrei*	Dégallier, 2015
		Dégallierister		*hielkemai*	Gomy, 2013
		Dégallierister		*kanaari*	Dégallier, 2015
		Dégallierister		*parenthesis*	Dégallier, 2015
		Dégallierister		*penatii*[b]	Gomy, 2001
		Dégallierister		*rogeri*	Dégallier, 2015
		Dégallierister		*surinamensis*	Gomy, 2013
		Mullerister		*striatinotum*	(Wenzel, 1944)
	Paromalini	*Carcinops*	*Carcinops*	*biinterrupta*	Wenzel and Dybas, 1941
		Carcinops	*Carcinops*	*carinata*	Wenzel and Dybas, 1941
		Carcinops	*Carcinops*	*exigua*	Wenzel, 1944

Continues

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
		Carcinops	*Carcinops*	*misella*	Marseul, 1855
		Carcinops	*Carcinops*	*miserula*	Marseul, 1862
		Carcinops	*Carcinops*	*ovatula*	Lewis, 1888
		Carcinops	*Carcinops*	*plaumanni*	Wenzel, 1944
		Carcinops	*Carcinops*	*pumilio*	(Erichson, 1834)
		Carcinops	*Carcinops*	*tenella*	(Erichson, 1834)
		Carcinops	*Carcinops*	*tristicula*	Marseul, 1870
		Carcinops	*Carcinops*	*troglodytes*	(Paykull, 1811)
		Carcinops	*Carcinops*	*tuberata*	Wenzel, 1944
		Paromalus	*Isolomalus*	*addendus*	Schmidt, 1896
		Paromalus	*Isolomalus*	*bicinctus*	Marseul, 1870
		Paromalus	*Isolomalus*	*causticus*	Marseul, 1862
		Paromalus	*Isolomalus*	*concentricus*	Marseul, 1870
		Paromalus	*Isolomalus*	*elongatus*	(Lewis, 1907)
		Paromalus	*Isolomalus*	*inunctus*	Marseul, 1862
		Paromalus	*Isolomalus*	*irregularis*	Schmidt, 1896
		Paromalus	*Isolomalus*	*jejunus*	Lewis, 1888
		Paromalus	*Isolomalus*	*leleupi*	Wenzel, 1976
		Paromalus	*Isolomalus*	*productus*	Marseul, 1855
		Paromalus	*Isolomalus*	*rugigenius*	Marseul, 1870
		Paromalus	*Isolomalus*	*samba*	Mazur, 1981
		Paromalus	*Isolomalus*	*sincerus*	Lewis, 1888
		Paromalus	*Isolomalus*	*trifolium*	Marseul, 1862
		Paromalus	*Isolomalus*	*truncatus*	(Lewis, 1907)
		Xestipyge		*garbigliettii*	(Marseul, 1867)
Haeteriinae	Haeteriini	*Alienister*		*patruum*	Reichensperger, 1926
		Aristomorphus		*borgmeieri*	(Reichensperger, 1931)
		Aristomorphus		*latipes*	(Reichensperger, 1933)
		Aristomorphus		*ogloblini*	(Bruch, 1933)
		Aristomorphus		*perversus*	(Reichensperger, 1923)
		Aristomorphus		*rutilus*	Lewis, 1913
		Brasilister		*flechtmanni*	Dégallier, 1998
		Chelonarhister		*castroi*	Dégallier, 2004
		Chelyocephalus		*varicolor*	Schmidt, 1983
		Coelister		*cavernosus*	(Schmidt, 1889)
		Colonides		*collegii*	(Reichensperger, 1923)
		Colonides		*hubrichi*	Bruch, 1923
		Colonides		*quadriglumis*	(Reichensperger, 1923)
		Cossyphodister		*schwarzmaieri*	Reichensperger, 1936
		Discoscelis		*canaliculata*	Schmidt, 1889
		Enicosoma		*vespertinum*	Lewis, 1904
		Euclasea		*diadocha*	(Reichensperger, 1939)
		Euclasea		*novaeteutoniae*	(Reichensperger, 1939)
		Euclasea		*pumila*	(Reichensperger, 1926)
		Euclasea		*splendens*	Reichensperger, 1939
		Euclasea		*tuberculata*	Lewis, 1893
		Euxenister		*asperatus*	Reichensperger, 1923
		Euxenister		*caroli*	Reichensperger, 1923
		Fistulaster		*hamata*	Helava, 1985
		Hemicolonides		*parvulus*	(Lewis, 1891)
		Hemicolonides		*plaumanni*	Reichensperger, 1939
		Hesperodromus		*sodalis*	Schmidt, 1889
		Hetaeriobius		*borgmeieri*	(Reichensperger, 1925)
		Hetaeriobius		*bucki*	Reichensperger, 1925
		Hippeutister		*plaumanni*	Reichensperger, 1936
		Homalopygus		*amnicola*	Lewis, 1898
		Homalopygus		*commensalis*	Lewis, 1885
		Homalopygus		*fidelis*	Reichensperger, 1931
		Homalopygus		*geminatus*	Lewis, 1901
		Homalopygus		*iniquus*	Bickhardt, 1914
		Homalopygus		*latipes*	Boheman, 1858
		Homalopygus		*latisternus*	Lewis, 1907
		Homalopygus		*longipes*	Marseul, 1870
		Iugulister		*clarissae*	Reichensperger, 1958

