



Differences in mutational processes and intra-tumour heterogeneity between organs

The local selective filter hypothesis

Mathieu Giraudeau^{1,*,1,†} Tuul Sepp^{2,†} Beata Ujvari^{3,4} François Renaud,¹ Aurélie Tasiemski,⁵ Benjamin Roche,^{1,6,7} Jean-Pascal Capp^{8,†} and Frédéric Thomas^{1,†}

¹CREEC, UMR IRD 224-CNRS 5290-Université de Montpellier, Montpellier, France; ²Institute of Ecology and Earth Sciences, University of Tartu, Vanemuise 46, Tartu 51014, Estonia; ³School of Natural Sciences, University of Tasmania, Private Bag 55, Hobart, Tasmania 7001, Australia; ⁴Centre for Integrative Ecology, School of Life and Environmental Sciences, Deakin University, Waurn Ponds, Victoria 3216, Australia; ⁵Université de Lille-sciences et technologies, UMR 8198 Evo-Eco-Paleo, Villeneuve d'Ascq/CNRS/INSERM/CHU Lille, Institut Pasteur de Lille, U1019-Unité Mixte de Recherche 8204, Lille, France; ⁶IRD, Sorbonne Université, UMMISCO, F-93143, Bondy, France; ⁷Departamento de Etología, Fauna Silvestre y Animales de Laboratorio, Facultad de Medicina Veterinaria y Zootecnia, Universidad Nacional Autónoma de México (UNAM), Ciudad de México, México and ⁸INSA/Université Fédérale de Toulouse, Laboratoire d'Ingénierie des Systèmes Biologiques et des Procédés, UMR CNRS 5504, UMR INRA 792, Toulouse, France

*Corresponding author. Tel: 33 (0) 4 67 41 63 18; E-mail: giraudeau.mathieu@gmail.com.

†These authors contributed equally.

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ABSTRACT

Extensive diversity (genetic, cytogenetic, epigenetic and phenotypic) exists within and between tumours, but reasons behind these variations, as well as their consistent hierarchical pattern between organs, are poorly understood at the moment. We argue that these phenomena are, at least partially, explainable by the evolutionary ecology of organs' theory, in the same way that environmental adversity shapes mutation rates and level of polymorphism in organisms. Organs in organisms can be considered as specialized ecosystems that are, for ecological and evolutionary reasons, more or less efficient at suppressing tumours. When a malignancy does arise in an organ applying strong selection pressure on tumours, its constituent cells are expected to display a large range of possible surviving strategies, from hyper mutator phenotypes relying on bet-hedging to persist (high mutation rates and high diversity), to few poorly variable variants that become invisible to natural defences. In contrast, when tumour suppression is weaker, selective pressure favouring extreme surviving strategies is relaxed, and tumours are moderately variable as a result. We provide a comprehensive overview of this hypothesis.

Lay summary: Different levels of mutations and intra-tumour heterogeneity have been observed between cancer types and organs. Anti-cancer defences are unequal between our organs. We propose that mostly aggressive neoplasms (i.e. higher mutational and ITH levels), succeed in emerging and developing in organs with strong defences.

KEYWORDS: cancer; intra-tumoural heterogeneity; selection; mutation

Extensive genetic, cytogenetic and epigenetic variation, as well as phenotypic diversity exist between and within tumours (i.e. inter- and intra-tumour heterogeneity, respectively, the latter being called ITH thereafter) [1–4]. Three main types of stochastic phenomena leading to ITH can be distinguished. The most well-known and by far most studied is genetic variability resulting from mutational processes [5]; a more recent but also intensively studied field is epigenetic variability [6] and finally the most largely unexplored is gene expression variability [7]. The implications and clinical importance of these different sources of ITH are considerable since ITH may underlie incomplete treatment responses, acquired and/or innate resistance, and disease relapse in response to chemotherapy and targeted agents [8–12]. Because ITH is an important clinical determinant of patient outcomes, its origins have been the subject of much discussion by investigators. While genomic instability seems to be *the* major proximate process generating ITH [13], no consensus has however emerged between the several (non-mutually exclusive) hypotheses proposed to explain its establishment and maintenance [14, 15] (Box 1). Interestingly, several studies have also highlighted that both mutational processes and ITH between cancer types display a relatively constant hierarchical pattern between organs, with for instance melanoma and lung cancers being on average the most heterogenous cancers [16–19]. Despite extensive research, the processes behind this hierarchy remain unclear as well.

