

Research article

New fossil genus and new extant species of diatoms (Stephanodiscaceae, Bacillariophyceae) from Pleistocene sediments in the Neotropics (Guatemala, Central America): adaptation to a changing environment?

Christine PAILLÈS^{1,*}, Florence SYLVESTRE², Alain TONETTO³,
 Jean-Charles MAZUR⁴ & Sandrine CONROD⁵

^{1,2,4,5} Aix-Marseille Université, CNRS, IRD, Collège de France, INRAE, CEREGE,
 13545 Aix-en-Provence cedex 4, France.

³ Aix-Marseille Université, Fédération de Chimie, PRATIM, 3 Place Victor Hugo,
 13331 Marseille Cedex 3, France.

* Corresponding author: pailles@cerege.fr

² Email: sylvestre@cerege.fr

³ Email: alain.tonetto@univ-amu.fr

⁴ Email: mazur@cerege.fr

⁵ Email: conrod@cerege.fr

Abstract. Several taxa of Stephanodiscaceae were found in the upper section of Pleistocene sediments from Lake Petén-Itzá (Guatemala). A new fossil genus *Cyclocostis* Paillès gen. nov. and new extant species *Discostella gabinii* Paillès & Sylvestre sp. nov. are described. *Cyclocostis* gen. nov. is characterized by a strongly tangentially undulated valve surface, coarse unequal striation reaching a central punctum in the valve center, an absence of central lamina and domed criba, widely open alveoli with one median recessed costa bearing marginal fultoportulae and a single rimoportula all within a ring. A single valve face fultoportula is present on the raised part of the valve opposite the rimoportula. Differences relative to similar genera and the delimitation of a new genus are discussed. *Discostella gabinii* sp. nov. is distinguished by circular and flat valves, a small central area bearing 5 to >30 scattered large areolae giving a colliculate appearance, medium-sized alveoli, marginal fultoportulae on every 4–5th costa, a single rimoportula and internally smooth valve center. Differences to similar taxa in the genus *Discostella* are discussed. The succession of the species of *Cyclotella*, *Discostella* and *Cyclocostis* gen. nov. in our record could represent eco-phenotypic responses to particular environmental stress/change.

Keywords. Diatom, Stephanodiscaceae, genus, *Cyclocostis rolfii*, *Discostella gabinii*, Pleistocene, Guatemala.

Paillès C., Sylvestre F., Tonetto A., Mazur J.-C. & Conrod S. 2020. New fossil genus and new extant species of diatoms (Stephanodiscaceae, Bacillariophyceae) from Pleistocene sediments in the Neotropics (Guatemala, Central America): adaptation to a changing environment? *European Journal of Taxonomy* 726: 1–23. <https://doi.org/10.5852/ejt.2020.726.1169>

Introduction

Publications describing freshwater diatoms from Central America are scarce. If few paleolimnological studies based on diatoms have been conducted in Costa Rica (e.g., Chávez & Haberyan 1996; Haberyan & Horn 1999, 2005), Nicaragua (e.g., Slate *et al.* 2013) and Panama (e.g., Temoltzin-Loranca *et al.* 2018), even fewer have been conducted in the Yucatan Peninsula, and these are generally limited to the Holocene (Whitmore *et al.* 1996; Rosenmeier *et al.* 2004). Longer diatom records originated from the Mexico Basin (Bradbury 2000; Ortega *et al.* 2010). Only the record of Cohuo *et al.* (2018) is based on the same sedimentary sequence as the present study. Furthermore, since the majority of fresh water bodies in this region are alkaline, calcium-bicarbonate dominated systems (Pérez *et al.* 2013), valve preservation is usually poor (Metcalf *et al.* 2000). A few taxonomic studies have been recently published on diatom species from, e.g., Panama (Lange-Bertalot & Metzeltin 2009) or El Salvador (Wetzel & Ector 2014; Krahn *et al.* 2018), with one dealing with centric diatom species of *Cyclotella* (Kütz.) Bréb. from Guatemala (Paillès *et al.* 2018).