Continues

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
		Kleptisister		*hirsuta*	Helava, 1985
		Lissosternus		*gracilipes*	Lewis, 1905
		Neoterapus		*bicarinatus*	(Lewis, 1898)
		Nomadister		*papillatus*	Borgmeier, 1948
		Notocoelis		*satur*	Lewis, 1900
		Paratroterius		*effertus*	(Lewis, 1891)
		Parodites		*wassmanni*	Reichensperger, 1923
		Paroecister		*zikani*	Reichensperger, 1923
		Plagioscelis		*daedalus*	(Lewis, 1891)
		Plagioscelis		*striaticeps*	(Bruch, 1929)
		Plaumannister		*volitans*	Reichensperger, 1958
		Procolonides		*bruchi*	Reichensperger, 1935
		Pterotister		*schwarzmaieri*	Borgmeier, 1948
		Reninoides		*declinatus*	Helava, 1985
		Reninopsis		*reichenspergeri*	Helava, 1985
		Reninus		*arechavaletae*	(Marseul, 1870)
		Reninus		*bruchi*	Reichensperger, 1927
		Reninus		*curvatus*	(Lewis, 1912)
		Reninus		*meticulosus*	(Lewis, 1885)
		Reninus		*pygidialis*	(Reichensperger, 1926)
		Reninus		*seminitens*	Schmidt, 1893
		Reninus		*turritus*	Lewis, 1902
		Scapicoelis		*ayri*	Dégallier and Tishechkin, 2022
		Scapicoelis		*bichoae*	Dégallier and Tishechkin, 2022
		Scapicoelis		*gomyi*	Dégallier and Tishechkin, 2022
		Scapicoelis		*kinitii*	Dégallier and Tishechkin, 2022
		Scapicoelis		*koriphensis*	Dégallier and Tishechkin, 2022
		Scapicoelis		*lackneri*	Dégallier and Tishechkin, 2022
		Scapicoelis		*leivasi*	Dégallier and Tishechkin, 2022
		Scapicoelis		*mariaeclarae*	Dégallier and Tishechkin, 2022
		Scapicoelis		*marinae*	Dégallier and Tishechkin, 2022
		Scapicoelis		*minor*	Dégallier and Tishechkin, 2022
		Scapicoelis		*nazareti*	Dégallier and Tishechkin, 2022
		Scapicoelis		*parallelus*	Dégallier and Tishechkin, 2022
		Scapicoelis		*rolandi*	Dégallier and Tishechkin, 2022
		Scapicoelis		*tibialis*	Marseul, 1862
		Scapicoelis		*tuberculatus*	Dégallier and Tishechkin, 2022
		Scapolister		*sternalis*	Borgmeier, 1930
		Synetister		*pilosus*	Reichensperger, 1924
		Synoditinus		*herteli*	Reichensperger, 1929
		Terapus		*fletchmanni*	Dégallier, 1998
		Terapus		*marseuli*	Westwood, 1874
		Terapus		*mirificus*	Reichensperger, 1926
		Terapus		*scaphipes*	Reichensperger, 1933
		Teratosoma		*longipes*	Lewis, 1885
		Termitoxenus		*setaceus*	Schmidt, 1889
		Termitoxenus		*strigicollis*	Lewis, 1898
		Thaumataerius		*emersoni*	Mann, 1923
		Troglosternus		*dasypus*	Bickhardt, 1917
		Troglosternus		*ecitonis*	Mann, 1925
		Troglosternus		*horridus*	Dégallier and Tishechkin, 2017
		Troglosternus		*rugosus*	Dégallier and Tishechkin, 2018
		Tubulister		*curvipilosus*	Borgmeier, 1948
		Tylois		*mirificus*	Lewis, 1901
		Ulkeus		*gratianus*	(Bruch, 1926)
		Ulkeus		*sahlbergi*	(Schmidt, 1893)
		Xenister		*schwarzmaieri*	Borgmeier, 1929
	Nymphisterini	*Aemulister*		*borgmeieri*	Reichensperger, 1938
		Alienodites		*amazonicus*	Tishechkin, 2007
		Alloiodites		*dispar*	(Reichensperger, 1939)
		Alloiodites		*plaumanni*	(Reichensperger, 1939)
		Alloiodites		*regulus*	(Reichensperger, 1939)
		Amiculus		*major*	(Dégallier, 1994)
		Anasynodites		*striatus*	(Reichensperger, 1923)

Continues

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
		Aristonister		*sericeus*	(Borgmeier, 1948)
		Bruchodites		*praeclusus*	(Reichensperger, 1939)
		Cheilister		*lucidulus*	Reichensperger, 1924
		Chrysetaerius		*iheringi*	Reichensperger, 1923
		Daitrosister		*ecitonis*	(Bruch, 1923)
		Daitrosister		*irregularis*	(Reichensperger, 1938)
		Daitrosister		*longipilus*	(Reichensperger, 1931)
		Daitrosister		*setulosus*	(Reichensperger, 1923)
		Ecclisister		*bickhardti*	(Reichensperger, 1923)
		Guianahister		*ashei*	Tishechkin, 2007
		Helavadites		*elegantulus*	(Reichensperger, 1939)
		Helavadites		*gibbidorsum*	(Schmidt, 1893)
		Leptosister		*patulus*	Helava, 1985
		Mesynodites		*aciculatus*	Schmidt, 1893
		Mesynodites		*affinis*	Reichensperger, 1931
		Mesynodites		*attaphilus*	Bruch, 1933
		Mesynodites		*Dégallieri*	Tishechkin, 2007
		Mesynodites		*intermedius*	Reichensperger, 1933
		Mesynodites		*schuppii*	(Schmidt, 1893)
		Mesynodites		*sodalis*	(Reichensperger, 1924)
		Mesynodites		*virgatus*	Reichensperger, 1931
		Metasynodites		*legionarius*	(Reichensperger, 1929)
		Metasynodites		*minor*	Reichensperger, 1931
		Metasynodites		*paschalis*	(Reichensperger, 1930)
		Microsynodites		*schmidti*	(Lewis, 1893)
		Microsynodites		*strigilatus*	(Reichensperger, 1931)
		Monotonodites		*nitidus*	(Reichensperger, 1923)
		Monotonodites		*pauperellus*	(Lewis, 1893)
		Monotonodites		*raptantis*	(Reichensperger, 1925)
		Mutodites		*evanescens*	(Reichensperger, 1935)
		Mutodites		*megacantha*	(Reichensperger, 1938)
		Mutodites		*semistriatus*	(Bruch, 1933)
		Mutodites		*speculum*	(Reichensperger, 1931)
		Nicolasites		*graniformis*	(Schmidt, 1893)
		Nymphister		*unstriatus*	Dégallier and Tishechkin, 2021
		Panoplitellus		*comes*	Reichensperger, 1923
		Paratropinus		*variepunctatus*	Reichensperger, 1923
		Psalidister		*burchelli*	Reichensperger, 1924
		Psalidister		*carinulatus*	Reichensperger, 1924
		Psalidister		*foveatus*	Reichensperger, 1924
		Psalidister		*furcatus*	Reichensperger, 1924
		Psalidister		*quadriglumis*	Reichensperger, 1924
		Reichenspergerites		*robustus*	(Reichensperger, 1939)
		Sternocoelopsis		*auricomus*	Reichensperger, 1923
		Sternocoelopsis		*veselyi*	Reichensperger, 1923
		Symphilister		*collegianus*	Reichensperger, 1923
		Symphilister		*hamati*	Reichensperger, 1929
		Trichoreninus		*exclamationis*	(Reichensperger, 1931)
		Trichoreninus		*imbricatus*	Lewis, 1893
		Trichoreninus		*major*	Bruch, 1923
		Trichoreninus		*schwarzmaieri*	(Reichensperger, 1931)
	Synoditulini	*Synoditulus*		*separatus*	(Reichensperger, 1924)
		Synoditulus		*separatus praedator*	Reichensperger, 1939
Histerinae	Exosternini	*Baconia*		*angulifrons*	Caterino and Tishechkin, 2013
		Baconia		*angusta*	Schmidt, 1893
		Baconia		*anthracina*	Caterino and Tishechkin, 2013
		Baconia		*atricolor*	Caterino and Tishechkin, 2013
		Baconia		*brunnea*	Caterino and Tishechkin, 2013
		Baconia		*bullifrons*	Caterino and Tishechkin, 2013
		Baconia		*burmeisteri*	(Marseul, 1870)
		Baconia		*castanea*	Caterino and Tishechkin, 2013
		Baconia		*cavifrons*	(Lewis, 1893)
		Baconia		*choaspites*	Lewis, 1901
		Baconia		*clemens*	Caterino and Tishechkin, 2013