In this paper, we propose that variation in mutational patterns and ITH result from the evolutionary ecology of organs' theory [20], and are therefore explained by the same rules than those governing mutational patterns and polymorphism in organisms living in more or less adverse habitats [21, 22]. For instance, genomic diversity is generally positively correlated with abiotic and biotic stress levels (e.g. [21, 23, 24]), leading sometimes to the selection of hyper-mutator phenotypes [25, 26]. Beyond a high-threshold level of stress, the diversity may also sometimes decline to a few adapted genotypes potentially displaying strong evolutionary convergence [27, 28]. Organs in organisms can be considered as specialized ecosystems in a living landscape, whose ecologies are more or less favourable to cancer progression. The evolutionary ecology of organs' theory predicts that the evolution of organ-specific resistance to malignant emergence and/or progression should be governed by their level of exposure to oncogenic factors together with the host's evolutionary responses in relation with the direct or indirect fitness importance of each organ [20]. Here, assuming

that the number of mutations typically found in a cancer is an indicator of the diversity of molecular characteristics of cancer cells, we discuss the extent to which this hypothesis could explain inter organ variability in mutational and ITH hierarchy patterns, as well as examine whether it could explain the predilection for metastatic site(s) by different cancer types.

THE LOCAL SELECTIVE FILTER HYPOTHESIS

Recently, Vittecoq *et al.* [29] argued that a promising research direction for discovering novel anticancer therapies consist in exploring cancer suppressive mechanisms in animals living in environments that favour cancer emergence and/or progression. Indeed, the same way as the lack of correlation between body size/life expectancy and cancer incidence led to Peto's paradox [30], a lack of correlation between exposure to oncogenic factors and cancer incidence might suggest that evolution has produced solutions to avoid and/or control malignant problems in those species. From an evolutionary perspective, a similar conceptual framework can be applied at the organs' level [31, 32]. Concretely, we expect that selection has locally shaped powerful natural defences against malignant emergence/progression, and hence mostly aggressive neoplasms (i.e. higher mutational and ITH levels), or conversely few invisible ones succeed in emerging and developing in organs strongly exposed to mutagenic substances. In contrast, organs that are less exposed to oncogenic factors have been less optimized by selection to be efficient at controlling malignant developments, and as a result, less aggressive neoplasms (i.e. lower mutational and ITH levels) may regularly emerge and progress in these tissues. These predictions seem in accordance with the hierarchical patterns observed for both mutational processes and ITH. Indeed, skin, lung or the digestive tract are, all things being equal, for instance undoubtedly more exposed to mutagenic substances than breast, pancreas or thyroid [33–35]. This phenomenon is of course exacerbated in our modern world [36]. Similarly, differential exposures to injuries and/or to infections, which can promote secondarily carcinogenesis, exist between organs [37].

Following the same idea, it has been long accepted in evolutionary immunology that strong immunological defences are also costly at the organ and tissue levels in terms of oxidative damage, since increased level of reactive oxygen species (ROS) is a by-product of elevated metabolism associated

with an immune response, but also a defence mechanism used by immune cells [38, 39]. The level of these oxidative costs (or the strength of protection against these costs) can be organ specific, as has been demonstrated in several studies on wild animals (e.g. [40, 41]). Accordingly, organs that are more efficient at controlling early-stage malignant emergence at the level of immune responses could be more vulnerable to tumour-promoting inflammation and mutations caused by ROS on the genomic level, resulting in higher ITH of neoplasms in these organs. Thus, tumour-promoting inflammation and antitumor immunity coexist at different points along the path of tumour progression, and environmental and micro-environmental conditions should dictate the balance between the two [42].

Carcinogenesis also typically occurs within the spatial constraints of the epithelial layer of the organ. In the breast and pancreas, for example, this involves tumour growth within a narrow duct while in the colon, premalignant lesions (e.g. polyps) grow into the lumen of the bowel and on the skin. The cell–cell interaction network is another factor that could explain the differential sensitivity of organs and tissues to neoplasm development. Indeed, it could be a major contributor among the cancer suppressive mechanisms in animals. When examining the connectivity of 144 cell types in terms of ligands and receptors, recent works found that hematopoietic lineages are outliers because they are far less connected than all other cell types [43]. These lineages are also known to be the less mutated [16] and seem to not necessitate strong genetic instability, suggesting that a major suppressive force might be situated at the level of the cell–cell interaction network. Nevertheless, apart from blood cancers, brain cancers such as glioma can also be characterized by low mutational load. Interestingly, gliomas are also the ones with the highest ITH among solid tumours [19], suggesting that the low mutational load in these cases is compensated by high ITH. As previously discussed (Box 2), if tissue disruption is an initiator event in oncogenesis [44, 45], a strong and dense cell–cell interaction network is expected to more efficiently prevent malignant development. Thus, the more cells are connected in a tissue or an organ, higher ITH is necessary for oncogenesis to occur, at least during the first steps of tumourigenesis. On the contrary, less connected tissues are expected to contribute to cancer types with the lowest ITH levels. Interestingly, works in ecology revealed a correlation between the connectivity between species in an ecosystem and the resistance to invaders [46], suggesting again that homology between species in ecological niches and cell types in organs could be relevant.