In the last 50 years, the genus *Cyclotella* (Brébisson 1838: 19) has received significant attention. There have been many attempts to classify species within the genus according to the morphological features by Lowe (1975), McFarland & Collins (1978), Serieyssol (1981), Servant-Vildary (1986), Loginova (1990), Håkansson (1990), Håkansson *et al.* (1993) and Tanaka (2007). In a revision of the genus *Cyclotella*, Håkansson (2002) subdivided it on the basis of the type of undulation of the valve, the morphology of the central area, the position of the rimoportula, and the position and number of satellite pores of marginal fultoportulae. This author also suggested that the number of satellite pores of marginal fultoportulae may be the most important character for defining phylogenetic groups within *Cyclotella*. Interestingly, Loginova (1990) observed that only fossil species of *Cyclotella* have marginal fultoportulae with three satellite pores. Under Håkansson's (2002) assumption, Prasad & Nienow (2006) identified one lineage of *Cyclotella* with three satellite pores and another with two satellite pores. Similarly, using the presence/absence of a central lamina (Servant-Vildary 1986) and the presence/absence of marginal chambers (Lange & Syversten 1989), the authors identified three groups of *Cyclotella*. Khursevich & Kociolek (2012) distinguished 11 groups of *Cyclotella* based on the structure of alveolae, the number and location of rimoportula, and the structure of striae. For Houk *et al.* (2010), the genus *Cyclotella* is subdivided into three groups. A revision of the classification of *Cyclotella* by Nakov *et al.* (2015) identified the position of the rimoportula as a synapomorphy. *Cyclotella* s. str. has a rimoportula located on a costa within the ring of marginal fultoportulae, as opposed to those of *Discostella* Houk & Klee, *Lindavia* (F.Schütt) De Toni & Forti and *Paleotertiarius* (Håk. & Khursevich) S.Blanco (Nakov *et al.* 2015). Ács *et al.* (2016) separated the genera *Lindavia*, *Pantocsekiella* K.T.Kiss & Ács and *Edtheriotia* Kociolek, You, Stepanek, R.L.Lowe & Wang from the genus *Cyclotella* on the basis of morphological and genetic characters. At this point, these new concepts make it difficult to disentangle the phylogeny and taxonomy of this group.

In 2006, the Petén-Itzá Scientific Drilling Project (PISDP) recovered a total of 1327 m of sediment from seven sites in Lake Petén-Itzá, Petén, northern Guatemala, for paleoclimate and paleoenvironmental studies (Hodell *et al.* 2008). The fossil diatom flora of Lake Petén-Itzá was analyzed in core PI-6 dated by tephro-chronology and covering the last 84 ka (Kutterolf *et al.* 2016). From this record, the new species *Cyclotella petenensis* Sylvestre, Paillès & Escobar and *C. cassandrae* Paillès & Sylvestre were described (Paillès *et al.* 2018). Based only on morphological observations, we herein describe a new genus belonging to the family Stephanodiscaceae Glezer & Makarova, *Cyclocostis* Paillès gen. nov., and a new species of *Discostella* from the same core, bringing the total to three new species and one new genus described from the Pleistocene sediments of Lake Petén-Itzá. So far, except for *C. cassandrae* and the new genus *Cyclocostis* gen. nov. (described herein) that are absent from the modern dataset (Pérez *et al.* 2013), the majority of species in the fossil record are still extant and will therefore provide a solid basis for paleolimnological reconstructions.

Material and methods

During the International Continental Scientific Drilling Program (ICDP) expedition PISDP in 2006, sediment cores were collected at site PI-6 in Lake Petén-Itzá (Hodell *et al.* 2008), providing a continuous record of sediment accumulation for the last 84 cal ka. Cores were stored on board at 5°C, then transferred to the Minneapolis Core-Repository in Minnesota (USA) where samples were taken from depths ranging between 0 and 70 mcd (meter composite depth) at 10-cm intervals. The core was dated by ^{14}C (44 ages) on terrestrial remains and three tephra layers dated by Ar/Ar (Escobar *et al.* 2012). An age model was derived using a weighted fit through 36 age-depth points. The sediments consisted of laminated light brown to greenish clays, gypsum and tephra layers (for more details see Hodell *et al.* 2008).