Continues

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
		Baconia		*coerulea*	(Bickhardt, 1917)
		Baconia		*crassa*	Caterino and Tishechkin, 2013
		Baconia		*diminua*	Caterino and Tishechkin, 2013
		Baconia		*dives*	(Marseul, 1862)
		Baconia		*emarginata*	Caterino and Tishechkin, 2013
		Baconia		*famelica*	Caterino and Tishechkin, 2013
		Baconia		*festiva*	(Lewis, 1891)
		Baconia		*foliosoma*	Caterino and Tishechkin, 2013
		Baconia		*fortis*	Caterino and Tishechkin, 2013
		Baconia		*fulgida*	(Schmidt, 1889)
		Baconia		*gibbifer*	Caterino and Tishechkin, 2013
		Baconia		*gounellei*	(Marseul, 1887)
		Baconia		*grossii*	Caterino and Tishechkin, 2013
		Baconia		*guartela*	Caterino and Tishechkin, 2013
		Baconia		*haeterioides*	Caterino and Tishechkin, 2013
		Baconia		*illustris*	(Lewis, 1900)
		Baconia		*jubaris*	(Lewis, 1901)
		Baconia		*leivasi*	Caterino and Tishechkin, 2013
		Baconia		*lewisi*	Mazur, 1984
		Baconia		*longipes*	Caterino and Tishechkin, 2013
		Baconia		*loricata*	Lewis, 1885
		Baconia		*micans*	(Schmidt, 1889)
		Baconia		*obsoleta*	Caterino and Tishechkin, 2013
		Baconia		*patula*	Lewis, 1885
		Baconia		*piluliformis*	Caterino and Tishechkin, 2013
		Baconia		*plebeia*	Caterino and Tishechkin, 2013
		Baconia		*punctiventer*	Caterino and Tishechkin, 2013
		Baconia		*redemptor*	Caterino and Tishechkin, 2013
		Baconia		*repens*	Caterino and Tishechkin, 2013
		Baconia		*riouka*	(Marseul, 1861)
		Baconia		*salobrus*	(Marseul, 1887)
		Baconia		*slipinskii*	Mazur, 1981
		Baconia		*tenuipes*	Caterino and Tishechkin, 2013
		Baconia		*varicolor*	(Marseul, 1887)
		Baconia		*viridimicans*	(Schmidt, 1893)
		Chapischema		*doppelganger*	Caterino and Tishechkin, 2014
		Conchita		*propygidiale*	Mazur, 1994
		Conocassis		*invaginata*	Caterino and Tishechkin, 2014
		Conocassis		*minor*	Caterino and Tishechkin, 2014
		Crenulister		*dentatus*	Caterino and Tishechkin, 2014
		Crenulister		*grossus*	Caterino and Tishechkin, 2014
		Crenulister		*impar*	Caterino and Tishechkin, 2014
		Crenulister		*paucitans*	Caterino and Tishechkin, 2014
		Crenulister		*umbrosus*	Caterino and Tishechkin, 2014
		Hypobletus		*apicalis*	(Hinton, 1935)
		Hypobletus		*delicatus*	(Lewis, 1889)
		Hypobletus		*orbatus*	Schmidt, 1896
		Hypobletus		*parensis*	(Marseul, 1861)
		Hypobletus		*semirufus*	(Lewis, 1891)
		Hypobletus		*subridens*	(Marseul, 1870)
		Kaszabister		*barrigai*	Dégallier, Mazur, Tishechkin and Caterino, 2012
		Kaszabister		*ferrugineus*	(Kirsch, 1873)
		Kaszabister		*rubellus*	(Erichson, 1834)
		Lacrimorpha		*acuminata*	Caterino and Tishechkin, 2014
		Lacrimorpha		*balbina*	Caterino and Tishechkin, 2014
		Lacrimorpha		*glabra*	Caterino and Tishechkin, 2014
		Mecistostethus		*flechtmanni*	Caterino, Tishechkin and Dégallier, 2012
		Mecistostethus		*loretoensis*	Bruch, 1932
		Megalocraerus		*rubricatus*	Lewis, 1902
		Nunbergia		*exosternoides*	Mazur, 1978
		Operclipygus		*abbreviatus*	Caterino and Tishechkin, 2013
		Operclipygus		*angustisternus*	(Wenzel, 1944)
		Operclipygus		*arnaudi*	Dégallier, 1982

Continues

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
		Operclipygus		*atlanticus*	Caterino and Tishechkin, 2013
		Operclipygus		*belemensis*	Caterino and Tishechkin, 2013
		Operclipygus		*bidessois*	(Marseul, 1889)
		Operclipygus		*britannicus*	Caterino and Tishechkin, 2013
		Operclipygus		*callifrons*	Caterino and Tishechkin, 2013
		Operclipygus		*carinisternus*	Caterino and Tishechkin, 2013
		Operclipygus		*carinistrius*	(Lewis, 1908)
		Operclipygus		*cavisternus*	Caterino and Tishechkin, 2013
		Operclipygus		*conquisitus*	(Lewis, 1902)
		Operclipygus		*crenulatus*	Caterino and Tishechkin, 2013
		Operclipygus		*depressus*	(Hinton, 1935)
		Operclipygus		*distinctus*	(Hinton, 1935)
		Operclipygus		*distractus*	(Schmidt, 1896)
		Operclipygus		*dubitabilis*	(Marseul, 1889)
		Operclipygus		*farctissimus*	Caterino and Tishechkin, 2013
		Operclipygus		*farctus*	(Marseul, 1864)
		Operclipygus		*florifaunensis*	Caterino and Tishechkin, 2013
		Operclipygus		*formicatus*	Caterino and Tishechkin, 2013
		Operclipygus		*fossipygus*	(Wenzel, 1944)
		Operclipygus		*friburgius*	(Marseul, 1864)
		Operclipygus		*hospes*	(Lewis, 1902)
		Operclipygus		*iheringi*	(Bickhardt, 1917)
		Operclipygus		*impositus*	Caterino and Tishechkin, 2013
		Operclipygus		*impuncticollis*	(Hinton, 1935)
		Operclipygus		*incisus*	Caterino and Tishechkin, 2013
		Operclipygus		*inflatus*	Caterino and Tishechkin, 2013
		Operclipygus		*innocuus*	Caterino and Tishechkin, 2013
		Operclipygus		*juninensis*	Caterino and Tishechkin, 2013
		Operclipygus		*kerga*	(Marseul, 1870)
		Operclipygus		*latemarginatus*	(Bickhardt, 1920)
		Operclipygus		*minutus*	Caterino and Tishechkin, 2013
		Operclipygus		*mutuca*	Caterino and Tishechkin. 2013
		Operclipygus		*novateutoniae*	Caterino and Tishechkin, 2013
		Operclipygus		*parallelus*	Caterino and Tishechkin, 2013
		Operclipygus		*parensis*	Caterino and Tishechkin, 2013
		Operclipygus		*perplexus*	Caterino and Tishechkin, 2013
		Operclipygus		*plaumanni*	Caterino and Tishechkin, 2013
		Operclipygus		*plicatus*	(Hinton, 1935)
		Operclipygus		*plicicollis*	(Schmidt, 1893)
		Operclipygus		*profundipygus*	Caterino and Tishechkin, 2013
		Operclipygus		*prolixus*	Caterino and Tishechkin, 2013
		Operclipygus		*prominens*	Caterino and Tishechkin, 2013
		Operclipygus		*proximus*	Caterino and Tishechkin, 2013
		Operclipygus		*punctifrons*	Caterino and Tishechkin, 2013
		Operclipygus		*punctulatus*	Caterino and Tishechkin, 2013
		Operclipygus		*quadratus*	Caterino and Tishechkin, 2013
		Operclipygus		*sejunctus*	(Schmidt, 1896)
		Operclipygus		*siluriformis*	Caterino and Tishechkin, 2013
		Operclipygus		*simplistrius*	Caterino and Tishechkin, 2013
		Operclipygus		*subrufus*	Caterino and Tishechkin, 2013
		Operclipygus		*subsphaericus*	Caterino and Tishechkin, 2013
		Operclipygus		*subterraneus*	Caterino and Tishechkin, 2013
		Operclipygus		*sulcistrius*	Marseul, 1870
		Operclipygus		*tenuis*	Caterino and Tishechkin, 2013
		Pachycraerus[c]		*chabrillaci*	Marseul, 1861
		Paratropus	*Paratropus*	*anthracinus*[d]	Lewis, 1891
		Phelister		*almeidae*	Caterino and Tishechkin, 2020
		Phelister		*amazoniae*	(Lewis, 1898)
		Phelister		*amplistrius*	Schmidt, 1893
		Phelister		*annulatus*	Caterino and Tishechkin, 2020
		Phelister		*arcuatus*	Caterino and Tishechkin, 2020
		Phelister		*asperatus*	Caterino and Tishechkin, 2020
		Phelister		*bistriatus*	Hinton, 1935
		Phelister		*blairi*	Hinton, 1935