The local selective filter hypothesis not only provides an explanation for the different levels of mutations and ITH observed between cancer types and organs, but also supports the fact that cancer could initiate when this selective filter at the tissue and organ levels is broken down. Interestingly, genes linked to multicellularity are systematically repressed in solid cancers while those that are more associated with unicellularity are upregulated

[47]. This observation, which is concordant with the atavism hypothesis [48, 49], suggests that cancer cells transit to a more ‘selfish’ unicellular mode of life through an active and directed process driven by selection [48]. Especially, genes linked to the extracellular matrix and adhesion as well as signalling and cell communication are mostly downregulated. Among the seven cancer types studied, those (breast and prostate) that have the most similar expression profile of multicellularity associated genes to normal tissue are also the least mutated [50], suggesting again that the level of genetic instability could be dependent on the need to break down the network of dense cellular interactions.

Finally, the nature and frequency of cancer stem cells are still a controversial debate. Inconsistencies in the numbers of such cells reported in the literature can be a consequence of the different definitions used by researchers. As suggested below (Box 2), oncogenesis could result from tissue disruption that generates differentiation problems because of the lack of tissue control [44, 45]. In our opinion, cancer stem cells have to be considered as cells acquiring highly unstable and variable phenotypes similar the ones of normal stem cells (due to high gene expression noise), but without the normal control normally exerted by the micro-environment.

If cancer development depends especially on the ability to counteract the cell–cell interaction network, it could be assumed that dedifferentiation (from differentiated cells) or failure of differentiation (from adult stem cells) is the best way to generate cells that are no more submitted to micro-environmental control because of the intrinsic plasticity and instability of such cancer stem-like cells [44, 51]. Consequently, the more cells of a tissue are under the control of their environment, the more they would need to acquire stem-cell like properties (and in higher number) to overcome this local selective filter. Tumours in tissues with stronger local selective filter would logically contain more cells with such unstable phenotypes. Thus, this framework could explain the differences in the frequency of cancer stem cells between tumours. As these cells are themselves a source of phenotypic heterogeneity, this would be also associated to higher non-genetic ITH in these tumours.

METASTATIC PREDILECTION SITE

Due to higher mutation rates, cancerous cell communities originating from neoplasias in organs with high resistance to malignant emergence should be able to produce metastases in a wider variety of organs compared with less diverse tumour cell communities. The understanding that metastasis results when tumour cells interact with a specific organ’s micro-environment stems from the ‘seed and soil’ hypothesis, stating that certain tumour cells (‘seed’) have specific affinity for the milieu of certain organs (‘soil’), and metastases form only when the seed and soil are compatible [52, 53]. Although this hypothesis has been one of the most persistent in the study of cancer, and supported by a wide range of experimental evidence [53], it has not been linked to



Box 1. Current hypotheses for the establishment and the maintenance of different types of intra-tumour heterogeneity (ITH)

Several non-mutually exclusive models have been recently published to explain the establishment and maintenance of ITH. For certain cancer/organ combination, it has been argued that a significant proportion of somatic mutations result from **exposures to mutagens**, e.g. ultraviolet light in skin cancers, or tobacco smoking in lung cancers [58]. While this process undoubtedly contributes to generate ITH, it cannot account, alone, for the extreme ITH values frequently observed in certain organs, especially in tiny tumours (e.g. [59]). Waclaw *et al.* [60] proposed a model for tumour evolution suggesting that **cell turnover** together with short-range migration can account for rapid cell mixing within the tumour. Alternatively, according to the **cancer stem cell hypothesis**, ITH results from the differentiation of few cells with stem cell properties (e.g. unrestricted self-renewal abilities) that produce various cell types in the tumour [61]. In parallel, the **linear clonal evolution hypothesis** suggests that ITH is due to the accumulation of various hereditary changes over time that confer selective advantages to some premalignant and malignant cells [62]. Finally, the **plasticity cell hypothesis** postulates that the majority of tumour cells, depending on micro-environmental conditions and/or cell intrinsic stochasticity, display varying degrees of stem cell-like characteristics [14]. In accordance with this idea, Lloyd *et al.* [63] suggested that (at least some) intra-tumour heterogeneity in the molecular properties of cancer cells is governed by predictable regional variations in environmental selection forces. In fact, a common point in these hypotheses is to argue that because ITH plays a crucial role in neoplasia, cancer progression and therapeutic resistance, its persistence, once initiated, is supported by various selective costs and benefits. Although realistic in many cases, this hypothesis has, however, some limitations because environments change unpredictably and evolution cannot anticipate the future. It is, therefore, challenging to explain the occurrence of ITH at the very first steps of the tumourigenesis. Genetic ITH can be so extreme even in tiny tumours, that Ling *et al.* [59] recently argued that evolution under a ‘non-Darwinian mode’ is plausible because genetic diversity observed would be orders of magnitude lower than predicted by simple classic Darwinian selection.