A 5 mm thick slice of sediment (0.5 g) was first decarbonated with hot 37% HCl for a few hours, the organic matter was then oxidized with hot 33% H_2O_2 for a few hours. Diatom suspensions were successively rinsed and decanted with distilled water. Aliquots of cleaned diatoms were diluted and evaporated at room temperature onto coverslips then mounted on glass slides using Naphrax[®] mounting medium. Diatom slides were examined under oil immersion using light microscopy (LM) at 630 $\times$ or 1000 $\times$ magnification using a Nikon Eclipse 80i microscope equipped with differential interference contrast optics and a Nikon D300 camera. Counting was generally performed on three slides. The total number of valves counted per sample varied from 200 in nearly sterile samples to > 1200 in rich samples. Diatom identification and taxonomy followed Krammer & Lange-Bertalot (1986, 1988, 1991a, 1991b) with the revised nomenclature in AlgaeBase (Guiry & Guiry 2020). Number of striae in 10 μm were determined following Genkal (1977), i.e., as the number of striae in 10 μm of the valve circumference, not in 10 μm of a transect (chord).

For scanning electron microscopy (SEM) observations, a few drops of cleaned diatom material were air-dried on circular coverslips which were then attached to aluminum stubs and gold-coated with Cressington 108 auto (Watford, UK). Diatoms were examined with a XL 30 ESEM Philips SEM at an accelerating voltage of 1–30 kV.

Taxonomic descriptions followed terminology given by Anonymous (1975), Ross *et al.* (1979), Theriot & Serreyssol (1994), Håkansson (2002), Tanaka (2007), Houk *et al.* (2010, 2014) and Khursevich & Kociolek (2012).

Abbreviations

LM = light microscopy
 mfp = marginal fulcrum
 rm = rimoportula
 SEM = scanning electron microscopy
 vffp = valve face fulcrum

Results

Sediments from core PI-6 revealed a rich and diverse diatom flora (153 species belonging to 42 genera). Ninety-six species are extant and were found in diverse waterbodies from the Yucatan Peninsula (Pérez *et al.* 2013). The upper part of the sedimentary sequence between 60–16 ka is characterized by an alternation of the species of *Cyclotella* and *Discostella* (Fig. 1): *Cyclotella meneghiniana* Kütz., *C. petenensis*, *Discostella stelligera* (Cleve & Grunow) Houk & Klee, *C. caspia* Grunow, *C. cassandrae*, a new species of *Discostella*, and what appears to be a new genus of Stephanodiscaceae.

Taxonomic treatment

Division Bacillariophyta G.Karst.
Class Mediophyceae Medlin & Kaczmarska
Order Thalassiosirales Glezer & Makarova
Family Stephanodiscaceae Glezer & Makarova

Genus *Cyclocostis* Paillès gen. nov.

Type species

Cyclocostis rolfii Paillès gen et sp. nov. (see below).

Diagnosis

A new morphotypic genus of Stephanodiscaceae. Cells solitary, circular in valve view. Marginal and central areas coarsely striated with branching striae merging in the middle of the valve into a central punctum. Central area strongly tangentially undulated, focus clear only on half the central area. Submarginal alveoli finely punctuated, transforming into radiating rows of large irregular areolae in the central area. Internally, alveoli delineated by thick elevated costae reaching the valve center to form a subcircular silica ring. Alveoli widely open with a median recessed costa bearing marginal fultoportulae. Central lamina absent. Single rimoportula within a ring of marginal fultoportulae reduced to a rounded tube tangentially orientated and born on a recessed costa. On the mantle, outer expressions of marginal fultoportulae and rimoportula consisting of simple rounded openings without projections. One single valve face fultoportula consisting of a central tube surrounded by three satellite pores that opens externally into a round opening on the raised part. Cingulum present consisting of an open valvocopula and several copulae.

Etymology

The genus name refers to the circular morphology of the valve and marked radiating costae.

Cyclocostis rolfii Paillès gen et sp. nov.
Figs 1–40

Etymology

This species is named in honor of Rolf Klee for his dedicated career on Stephanodiscaceae.

Type material

Holotype

Slide PC0608731 and sediment PC0608728 deposited at the Laboratoire de Cryptogamie, Muséum national d'histoire naturelle (MNHN) Paris, France. Specimen on slide PC0608731 (Fig. 4) represents the holotype designated here.

Isotype

Slide ZU 11/30 and sediment R1284 deposited at the Friedrich Hustedt Diatom Center in Bremerhaven, Germany.