Continues

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
		Phelister		*blairoides*	Caterino and Tishechkin, 2020
		Phelister		*brevistrius*	Marseul, 1853
		Phelister		*canalis*	Lewis, 1888
		Phelister		*carinifrons*	Schmidt, 1893
		Phelister		*carinistrius*	Lewis, 1908
		Phelister		*carinullus*	Hinton, 1935
		Phelister		*chapadae*	Lewis, 1900
		Phelister		*chicomendesi*	Caterino and Tishechkin, 2020
		Phelister		*curvipes*	Caterino and Tishechkin, 2020
		Phelister		*daugar*	Marseul, 1861
		Phelister		*egincola*	Marseul, 1889
		Phelister		*erwini*	Caterino and Tishechkin, 2020
		Phelister		*fimbriatus*	Caterino and Tishechkin, 2020
		Phelister		*finitimus*	Bickhardt, 1918
		Phelister		*foveicollis*	Lewis, 1902
		Phelister		*fractistrius*	Lewis, 1908
		Phelister		*fraternus*	Caterino and Tishechkin, 2020
		Phelister		*fulvulus*	Marseul, 1870
		Phelister		*geijskesi*	Kanaar, 1997
		Phelister		*geminus*	Caterino and Tishechkin, 2020
		Phelister		*globosus*	Caterino and Tishechkin, 2020
		Phelister		*gracilis*	Schmidt, 1889
		Phelister		*gregarius*	Caterino and Tishechkin, 2020
		Phelister		*haemorrhous*	Marseul, 1854
		Phelister		*interrogans*	Marseul, 1889
		Phelister		*matatlantica*	Caterino and Tishechkin, 2020
		Phelister		*microdens*	Caterino and Tishechkin, 2020
		Phelister		*miscellus*	Caterino and Tishechkin, 2020
		Phelister		*morbidus*	Caterino and Tishechkin, 2020
		Phelister		*nidicola*	Bickhardt, 1920
		Phelister		*notandus*	Schmidt, 1893
		Phelister		*panamensis*	J.E. Le Conte, 1860
		Phelister		*parana*	Caterino and Tishechkin, 2020
		Phelister		*parecis*	Caterino and Tishechkin, 2019
		Phelister		*pervagatus*	Caterino and Tishechkin, 2020
		Phelister		*petro*	Bickhardt, 1917
		Phelister		*praedatoris*	Reichensperger, 1939
		Phelister		*praesignis*	Caterino and Tishechkin, 2020
		Phelister		*pumilus*	(Erichson, 1834)
		Phelister		*puncticollis*	Hinton, 1935
		Phelister		*pusio*	(Erichson, 1847)
		Phelister		*pygmaeus*	Bickhardt, 1918
		Phelister		*rectisternus*	Lewis, 1908
		Phelister		*rio*	Caterino and Tishechkin, 2020
		Phelister		*rouzeti*	(Fairmaire, 1850)
		Phelister		*rubicundus*	Marseul, 1889
		Phelister		*rufinotus*	Marseul, 1861
		Phelister		*ruptistrius*	Schmidt, 1893
		Phelister		*salobrus*	Marseul, 1887
		Phelister		*sanguinipennis*	Marseul, 1853
		Phelister		*sculpturatus*	Schmidt, 1893
		Phelister		*serratus*	Caterino and Tishechkin, 2020
		Phelister		*sparsus*	Caterino and Tishechkin, 2020
		Phelister		*sphaericus*	Caterino and Tishechkin, 2020
		Phelister		*stellans*	Caterino and Tishechkin, 2020
		Phelister		*striatinotum*	Wenzel and Dybas, 1941
		Phelister		*testudo*	Lewis, 1908
		Phelister		*thiemei*	Schmidt, 1889
		Phelister		*trigonisternus*	Marseul, 1889
		Phelister		*umens*	Caterino and Tishechkin, 2020
		Phelister		*vazdemelloi*	Caterino and Tishechkin, 2020
		Phelister		*vilavelha*	Caterino and Tishechkin, 2020
		Pseudister		*distractus*	(Schmidt, 1896)
		Pseudister		*suturalis*	(Schmidt, 1896)