Recently, Thomas *et al.* [15] argued that generative mechanisms of ITH could also provide selective advantages to cells from the first steps of oncogenesis. In this hypothesis, malignant cells achieve greater success by **cooperating** in the process of tumour construction, providing the other with a common good, rather than by just being proliferative in isolation. There would be a concomitant selection of a bet-hedging strategy during oncogenesis, and hence ITH because this is necessary to generate the diversity of cell components needed to build, *de novo*, a novel and an intricate cooperative system like the solid tumour is.

Finally, the molecular heterogeneity within tumours could be fundamentally driven by variations in spatial and temporal distribution of blood flow (see for instance [64, 65]), suggesting that variations in patterns of angiogenesis in different organs could be the primary driver of molecular heterogeneity.

the mutation and ITH patterns. At the same time, it is logical to assume that the most lethal metastatic ‘seeds’ evolve as a result of selective pressure in the primary tumour [54], and the selective pressure assumed to be the highest in organs that are most protected against malignant developments.

For example, while melanoma (rated highest on mutational processes by [50] and 2nd by [17]) cells introduced in mouse circulation can cause tumour development in a wide variety of tissue types [55], human ovarian cancer (ranked 14th at ITH by [50] and 19th by [17]) cells, despite continuous entry of millions of tumour cells into the circulation, rarely cause metastases even to the lung, the first capillary bed encountered [56]. Similarly, prostate cancer (ranked 15th by [50] and 21st by [17]) ‘seeds’ have a very low probability to find a compatible ‘soil’ in any tissues, but colorectal cancer (1st by [17] and 7th by [50]) ‘seeds’ readily give metastasis in a number of organs, exhibiting a cascading spread of gastro-

intestinal tumours, where metastases in secondary sites spread ‘seeds’ to the organs that follow the blood drain route [57].

CONCLUDING REMARKS

We only see cancers that succeed in their development and this fraction corresponds to malignancies that bypass our natural defences. As soon as defences are unequal between the different parts of the body, the fraction of successful cancers is also expected to vary accordingly. By suggesting that the efficiency of natural defences against cancer development is organ specific and that it explains the hierarchy in mutation load and ITH between organs, our hypothesis highlights the role of the cellular environment in shaping the tumoural genomic and epigenomic content. In standard models of cancer, the cellular environment is destroyed as a consequence of cancer progression, thus has also a



Box 2. ITH on the level of gene expression

Apart from genetic ITH, high heterogeneity in gene expression is observed within cancers [7], even at the single-cell level [66]. Cancer cells harbour a continuum of heterogeneous phenotype states demarcated by gradients of marker expression rather than distinct subpopulations [67]. An early increase in non-genetic ITH, especially in gene expression variability from cell-to-cell, has been suggested to account for phenotypic diversification in early steps and ultimately to oncogenesis [44]. Indeed, gene expression variability is modulated during development and differentiation and many studies showed that following a phase of highly stochastic and widespread gene expression, cells progressively transit towards a more homogeneous, coordinated and restricted gene expression pattern [68–70]. Cellular interactions are major determinants in constraining and decreasing gene expression variability and seem to be the main ‘constraints’ leading to these stable differentiated states [71, 72]. For instance, direct cell contacts through gap junctions spatially coordinate prolactin gene expression in pituitary adult tissue [73]. Moreover, enzymatic digestion of extracellular proteins or pharmacological inhibition of gap junctions reduced transcriptional coordination between cells [73], showing that perturbation of cell communication can enhance gene expression variability and phenotypic heterogeneity among differentiated cells. Thus, tissue disruption could be the initial source of gene expression ITH [44, 45] and genetic instability has been proposed to be caused by this early gene expression ITH [45]. In accordance with this hypothesis, numerous studies have now shown that tissue disruption can be either the inducer or the repressor of the cancerous state [44, 74–76]. Therefore, the presence of epigenetic [77], gene expression [78] or micro-environmental [79] alterations that might precede the emergence of genetically abnormal cells further argues for a major role of non-genetic processes in the first steps of oncogenesis.