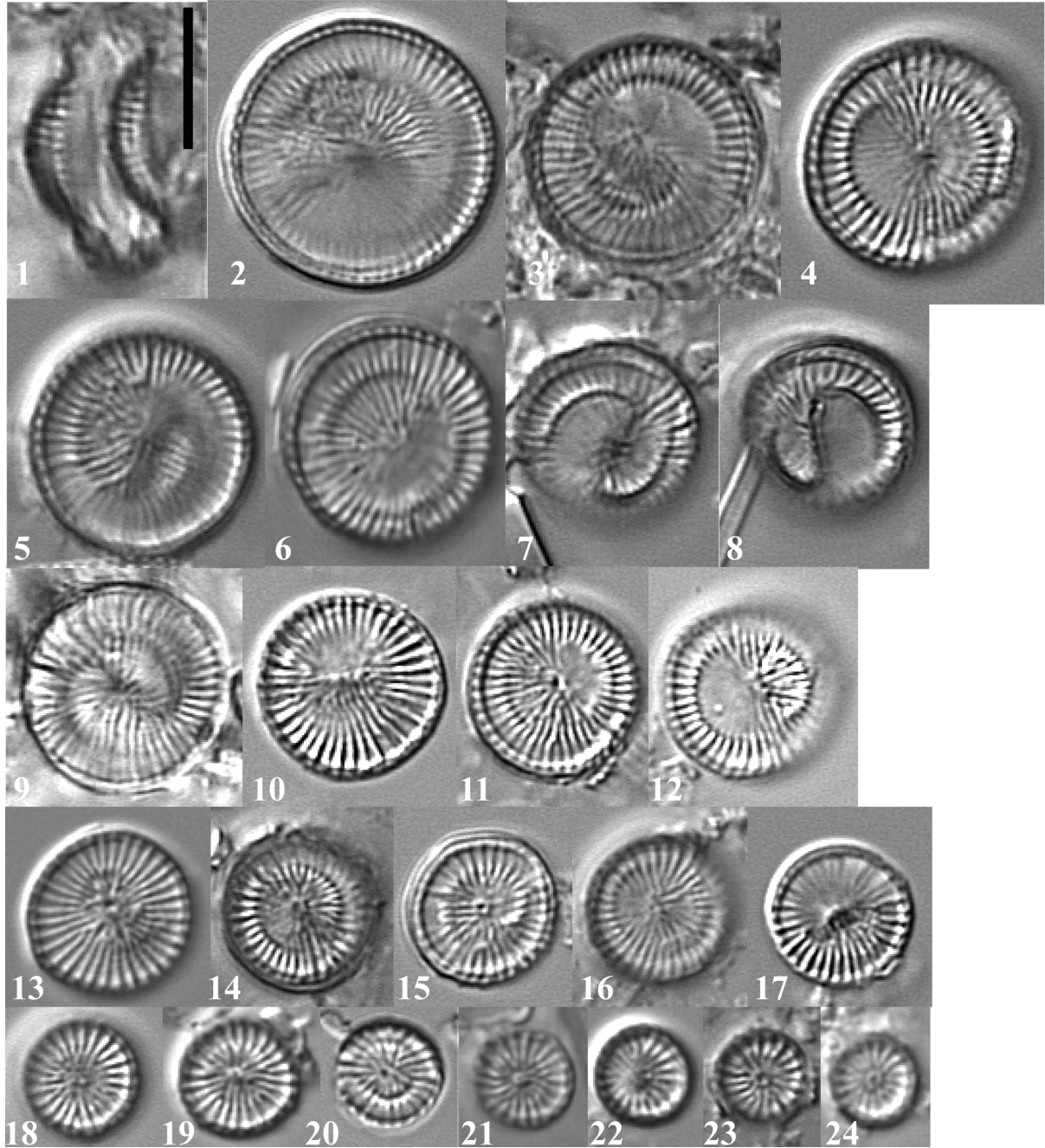
Type locality

GUATEMALA • Department of Petén, Lake Petén-Itzá; 16°15'50" N, 89°15'00" W; lacustrine sediment in core PI-06; sample GLAD9-PET06-6B-10H1- 98–99 cm (27.83 m below lake floor) consisting of light gray carbonated sediment; core collected in February 2006.