Continues

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
		Pyxister		*devorator*	Caterino and Tishechkin, 2014
		Pyxister		*labralis*	Caterino and Tishechkin, 2014
		Scaptorus		*pyramus*	Caterino and Tishechkin, 2014
		Tarsilister		*loretoensis*	Bruch, 1932
		Yarmister		*emersoni*	Wenzel, 1944
	Histerini	*Hister*		*abhorrens*	(Schmidt, 1889)
		Hister		*alegre*	Caterino, 1999
		Hister		*catarinae*	Caterino, 1999
		Hister		*cavifrons*	Marseul, 1854
		Hister		*conductus*	Marseul, 1862
		Hister		*curvatus*	Erichson, 1834
		Hister		*denysi*	Marseul, 1870
		Hister		*diadema*	Marseul, 1854
		Hister		*incisifrons*	Marseul, 1863
		Hister		*lissurus*	Marseul, 1854
		Hister		*lucia*	Leivas, Moura and Caterino, 2015
		Hister		*pioti*	Marseul, 1870
		Hister		*punctifer*	Paykull, 1811
		Hister		*putridus*	Erichson, 1834
		Hister		*sturnus*	Marseul, 1862
		Margarinotus	*Paralister*	*ignobilis*	(Marseul, 1854)
	Hololeptini	*Dimalus*		*platamodes*	Marseul, 1870
		Eutidium		*bahiense*	(Marseul, 1860)
		Eutidium		*labrosum*	(Lewis, 1901)
		Eutidium		*lepidum*	Lewis, 1904
		Eutidium		*maragnoni*	(Marseul, 1870)
		Eutidium		*monodon*	(Marseul, 1870)
		Eutidium		*pacale*	Lewis, 1903
		Eutidium		*tuberculatum*	(Lewis, 1901)
		Eutidium		*williamsi*	(Dillon, 1935)
		Hololepta	*Hololepta*	*aradiformis*	Erichson, 1834
		Hololepta	*Hololepta*	*attenuata*	Blanchard, 1843
		Hololepta	*Hololepta*	*bogotana*	Marseul, 1853
		Hololepta	*Hololepta*	*bractea*	Erichson, 1834
		Hololepta	*Hololepta*	*canaliculata*	Lewis, 1901
		Hololepta	*Hololepta*	*cayennensis*	Marseul, 1853
		Hololepta	*Hololepta*	*curta*	Marseul, 1853
		Hololepta	*Hololepta*	*excisa*	Marseul, 1853
		Hololepta	*Hololepta*	*guidonis*	Marseul, 1860
		Hololepta	*Hololepta*	*lamina*	Paykull, 1811
		Hololepta	*Hololepta*	*quadriformis*	Marseul, 1853
		Hololepta	*Hololepta*	*subhumilis*	Marseul, 1853
		Hololepta	*Hololepta*	*wenzeli*	Mazur, 1984
		Hololepta	*Leionota*	*braziliensis*	Dillon, 1935
		Hololepta	*Leionota*	*cerdo*	(Marseul, 1853)
		Hololepta	*Leionota*	*cimex*	(Marseul, 1870)
		Hololepta	*Leionota*	*devia*	Marseul, 1853
		Hololepta	*Leionota*	*inermis*	Bickhardt, 1914
		Hololepta	*Leionota*	*lata*	(Marseul, 1853)
		Hololepta	*Leionota*	*minuta*	Erichson, 1834
		Hololepta	*Leionota*	*patula*	Lewis, 1906
		Hololepta	*Leionota*	*punctulata*	Marseul, 1853
		Hololepta	*Leionota*	*quadridentata*	(Olivier, 1789)
		Hololepta	*Leionota*	*reichii*	(Marseul, 1853)
		Oxysternus		*maximus*	(Linnaeus, 1767)
	Omalodini	*Ebonius*		*lineiger*	(Marseul, 1870)
		Ebonius		*politus*	Lewis, 1885
		Omalodes	*Cornillus*	*binodulus*	(Lewis, 1910)
		Omalodes	*Cornillus*	*bullatus*	Lewis, 1905
		Omalodes	*Cornillus*	*tuberculatus*	Lewis, 1905
		Omalodes	*Cornillus*	*tuberculifer*	Desbordes, 1917
		Omalodes	*Cornillus*	*tuberculipygus*	Schmidt, 1889
		Omalodes	*Cornillus*	*tuberosus*	Lewis, 1899
		Omalodes	*Diplogrammicus*	*ebeninus*	Erichson, 1834

Continues

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
		Omalodes	*Diplogrammicus*	*marseuli*	Schmidt, 1889
		Omalodes	*Omalodes*	*amazonicus*	Marseul, 1861
		Omalodes	*Omalodes*	*angulatus*	(Fabricius, 1801)
		Omalodes	*Omalodes*	*anthracinus*	Marseul, 1854
		Omalodes	*Omalodes*	*areolatus*	Schmidt, 1889
		Omalodes	*Omalodes*	*bifoveolatus*	Marseul, 1853
		Omalodes	*Omalodes*	*bisulcatus*	Desbordes, 1919
		Omalodes	*Omalodes*	*cerqueirae*	Desbordes, 1919
		Omalodes	*Omalodes*	*chapadae*	Lewis, 1908
		Omalodes	*Omalodes*	*exul*	Marseul, 1854
		Omalodes	*Omalodes*	*faustus*	Erichson, 1834
		Omalodes	*Omalodes*	*fortunatus*	Lewis, 1898
		Omalodes	*Omalodes*	*foveola*	Erichson, 1834
		Omalodes	*Omalodes*	*gagatinus*	Erichson, 1847
		Omalodes	*Omalodes*	*laceratus*	Marseul, 1854
		Omalodes	*Omalodes*	*laevicollis*	Bickhardt, 1911
		Omalodes	*Omalodes*	*lucidus*	Erichson
		Omalodes	*Omalodes*	*omega*	(Kirby, 1818)
		Omalodes	*Omalodes*	*planifrons*	Marseul, 1854
		Omalodes	*Omalodes*	*praevius*	Marseul, 1861
		Omalodes	*Omalodes*	*pulvinatus*	Erichson, 1834
		Omalodes	*Omalodes*	*punctistrius*	Marseul, 1854
		Omalodes	*Omalodes*	*punctulatus*	Moura and Almeida, 2013
		Omalodes	*Omalodes*	*rivus*	Moura and Almeida, 2013
		Omalodes	*Omalodes*	*serenus*	Erichson, 1834
		Omalodes	*Omalodes*	*seriatus*	Schmidt, 1889
		Omalodes	*Omalodes*	*sinuaticollis*	Marseul, 1854
		Omalodes	*Omalodes*	*sobrinus*	Erichson, 1834
		Omalodes	*Omalodes*	*wagneri*	Desbordes, 1919
	Platysomatini	*Platysoma*	*Cylister*	*directum*[e]	Lewis, 1885
	Incertae sedis	*Scapomegas*		*aurifer*	Marseul, 1887
		Scapomegas		*auritus*	Marseul, 1855
		Scapomegas		*gibbus*	Marseul, 1855
		Sphyracus		*anjubaulti*	(Marseul, 1864)
		Sphyracus		*gryphus*	(Marseul, 1861)
Saprininae	Euspilotini	*Euspilotus*	*Hesperosaprinus*	*alvarengai*	Arriagada, 2012
		Euspilotus	*Hesperosaprinus*	*amazonius*	(Desbordes, 1923)
		Euspilotus	*Hesperosaprinus*	*arcipygus*	(Schmidt, 1890)
		Euspilotus	*Hesperosaprinus*	*aterrimus*	(Erichson, 1834)
		Euspilotus	*Hesperosaprinus*	*auctus*	(Schmidt, 1890)
		Euspilotus	*Hesperosaprinus*	*azureus*	(Sahlberg, 1823)
		Euspilotus	*Hesperosaprinus*	*blandus*	(Erichson, 1834)
		Euspilotus	*Hesperosaprinus*	*connectens*	(Paykull, 1811)
		Euspilotus	*Hesperosaprinus*	*disnexus*	(Schmidt, 1890)
		Euspilotus	*Hesperosaprinus*	*emys*	(Marseul, 1870)
		Euspilotus	*Hesperosaprinus*	*eremita*	(Marseul, 1870)
		Euspilotus	*Hesperosaprinus*	*excavata*	Arriagada, 2012
		Euspilotus	*Hesperosaprinus*	*flaviclava*	(Marseul, 1870)
		Euspilotus	*Hesperosaprinus*	*innubus*	(Erichson, 1834)
		Euspilotus	*Hesperosaprinus*	*inversus*	(Lewis, 1899)
		Euspilotus	*Hesperosaprinus*	*jenseni*	(Bickhardt, 1911)
		Euspilotus	*Hesperosaprinus*	*milium*	(Marseul, 1855)
		Euspilotus	*Hesperosaprinus*	*modestus*	(Erichson, 1834)
		Euspilotus	*Hesperosaprinus*	*parenthesis*	(Schmidt, 1890)
		Euspilotus	*Hesperosaprinus*	*pavidus*	(Erichson, 1834)
		Euspilotus	*Hesperosaprinus*	*strobeli*	(Steinheil, 1869)
		Euspilotus	*Hesperosaprinus*	*sydovi*	(Bickhardt, 1911)
		Euspilotus	*Neosaprinus*	*limatus*	(Marseul, 1870)
		Euspilotus	*Neosaprinus*	*pipitzi*	(Marseul, 1887)
		Euspilotus	*Neosaprinus*	*rubriculus*	(Marseul, 1855)
		Euspilotus	*Neosaprinus*	*wenzeli*	Mazur, 1984
		Euspilotus	*Platysaprinus*	*ferreri*	Lackner and Arriagada, 2020
		Euspilotus	*Platysaprinus*	*latimanus*	(Schmidt, 1890)
		Euspilotus	*Platysaprinus*	*myrmecophilus*	(Bickhardt, 1910)