The early increase in gene expression variability allows another type of bet-hedging that can synergize with genetic ITH to allow phenotypic diversification, the transcriptional ITH. When RNA-seq data were used to measure the level of transcriptional ITH, 12 major cancer types showed distinct levels of this type of ITH [7]. Interestingly, when these results were compared with previous data on genetic ITH, a positive correlation between genetic heterogeneity and transcriptional ITH was found [7]. Both types of ITH can thus be considered as relevant forces in a bet-hedging strategy where the level of heterogeneity would be dependent on the level of cooperation needed in the process of tumour construction to bypass suppressive forces in tissues.

Finally epigenetic alterations are also increasingly acknowledged as being able to initiate transformation, as genetic alterations do, by providing the gene expression plasticity necessary to provide stochastic oncogenic epigenetic changes [80]. Epigenetic instability can also allow phenotypic diversification in the bet-hedging strategy that we proposed here in the early steps of oncogenesis. Interestingly, while the global levels of genetic and epigenetic variations between tumour types are mostly uncorrelated [81], when epigenetic and genetic ITH were measured by analysis of DNA methylation and copy number alterations in aggressive prostate cancer, the structure of phylogenetic trees constructed from the epigenetic and genetic data were very close, indicating a similarity in evolutionary process [82]. Other works revealed such correlation [6]: for instance, the level of DNA methylation ITH within an individual’s leukaemia was positively correlated with the level of genetic ITH [83]. Landau *et al.* [84] also found this correlation in chronic lymphocytic leukaemia between high numbers of sub-clonal mutations and high DNA methylation ITH. Finally, this correlation between genetic and epigenetic heterogeneity was completed in this last work by an additional correlation identified with data from single-cell RNA sequencing: promoters with high methylation ITH showed high cell-to-cell expression heterogeneity of the corresponding gene [84]. Altogether these works reveal that genetic, epigenetic and gene expression ITH are mostly correlated and suggest that different levels of all these types of ITH, and thus different levels of bet-hedging, are needed depending on the tissue and organ considered. However, despite a diversity of hypotheses that explain why ITH is omnipresent, the reasons governing these different levels of ITH are still unclear.

passive role and no driver role in tumour evolution. On the contrary, our hypothesis considers that the level of diversity needed for cancer development is precisely dependent on and the consequence of the ability of an organ to suppress this development. It contributes to the continuously growing body of works that place

major influences in oncogenesis at higher levels of organization than the genomic level while not denying the major contribution of genetic and epigenetic instability in cancer progression.

An evolutionary theory needs to consider more than just humans, and needs to consider ‘historical’ cancer patterns that would be

more in line with the selective pressure experienced by our ancestors (which largely determined our genetic makeup and tumour suppressive strategies). While historical cancer data are undoubtedly difficult to find, we encourage scientists to explore our hypothesis using different datasets among various animal species. We also encourage researchers to perform experimental studies specifically designed to test whether cells with high mutation rate are, as proposed here, more likely to metastasize than others.