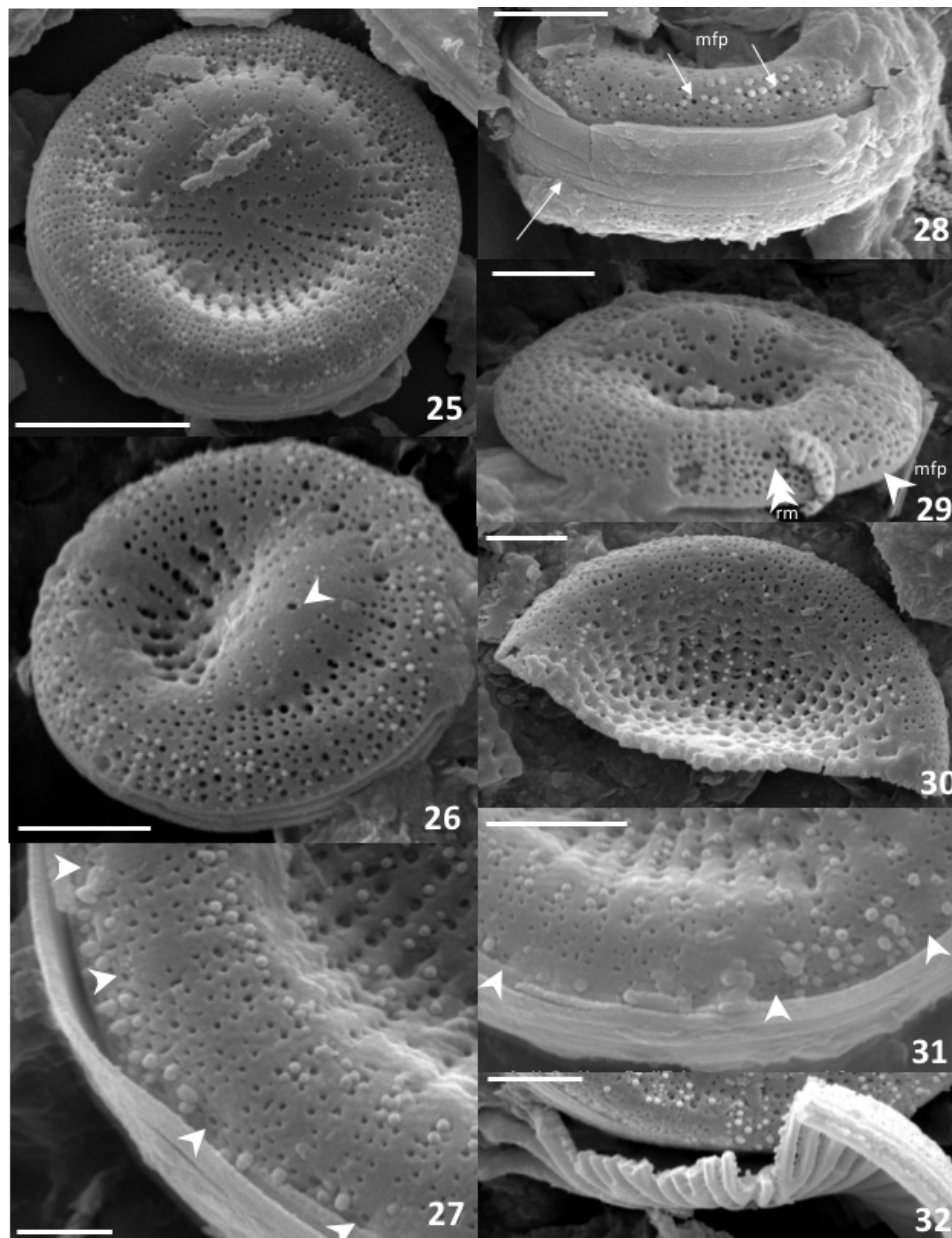
Description

Light microscopy (Figs 1–24)

In girdle view, cells quadrangular, displaying strongly undulated valve faces (Fig. 1). In valve view, frustules circular, 7–22 µm in diameter (Figs 2–24). Marginal area (outer 1/3 of valve) concentrically undulated and consisting of an external ring of small bright ‘chambers’ transforming into strong radiating costae – 10 to 12 striae in 10 µm. The central 2/3 of valve surface strongly tangentially undulated, forming an S shape in large specimens (Figs 7–8). The central area with radial anastomosing striae of unequal



Figs 1–24. Type material of fossil lacustrine diatom *Cyclocostis rolfii* Paillès gen. et sp. nov., Lake Petén-Itzá (Guatemala); LM girdle view (1) and valve views (2–24). **4.** Holotype (MNHN, slide PC0608731). **7–8.** Valve surface strongly tangentially undulated, forming an S shape. Scale bar = 10 µm.