Continues

Subfamily	Tribe	Genus	Subgenus	Species	Author/Year
		Euspilotus	*Platysaprinus*	*vazdemelloi*	Lackner and Arriagada, 2020
		Geomysaprinus	*Priscosaprinus*	*bicirculus*	(Marseul, 1870)
		Phoxonotus	*Alienosternus*	*fryi*	Lewis, 1879
		Phoxonotus	*Alienosternus*	*venustus*	(Erichson, 1834)
		Phoxonotus	*Phoxonotus*	*tuberculatus*	Marseul, 1862
		Tatianaella		*dolata*	(Marseul, 1862)
	Hypocaccini	*Hypocaccus*	*Hypocaccus*	*brasiliensis*	(Paykull, 1811)
	Saprinini	*Saprinus*	*Saprinus*	*guyanensis*	Marseul, 1855
		Saprinus	*Saprinus*	*semirosus*	Marseul, 1870
		Saprinus	*Xerosaprinus*	*diptychus*	Marseul, 1855
		Saprinus	*Xerosaprinus*	*ignotus*	Marseul, 1855
Tribalinae		*Epierus*		*alberti*	Marseul, 1861
		Epierus		*alutaceus*	Marseul, 1855
		Epierus		*bisbistriatus*	Marseul, 1854
		Epierus		*fulvicornis*	Fabricius, 1801
		Epierus		*humeristrius*	Schmidt, 1889
		Epierus		*invidus*	Marseul, 1861
		Epierus		*kraatzi*	Schmidt, 1889
		Epierus		*mariae*	Marseul, 1861
		Epierus		*notius*	Marseul, 1861
		Epierus		*planulus*	Erichson, 1834
		Epierus		*vandepolli*	Schmidt, 1889
		Idolia		*punctisternum*	Lewis, 1885
		Plagiogramma		*amplistria*	Kovarik and Dégallier, 2021
		Plagiogramma		*angularis*	(Schmidt, 1896)
		Plagiogramma		*arciger*	(Marseul, 1854)
		Plagiogramma		*brasiliense*	Tarsia, 1935
		Plagiogramma		*epulo*	(Marseul, 1870)
		Plagiogramma		*erwini*	Kovarik and Dégallier, 2021
		Plagiogramma		*fissus*	(Marseul, 1861)
		Plagiogramma		*fornicata*	(Marseul, 1887)
		Plagiogramma		*glabra*	(Bickhardt, 1911)
		Plagiogramma		*inscriptus*	(Schmidt, 1896)
		Plagiogramma		*mundus*	(Erichson, 1834)
		Plagiogramma		*nitescens*	(Marseul, 1861)
		Plagiogramma		*paradoxa*	Mazur, 1988
		Plagiogramma		*parvoperuana*	Kovarik and Dégallier, 2021
		Plagiogramma		*peruana*	(Schmidt, 1889)
		Plagiogramma		*tersus*	(Erichson, 1834)

a) In the original description the species is mentioned as "Amérique mérid; Brésil?", that is, with a dubious location. The genus *Plegaderus* has most of its distribution in North America and Europe, therefore, the type material is possibly not from Brazil. While this issue is not resolved, we decided to maintain the species record for Brazil, but the resolution of this situation must be treated as a priority for future studies.

b) According to Dégallier et al. (2015; 2023), the South American specimens, including the Brazilian ones, probably belong to a taxon distinct from the Central America populations. The resolution of this situation must be addressed as a priority for future studies.

c) In the original description the species is mentioned as "Brésil (Coll. Chabrillac)." According to Cambefort (2006), Mr. François Chabrillac collected in Brazil, and his collection would be rich in specimens from South America. However, the genus *Pachycraerus* is distributed essentially on the African continent, therefore, the type material is possibly not from Brazil. During a 2017 visit to the Muséum National d'Histoire Naturelle – Paris (MNHN), the first and last authors of this paper were unable to locate the type material of *Pachycraerus chabrillaci*. Additionally, the genus *Pachycraerus* can be confused with other genera of Exosternini, such as *Baconia*, *Megalocraerus*, among others, making it important to study the type material for an adequate taxonomic resolution of this species. While this issue is not resolved, we decided to maintain the species record for Brazil, but the resolution of this situation must be treated as a priority for future studies.

d) This species is possibly not a *Paratropus*, but rather a *Fistulaster* Helava in Helava et al. (1985) (Dégallier, unpublished). This definition must be treated as a priority for future studies.

e) In the original description of *Platysoma* (*Cylister*) *directum*, Lewis indicated the origin of the species as "Para", later interpreted as belonging to Brazil. However, this species is the only Platysomatini taxon recorded for South America, as the tribe has its great diversity in Africa, Asia, Indonesia with some records for North America, Central America, Australia and New Zealand.