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REFERENCES

- Fidler IJ. Tumor heterogeneity and the biology of cancer invasion and metastasis. *Cancer Res* 1978;**38**:2651–60.
- Marusyk A, Polyak K. Tumor heterogeneity: causes and consequences. *Biochim Biophys Acta* 2010;**1805**:105–17.
- Marusyk A, Almendro V, Polyak K. Intra-tumour heterogeneity: a looking glass for cancer? *Nat Rev Cancer* 2012;**12**:323–34.
- Yates LR, Campbell PJ. Evolution of the cancer genome. *Nat Rev Genet* 2012;**13**:795–806.
- Greaves M, Maley CC. Clonal evolution in cancer. *Nature* 2012. DOI: 10.1038/nature10762
- Mazor T, Pankov A, Song JS *et al.* Intratumoral heterogeneity of the epigenome. *Cancer Cell* 2016;**29**:440–51.
- Park Y, Lim S, Nam JW *et al.* Measuring intratumor heterogeneity by network entropy using RNA-seq data. *Sci Rep* 2016;**6**:DOI: 10.1038/srep37767
- Pribluda A, De La Cruz CC, Jackson EL. Intratumoral heterogeneity: from diversity comes resistance. *Clin Cancer Res* 2015;**21**:2916–23.
- Allison KH, Sledge GW. Heterogeneity and Cancer. *Oncology (Williston Park)*. 2014;**28**:1–9.
- Mengelbier LH, Karlsson J, Lindgren D *et al.* Intratumoral genome diversity parallels progression and predicts outcome in pediatric cancer. *Nat Commun* 2015;**6**:DOI: 10.1038/ncomms7125
- Maley C. Multistage carcinogenesis in Barrett's esophagus. *Cancer Lett* 2007;**245**:22–32.
- Roerink SF, Sasaki N, Lee-Six H *et al.* Intra-tumour diversification in colorectal cancer at the single-cell level. *Nature* 2018;**556**:457–62.
- Burrell RA, McGranahan N, Bartek J *et al.* The causes and consequences of genetic heterogeneity in cancer evolution. *Nature* 2013;**501**:338–45.
- Michor F, Polyak K. The origins and implications of intratumor heterogeneity. *Cancer Prev Res* 2010;**3**:1361–4.
- Thomas F, Ujvari B, Gidoin C *et al.* Toward an ultimate explanation of intratumor heterogeneity. *Ecol Evol Cancer* 2017;**2**:19–22.
- Alexandrov LB, Nik-Zainal S, Wedge DC *et al.* Signatures of mutational processes in human cancer. *Nature* 2013;**500**:415–21.
- Vormehr M, Diken M, Boegel S *et al.* Mutanome directed cancer immunotherapy. *Curr Opin Immunol* 2016;**39**:14–22.
- Andor N, Graham TA, Jansen M *et al.* Pan-cancer analysis of the extent and consequences of intratumor heterogeneity. *Nat Med* 2016. DOI: 10.1038/nm.3984
- McGranahan N, Swanton C. Clonal heterogeneity and tumor evolution: past, present, and the future. *Cell* 2017;**168**:613–28.
- Thomas F, Nesse RM, Gatenby R *et al.* Evolutionary ecology of organs: a missing link in cancer development? *Trends Cancer* 2016;**2**:409–15.
- Nevo E. Evolution of genome-phenome diversity under environmental stress. *Proc Natl Acad Sci* 2001. DOI: 10.1073/pnas.101109298
- Hedrick PW. Genetic polymorphism in heterogeneous environments: the age of genomics. *Annu Rev Ecol Evol Syst* 2006. DOI: 10.1146/annurev.ecolsys.37.091305.110132
- Galhardo RS, Hastings PJ, Rosenberg SM. Mutation as a stress response and the regulation of evolvability. *Crit Rev Biochem Mol Biol* 2007. DOI: 10.1080/10409230701648502
- Woodford N, Ellington MJ. The emergence of antibiotic resistance by mutation. *Clin Microbiol Infect* 2007. DOI: 10.1111/j.1469-0691.2006.01492.x
- Waine DJ, Honeybourne D, Smith EG *et al.* Association between hypermutator phenotype, clinical variables, mucoid phenotype, and antimicrobial resistance in *Pseudomonas aeruginosa*. *J Clin Microbiol* 2008. DOI: 10.1128/JCM.00357-08
- Swings T, van Den Bergh B, Wuyts S *et al.* Adaptive tuning of mutation rates allows fast response to lethal stress in *Escherichia coli*. *Elife* 2017. DOI: 10.7554/eLife.22939
- Kis-Papo T, Kirzhner V, Wasser SP *et al.* Evolution of genomic diversity and sex at extreme environments: fungal life under hypersaline Dead Sea stress. *Proc Natl Acad Sci U S A* 2003. DOI: 10.1073/pnas.2036284100
- Pan S, Zhang T, Rong Z *et al.* Population transcriptomes reveal synergistic responses of DNA polymorphism and RNA expression to extreme environments on the Qinghai–Tibetan Plateau in a predatory bird. *Mol Ecol* 2017. DOI: 10.1111/mec.14090
- Vittecoq M, Giraudeau M, Sepp T *et al.* Turning natural adaptations to oncogenic factors into an ally in the war against cancer. *Evol Appl* 2018. DOI: 10.1111/eva.12608
- Caulin AF, Maley CC. Peto's paradox: evolution's prescription for cancer prevention. *Trends Ecol Evol* 2011;**26**:175–82.
- Nunney LL. selection and the evolution of multistage carcinogenesis. *Proc R Soc B Biol Sci* 1999. DOI: 10.1098/rspb.1999.0664
- Noble R, Kaltz O, Hochberg ME. Peto's paradox and human cancers. *Philos Trans R Soc Lond B Biol Sci* 2015;**370**:20150104.
- D'Orazio J, Jarrett S, Amaro-Ortiz A *et al.* UV radiation and the skin. *Int J Mol Sci* 2013. DOI: 10.3390/ijms140612222
- Wakabayashi K, Nagao M, Esumi H *et al.* Food-derived mutagens and carcinogens. *Cancer Res* 1992;**52**:2092s–8s.
- Wogan GN, Hecht SS, Felton JS *et al.* Environmental and chemical carcinogenesis. *Semin Cancer Biol* 2004. DOI: 10.1016/j.semcancer.2004.06.010
- Aktipis CA, Nesse RM. Evolutionary foundations for cancer biology. *Evol Appl* 2013;**6**:144–59.
- Ewald PW, Swain Ewald HA. Toward a general evolutionary theory of oncogenesis. *Evol Appl* 2013. DOI: 10.1111/eva.12023
- Klasing KC. Nutrition and the immune system. *Br Poult Sci* 2007;**48**:525–37.
- Hasselquist D, Nilsson JÅ. Physiological mechanisms mediating costs of immune responses: what can we learn from studies of birds? *Anim Behav* 2012;**83**:1303–12.