Figs 25–32. Type material of fossil lacustrine diatom *Cyclocostis rolfii* Paillès gen. et sp. nov.; SEM external valve views. **25.** Valve view of concentrically undulated marginal area and tangentially undulated central area; punctuated striae become in the central area rows of larger areolae arranged in a stellate pattern. **26.** Valve surface with scattered papillae; the external opening of the single valve face fultoportula is located on the raised part (white arrowhead). **27.** Marginal area showing the external openings of marginal fultoportulae, collared but with no projections (white arrowheads). **28.** Side view of marginal area showing striation, papillae, external openings of marginal fultoportulae (mfp – two white arrows), and the cingulum consisting of an open valvocopula and several copulae (white arrow). **29.** Detail of the central area with large areolae; external areolae are bigger and occluded by volae in places where ribs are fusing. **30.** Broken valve view showing the different striation between the margins and the center, the steep transversal undulation and the valve thickness. **31.** Marginal area with the external openings of marginal fultoportulae (white arrowheads), papillae, and the cingulum. **32.** Broken valve view showing the simple structure of anastomosing ribs covered by a finely perforated silica layer. Scale bars: 25 = 5 μm ; 26, 28–32 = 2 μm ; 27 = 1 μm (27).

length, some extending deep in the central zone to a central bright punctum. Marginal and central areas not distinctively structured. In small specimens, strong transversal undulation is attenuated and radiating striae are converging to a central ring.

Scanning electron microscopy (Figs 25–40)

Valves strongly undulated externally (Figs 25–26). Cingulum present, consisting of an open valvocopula and several copulae (Figs 27–28). Valve surface is irregular with sprinkled granules and prominent embossed ribs. The marginal striated area is circumferentially undulated with numerous granules; the mantle is gently sloping (Fig. 27). Striae consisting of 3–4 rows of finely aligned areolae alternating with reduced hyaline interstriae bearing near the valve margin big rounded openings corresponding to the external openings of marginal fuloportulae (Figs 27–28). The central area displays a steep tangential undulation (Fig. 29). If the junction between the valve face and the mantle is steeply marked vertically both on the elevated and depressed sides (Fig. 30), horizontally, from the elevated to depressed parts, the incline is smooth and gradual. On the internal side of the marginal area, striae become single rows of large rounded to oblong areolae that progressively become smaller and arranged to some extent into a stellate pattern (Fig. 31). Where ribs are fusing, external areolae are bigger and occluded by volae. Broken specimen displays a simple valve structure: a basal siliceous layer composed of anastomosing ribs starting from a central hollow and continuing to the valve rim where intercostal spaces are covered by a finely perforated silica layer (Fig. 32).

Internally, there is no central lamina inside the valve (Fig. 33). Costae are strongly silicified and elevated, extending from the valve rim to the valve center and fusing into a thick silicified hollow (Figs 34–35). The alveolar structure could be classified as complex as the alveolus bears in its middle a recessed/sunken costa that carries the marginal fuloportula (mfp) (Fig. 36). As such, mfp are located on every second striae and are composed of one tube and three satellite pores (Figs 37–38). The ring of mfp and rm stands just beneath marginal lamina. One single rimoportula (rm) positioned on a recessed costa consisting of a short tube with a tangential slit that is always diametrically opposed to the raised side (Fig. 39). One single valve face fuloportula (vffp) – composed of one tube and three satellite pores – is always diametrically opposed to the rimoportula (Fig. 39). It appears eccentric since positioned on the raised part (Fig. 40). The external opening of the vffp is difficult to observe as it is positioned on the external slope of the raised central part (see in Fig. 26).

Time range

23–28 ka, abruptly absent after, considered extinct.