Dégallier et al. (2005) studied the type material of *Platysoma directum* and compared it with some other species, including those distributed in Central America, concluding that *Platysoma directum* may represent a species with a distribution further south on the American continent.

Appendix 2. Recent revisions and identification keys of the Histeridae taxa recorded in the Brazilian territory.

Subfamily/ Tribe	Genus	Bibliography source	Information content
Abraeinae		Arriagada et al. (2023)	Identification key to the Argentine genera.
Acritini	*Aeletes*	Leivas et al. (2012a)	Identification key to the species: Brazil.
Teretriini	*Teretrius*	Marseul (1856)	Identification key to the species.
Trypanaeini	*Coptotrophis*	Bickhardt (1917)	Identification key to the species.
	Trypanaeus	Marseul (1856), Bickhardt (1917)	Identification key to the species.
	Xylonaeus	Bickhardt (1917)	Identification key to the species.
Dendrophilinae		Arriagada et al. (2023)	Identification key to the Argentine genera.
Bacaniini	*Bacanius*	Wenzel and Dybas (1941)	Identification key to the species: Neotropical.
	Degallierister	Dégallier et al. (2015; 2023)	Identification key and distributional data of the species.
	Carcinops	Marseul (1864)	Identification key to the species.
	Paromalus	Marseul (1870)	Identification key to the species.
Haeteriinae		Helava et al. (1985), Arriagada et al. (2023)	Identification key to the genera, identification key to the Argentine species, respectively.
Haeteriini			
	Coelister	Dégallier (1998a)	Taxonomic revision.
	Colonides	Dégallier (1998b)	Identification key to the species.
	Plagioscelis	Dégallier (1998a)	Identification key to the species.
	Scapicoelis	Dégallier and Tishechkin (2022)	Identification key to the species.
	Termitoxenus	Dégallier (1998a)	Identification key to the species.
	Troglosternus	Dégallier et al. (2018)	Identification key to the species.
Nymphisterini		Tishechkin (2007)	Identification key to the genera.
	Nymphister	Dégallier et al. (2021)	Identification key to the species.
Synoditulini	*Synoditulus*	Helava et al. (1985); Tishechkin (2007)	Identification key to the genera.
Histerinae		Arriagada et al. (2023)	Identification key to the Argentine genera.
Exosternini	*Baconia*	Caterino and Tishechkin (2013a)	Identification key to the species.
	Chapischema	Caterino and Tishechkin (2014)	Identification key to the species.
	Conchita	Mazur (1994)	Original description.
	Conocassis	Caterino and Tishechkin (2014)	Identification key to the species.
	Crenulister	Caterino and Tishechkin (2014)	Identification key to the species.
	Kaszabister	Dégallier et al. (2012)	Identification key to the species.
	Lacrimorpha	Caterino and Tishechkin (2014)	Identification key to the species.
	Mecistostethus	Caterino and Tishechkin (2012)	Identification key to the species.
	Megalocraerus	Caterino and Tishechkin (2016)	Identification key to the species.
	Operclipygus	Caterino and Tishechkin (2013b)	Identification key to the species.
	*Phelister**	Caterino and Tishechkin (2019), Caterino and Tishechkin (2020)	Identification key to the *haemorrhous* and *blairi* species groups, respectively.
	Pseudister	Bickhardt (1917)	Identification key to the species (part).
	Pyxister	Caterino and Tishechkin (2014)	Identification key to the species.
	Scaptorus	Caterino and Tishechkin (2014)	Original description.
Histerini		Leivas et al. (2015a)	Key to the genera: Brazil.
	Hister	Caterino (1999a,1999b, 2002)	Identification key to the *servus* species group, *coenosus* species group and *militaris* species group, respectively.
Hololeptini		Dillon (1935)	Identification key to the genera (cited as Hololeptinae).
	Dimalus	Dégallier et al. (2010)	Identification key to the species: French Guiana.
	Eutidium	Marseul (1860), Desbordes (1916), Dillon (1935), Dégallier et al. (2010)	Identification key to the species (part, cited as *Phylloma*), identification key to the species (cited as *Orphinium* Lewis), identification key to the species, identification key to the species: French Guiana, respectively.
	Hololepta	Marseul (1853), Dégallier et al. (2010), Desbordes (1928)	Identification key to the species, identification key to the subgenus *Leionota* (cited as *Lioderma*), Identification key to the species: French Guiana, respectively.
	Oxysternus	Dégallier et al. (2010)	Identification key to the species: French Guiana
Omalodini		Leivas et al. (2015b)	Key to the genera.
	Ebonius	Dégallier et al. (2011)	Identification key to the species.
	Omalodes	Desbordes (1917), Desbordes (1919)	Identification key to the subgenus *Cornillus*, Identification key to the species, respectively.
Incertae sedis	*Scapomegas*	Leivas et al. (2012b)	Identification key to the species.
Saprininae		Arriagada et al. (2023)	Identification key to the Argentine genera.
Euspilotini	*Euspilotus*	Dégallier et al. (2012), Lackner and Arriagada (2020)	Identification key to the species: French Guiana, and identification key to the subgenus *Platysaprinus*, respectively.
	Phoxonotus	Lackner (2016)	Identification key to the species.
	Saprinus	Dégallier et al. (2012)	Identification key to the species: French Guiana
	Tatianaella	Arriagada (2018)	Original description.
Tribalinae		Arriagada et al. (2023)	Identification key to the Argentine genera.
	Epierus	Marseul (1864); Bickhardt (1920)	Identification key to the species; identification key for part of the American species, respectively.
	Plagiogramma	Dégallier et al. (2021)	Identification key to the species: French Guiana.

*See Caterino and Tishechkin (2013c) about the proposed conservation of usage of the name *Phelister*.

Appendix 3. The Histeridae species recorded in Brazilian conservation areas.