40. Yang D-B, Xu Y-C, Wang D-H *et al.* Effects of reproduction on immunosuppression and oxidative damage, and hence support or otherwise for their roles as mechanisms underpinning life history trade-offs, are tissue and assay dependent. *J Exp Biol* 2013;**216**:4242–50.
41. Tkachenko H, Kurhaluk N, Grudniewska J *et al.* Tissue-specific responses of oxidative stress biomarkers and antioxidant defenses in rainbow trout *Oncorhynchus mykiss* during a vaccination against furunculosis. *Fish Physiol Biochem* 2014;**40**:1289–300.
42. Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell* 2010;**140**:883–99.
43. Ramiłowski JA, Goldberg T, Harshbarger J *et al.* A draft network of ligand-receptor-mediated multicellular signalling in human. *Nat Commun* 2015;**6**:DOI: 10.1038/ncomms8866
44. Capp JP. Tissue disruption increases stochastic gene expression thus producing tumors: cancer initiation without driver mutation. *Int J Cancer* 2017;**140**:2408–13.
45. Capp JP, Bataille R. Multiple myeloma exemplifies a model of cancer based on tissue disruption as the initiator event. *Front Oncol* 2018;**8**:355.
46. Smith-Ramesh LM, Moore AC, Schmitz OJ. Global synthesis suggests that food web connectance correlates to invasion resistance. *Global Change Biol* 2017;**23**:465–73.
47. Trigos AS, Pearson RB, Papenfuss AT *et al.* Altered interactions between unicellular and multicellular genes drive hallmarks of transformation in a diverse range of solid tumors. *Proc Natl Acad Sci U S A* 2017;**114**:6406–11.
48. Vincent M. Cancer: a de-repression of a default survival program common to all cells?: a life-history perspective on the nature of cancer. *Bioessays* 2012;**34**:72–82.
49. Thomas F, Ujvari B, Renaud F *et al.* Cancer adaptations: atavism, de novo selection, or something in between? *Bioessays* 2017. DOI: 10.1002/bies.201700039
50. Alexandrov LB, Nik-Zainal S, Wedge DC *et al.* Deciphering signatures of mutational processes operative in human cancer. *Cell Rep* 2013;**3**:246–59.
51. Capp JP. Stochastic gene expression, disruption of tissue averaging effects and cancer as a disease of development. *Bioessays* 2005;**27**:1277–85.
52. Paget S. The distribution of secondary growths in cancer of the breast. *Lancet* 1889;**133**:571–3.
53. Fidler IJ. The pathogenesis of cancer metastasis: the “seed and soil” hypothesis revisited. *Nat Rev Cancer* 2003;**3**:453–8.
54. de Groot AE, Roy S, Brown JS *et al.* Revisiting seed and soil: examining the primary tumor and cancer cell foraging in metastasis. *Mol Cancer Res* 2017;**15**:361–70.
55. Hart IR, Fidler IJ. Role of organ selectivity in the determination of metastatic patterns of b16 melanoma. *Cancer Res* 1980;**40**:2281–7.
56. Tarin D, Price JE, Kettlewell MGW *et al.* Mechanisms of human tumor metastasis studied in patients with peritoneovenous shunts. *Cancer Res* 1984;**44**:3584–92.
57. Riihimäki M, Hemminki A, Sundquist J *et al.* Patterns of metastasis in colon and rectal cancer. *Sci Rep* 2016;**6**:29765.
58. Pfeifer GP. Environmental exposures and mutational patterns of cancer genomes. *Genome Med* 2010. DOI: 10.1186/gm175
59. Ling S, Hu Z, Yang Z *et al.* Extremely high genetic diversity in a single tumor points to prevalence of non-Darwinian cell evolution. *Proc Natl Acad Sci U S A* 2015;**112**:1519556112.
60. Waclaw B, Bozic I, Pittman ME *et al.* Spatial model predicts dispersal and cell turnover cause reduced intra-tumor heterogeneity. *Nature* 2015;**525**:261–7.
61. Campbell LL, Polyak K. Breast tumor heterogeneity: cancer stem cells or clonal evolution? *Cell Cycle* 2007;**6**:2332–8.
62. Gerlinger M, Swanton C. How Darwinian models inform therapeutic failure initiated by clonal heterogeneity in cancer medicine. *Br J Cancer* 2010;**103**:1139–43.
63. Lloyd MC, Cunningham JJ, Bui MM *et al.* Darwinian dynamics of intratumoral heterogeneity: not solely random mutations but also variable environmental selection forces. *Cancer Res* 2016;**76**:3136–44.