Remarks

Cyclocostis rolfii gen et sp. nov. belongs unequivocally to the family Stephanodiscaceae (Glezer & Makarova 1986). In LM, it resembles *Discostella woltereckii* (Hust.) Houk & Klee in Klee & Houk (1996). However, SEM observations reveal a completely different structure in *C. rolfii* gen et sp. nov., particularly in the position of mfp and rm on costae, ruling out its belonging to the genus *Discostella*. With alternating and unequal striation pattern on the valve face, *C. rolfii* gen et sp. nov. also resembles *Cyclotella stoermeri* Khursevich & Kociolek in Kociolek & Khursevich (2013) in LM. Even though they share the absence of a central area, unequal striation, reduced alveoli and position of mfp and rm on recessed costae, *C. rolfii* gen et sp. nov. differs by the absence of centripetal roofing (central lamina) and thus the alveoli have no distinct border at the valve center side. The unequal striation visible in LM in *C. rolfii* gen et sp. nov. only corresponds to the way ribs are arranged internally: elevated, strongly silicified, anastomosing and joining in a central hollow. These thick radial ribs are the external characteristic features of the genus *Stephanocostis* Genkal & Kuzmina represented by *S. chantaica* Genkal & Kuzmina. However, these structures are internal in *C. rolfii* gen et sp. nov., whereas they are

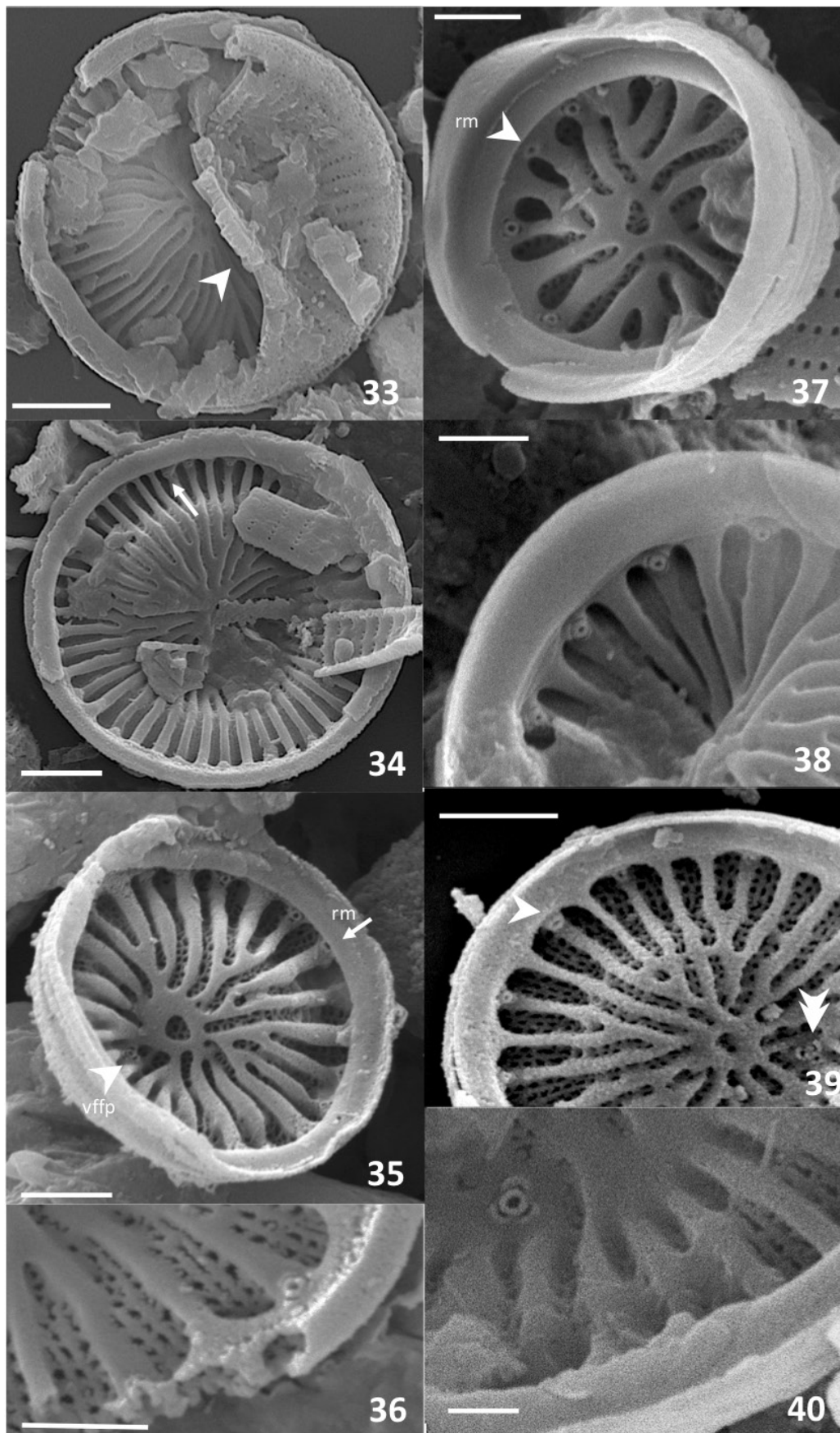
external in *S. chantaica* whose internal structure is flat with fine areolae forming $\pm$ radial rows of star-like criba.