Region / State	Conservation Unit	Biome	Taxa	Citation
North				
Amazonas	Adolpho Ducke Forest Reserve	Amazon	*Omalodes bifoveolatus* Marseul, *Omalodes foveola* Erichson, *Omalodes lucidus* Erichson, *Phelister sanguinipennis* Marseul, *Scapomegas auritus* Marseul	Mise et al. (2010), Leivas et al. (2012)
Northwest	Parque Nacional de Sete Cidades (PNSC)	Caatinga	*Omalodes foveola* Erichson, *Hister cavifrons* Marseul, *Phelister panamensis* J.E. Le Conte, *Euspilotus blandus* (Erichson), *Euspilotus parenthesis* (Schmidt), *Euspilotus innubus* (Erichson), *Euspilotus eremita* (Marseul), *Euspilotus pavidus* (Erichson), *Euspilotus sydovi* (Bickhardt), *Euspilotus milium* (Marseul), *Saprinus* (*Xerosaprinus*) *diptychus* (Marseul)	Rafael et al. (2017)
	Parque Nacional de Ubajara (PNU)	Caatinga	*Omalodes foveola* Erichson, *Phelister brevistrius* Marseul, *Scapomegas gibbus* Marseul, *Euspilotus blandus* (Erichson),	Rafael et al. (2017)
Paraíba	Reserva Particular de Patrimônio Natural Fazenda Almas	Caatinga	*Euspilotus azureus* (Sahlberg), *Hister punctifer* Paykull, *Hololepta reichii* Marseul, *Omalodes foveola* Erichson *Saprinus* (*Xerosaprinus*) *diptychus* (Marseul).	Santos et al. (2014)
Southeast				
Minas Gerais	Area de Proteção Ambiental Serra de São Jose		*Carcinops troglodytes* (Paykull), *Euspilotus alvarengai* Arriagada, *Euspilotus azureus* (Sahlberg), *Euspilotus excavata* Arriagada, *Hister cavifrons* Marseul, *Hister lissurus* Marseul, *Omalodes foveola* Erichson, *Omalodes omega* (Kirby), *Scapomegas aurifer* Marseul	Vieira et al. (2018)
Minas Gerais	Refúgio da Vida Silvestre Lilbélulas da Serra de São Jose	Cerrado and Atlântic Forest	*Carcinops troglodytes* (Paykull), *Euspilotus alvarengai* Arriagada, *Euspilotus azureus* (Sahlberg), *Euspilotus excavata* Arriagada, *Hister cavifrons* Marseul, *Hister lissurus* Marseul, *Omalodes foveola* Erichson, *Omalodes omega* (Kirby), *Scapomegas aurifer* Marseul	Vieira et al. (2018)
South				
Paraná	Área Especial de Interesse Turístico do Marumbi (AEIT) do Marumbí	Atlantic Forest	*Baconia bullifrons* Caterino and Tishechkin, *Baconia grossii* (Caterino and Tishechkin), *Colonides hubrichi* Bruch, *Colonides quadriglumis* (Reichensperger), *Hister punctifer* Paykull, *Iugulister clarissae* Reichensperger, *Omalodes* (s. str.) *angulatus* (Fabricius), *Omalodes* (s. str.) *bisulcatus* Desbordes, *Omalodes* (s. str.) *foveola* (Erichson), *Omalodes* (s. str.) *laceratus* Marseul, *Omalodes* (s. str.) *planifrons* Marseul, *Omalodes (Omalodes) praevius* Marseul, *Operclipygus distractus* (Schmidt), *Operclipygus friburgius* (Marseul), *Operclipygus iheringi* (Bickhardt), *Operclipygus sejunctus* (Schmidt), *Operclipygus subrufus* Caterino and Tishechkin, *Operclipygus subterraneus* Caterino and Tishechkin, *Scapomegas aurifer* Marseul, *Phelister almeidae* Caterino and Tishechkin, *Phelister gregarius* Caterino and Tishechkin, *Phelister matatlantica* Caterino and Tishechkin, *Phelister parana* Caterino and Tishechkin, *Phelister rio* Caterino and Tishechkin, *Scapicoelis bichoae* Dégallier and Tishechkin	Leonel and Leivas (2023)
Paraná	Estação Ecológica Caiuá	Atlantic Forest	*Euspilotus (Hesperosaprinus) azureus* (Sahlberg), *Omalodes* (s. str.) *angulatus* (Fabricius)	Leonel and Leivas (2023)
Paraná	Estação Ecológica Ilha do Mel	Atlantic Forest	*Hister cavifrons* Marseul	Leonel and Leivas (2023)
Paraná	Parque Ecológico Samuel Klabin	Atlantic Forest	*Omalodes* (s. str.) *angulatus* (Fabricius)	Leonel and Leivas (2023)
Paraná	Parque Estadual de Guartelá	Cerrado	*Baconia* (s. str.) *angusta* Schmidt, *Baconia guartela* Caterino and Tishechkin, *Hister cavifrons* (Marseul), *Hister lissurus* Marseul, *Hister punctifer* Paykull, *Omalodes* (s. str.) *laceratus* Marseul, *Operclipygus distractus* Schmidt, *Operclipygus iheringi* (Bickhardt), *Operclipygus latemarginatus* Bickhardt, *Operclipygus subrufus* Caterino and Tishechkin, *Operclipygus subterraneus* Caterino and Tishechkin, *Phelister panamensis* J.E. Le Conte, *Phelister pumilus* (Erichson), *Scapomegas aurifer* Marseul.	Leonel and Leivas (2023)
Paraná	Parque Estadual do Cerrado	Cerrado	*Hister cavifrons* Marseul, *Omalodes* (s. str.) *foveola* (Erichson)	Leonel and Leivas (2023)
Paraná	Parque Estadual do Marumbi	Atlantic Forest	*Omalodes* (s. str.) *fortunatus* Lewis	Leonel and Leivas (2023)
Paraná	Parque Estadual Mata São Francisco	Atlantic Forest	*Omalodes* (s. str.) *angulatus* (Fabricius), *Omalodes* (s. str.) *foveola* (Erichson), *Omalodes* (s. str.) *omega* (Kirby), *Operclipygus* aff. *wenzeli* Caterino and Tishechkin, *Operclipygus farctus* (Marseul), *Operclipygus proximus* Caterino and Tishechkin, *Phelister panamensis* J.E. Le Conte, *Reninus meticulosus* (Lewis), *Scapomegas aurifer* Marseul, *Scapomegas gibbus* Marseul	Leonel and Leivas (2023)
Paraná	Parque Estadual São Camilo	Atlantic Forest	*Hister cavifrons* Marseul, *Omalodes* (s. str.) *foveola* (Erichson)	Leonel and Leivas (2023)
Paraná	Parque Estadual Vila Velha	Atlantic Forest	*Operclipygus sejunctus* (Schmidt), *Operclipygus subterraneus* Caterino and Tishechkin, *Phelister brevistrius* Marseul.	Leonel and Leivas (2023)
Paraná	Parque Mata dos Godoy	Atlantic Forest	*Scapomegas aurifer* Marseul	Leonel and Leivas (2023)
Paraná	Reserva Biológica das Perobas	Atlantic Forest	*Euspilotus azureus* (Sahlberg), *Hister cavifrons* Marseul, *Plagioscelis striaticeps* (Bruch), *Scapomegas gibbus* Marseul	Leonel and Leivas (2023)
Paraná	Reserva Estadual de Vila Rica	Atlantic Forest	*Omalodes* (s. str.) *foveola* (Erichson)	Leonel and Leivas (2023)
Rio Grande do Sul	Parque Estadual do Turvo	Atlantic Forest	*Omalodes* (s. str.) *angulatus* (Fabricius)	Leonel and Leivas (2023)
Rio Grande do Sul	Parque da Ferradura	Atlantic Forest	*Kaszabister rubellus* (Erichson)	Leonel and Leivas (2023)