64. Verduzco D, Lloyd M, Xu L *et al.* Intermittent hypoxia selects for genotypes and phenotypes that increase survival, invasion, and therapy resistance. *PLoS One* 2015. DOI: 10.1371/journal.pone.0120958
65. Chen A, Sceneay J, Gödde N *et al.* Intermittent hypoxia induces a metastatic phenotype in breast cancer. *Oncogene* 2018. DOI: 10.1038/s41388-018-0259-3
66. Patel AP, Tirosh I, Trombetta JJ *et al.* Single-cell RNA-seq highlights intratumoral heterogeneity in primary glioblastoma. *Science* 2014;**344**:1396–401.
67. Amir EAD, Davis KL, Tadmor MD *et al.* ViSNE enables visualization of high dimensional single-cell data and reveals phenotypic heterogeneity of leukemia. *Nat Biotechnol* 2013;**31**:545–52.
68. Efroni S, Duttagupta R, Cheng J *et al.* Global transcription in pluripotent embryonic stem cells. *Cell Stem Cell* 2008;**2**:437–47.
69. Richard A, Boullu L, Herbach U *et al.* Single-cell-based analysis highlights a surge in cell-to-cell molecular variability preceding irreversible commitment in a differentiation Process. *PLoS Biol* 2016;**14**:DOI: 10.1371/journal.pbio.1002585
70. Moussy A, Cosette J, Parmentier R *et al.* Integrated time-lapse and single-cell transcription studies highlight the variable and dynamic nature of human hematopoietic cell fate commitment. *PLoS Biol* 2017;**15**:DOI: 10.1371/journal.pbio.2001867
71. Ohnishi Y, Huber W, Tsumura A *et al.* Cell-to-cell expression variability followed by signal reinforcement progressively segregates early mouse lineages. *Nat Cell Biol* 2014;**16**:27–37.
72. Peláez N, Gavaldà-Mirallas A, Wang B *et al.* Dynamics and heterogeneity of a fate determinant during transition towards cell differentiation. *Elife* 2015;**4**:DOI: 10.7554/eLife.08924
73. Featherstone K, Hey K, Momiji H *et al.* Spatially coordinated dynamic gene transcription in living pituitary tissue. *Elife* 2016;**5**:DOI: 10.7554/eLife.08494
74. Walkley CR, Olsen GH, Dworkin S *et al.* A microenvironment-induced myeloproliferative syndrome caused by retinoic acid receptor gamma deficiency. *Cell* 2007;**129**:1097–110.
75. Raaijmakers MHGP, Mukherjee S, Guo S *et al.* Bone progenitor dysfunction induces myelodysplasia and secondary leukaemia. *Nature* 2010;**464**:852–7.
76. Kode A, Manavalan JS, Mosialou I *et al.* Leukaemogenesis induced by an activating β -catenin mutation in osteoblasts. *Nature* 2014;**506**:240–4.
77. Teschendorff AE, Gao Y, Jones A *et al.* DNA methylation outliers in normal breast tissue identify field defects that are enriched in cancer. *Nature Communications* 2016;**7**:10478.
78. Abdalla M, Tran-Thanh D, Moreno J *et al.* Mapping genomic and transcriptomic alterations spatially in epithelial cells adjacent to human breast carcinoma. *Nature Communications* 2017;**8**:1245.

79. Tyekucheva S, Bowden M, Bango C *et al.* Stromal and epithelial transcriptional map of initiation progression and metastatic potential of human prostate cancer. *Nature Communications* 2017; doi:10.1038/s41467-017-00460-4.
80. Flavahan WA, Gaskell E, Bernstein BE. Epigenetic plasticity and the hallmarks of cancer. *Science* 2017;**357**:eaal2380.
81. Salas LA, Johnson KC, Koestler DC *et al.* Integrative epigenetic and genetic pan-cancer somatic alteration portraits. *Epigenetics* 2017;**12**: 561–74.
82. Brocks D, Assenov Y, Minner S *et al.* Intratumor DNA methylation heterogeneity reflects clonal evolution in aggressive prostate cancer. *Cell Reports* 2014;**8**:798–806.
83. Oakes CC, Claus R, Gu L *et al.* Evolution of DNA methylation is linked to genetic aberrations in chronic lymphocytic leukemia. *Cancer Discovery* 2014;**4**:348–61.
84. Landau DA, Clement K, Ziller MJ *et al.* Locally disordered methylation forms the basis of intratumor methylome variation in chronic lymphocytic leukemia. *Cancer Cell* 2014;**26**:813–25..