The strongly tangentially undulated valve is a character shared by *Cyclotella* and *Pliocaenicus* Round & Håk., the latter genera being included into *Lindavia* by Nakov *et al.* (2015). The position and structure of mfp are also a common character. But unlike in some *Cyclotella* or *Pliocaenicus*, no domed criba are observed internally in *Cyclocostis* gen. nov. As the position of the rm within the ring of mfp in *C. rolfii* gen et sp. nov. differs from *Pliocaenicus*, it rules out its belonging to this genus.

The alveoli of *C. rolfii* gen et sp. nov. could be classified as complex, because there is a median fulcrum born on a recessed costa. However, structurally they are simple since not partially occluded by central lamina and widely open towards the valve center. The structure of alveoli of *C. rolfii* gen et sp. nov. rather corresponds to some extent to the marginal chambers defined by Lange & Syvertsen (1989) as “a marginal space characterized by an opening on the inside of the valve encompassing two or more alveolus openings that is limited by coarse interstriae”. These marginal chambers are present in *Cyclostephanos novaezeelandiae* (Cleve) Round in Theriot *et al.* (1987) or *C. dubius* (Hust.) Round in Theriot *et al.* (1987). However, the genus *Cyclostephanos* Round is largely heterogenous in terms of frustule morphology as it includes species with and without alveolar chambers. Except the marginal chambers, *C. rolfii* gen et sp. nov. shares no other characters with the genus *Cyclostephanos*.

The characteristics of the striae, composed of fine pori on the mantle and becoming uniseriate with large radially arranged areolae towards the center of the valve, as observed in *C. rolfii* gen et sp. nov., are shared with *Paleotertiarius*. Even though *C. rolfii* gen et sp. nov. and *Paleotertiarius* share characters such as strong ribs and the structure and position of mfp, in *Paleotertiarius*, flat or concentrically undulated valves, internal domed criba and rimoportula located on the side of a costa inside the alveolus are morphological differences that exclude *C. rolfii* gen et sp. nov. from belonging to *Paleotertiarius*.

Figs 33–40 (next page). Type material of fossil lacustrine diatom *Cyclocostis rolfii* Paillès gen. et sp. nov.; SEM internal valve views. **33.** Broken valve revealing internal structure; note the reduced cell cavity (white arrowhead). **34.** Valve interior showing strong anastomosing ribs merging to a central siliceous ring; central lamina absent; marginal fulcrum located on every 2nd recessed costa; note the slightly deflected rimoportula (white arrow). **35.** Valve interior of a small specimen with strong thick ribs and reduced alveoli, central lamina absent; note the single valve face fulcrum (white arrowhead) located near the valve margin on the raised central area; open valvocopula in place. **36.** Broken valve margin showing complex alveolar structure: two thick ribs with a median recessed costa, more inwardly bent, carrying marginal fulcrum composed of one tube and three satellite pores. **37.** Valve interior of a small specimen with valvocopula. **38.** Heart shaped alveoli due to the marginal fulcrum located on median recessed costa. **39.** Internal valve view with marginal fulcrum and rimoportula arranged in a ring below the valve margin; the single valve face fulcrum (double white arrowhead) is always on the opposite side from the rimoportula (single white arrowhead). **40.** Detail of the single valve face fulcrum composed of a short tube and three satellite pores always in an eccentric position. Scale bars: 33, 34 = 5 μ m; 35, 39 = 2 μ m; 36–38 = 1 μ m; 40 = 0.5 μ m.



Genus *Discostella* Houk & Klee

Discostella gabinii Paillès & Sylvestre sp. nov.

Figs 41–58

Etymology

This taxon is named in honor of deceased Gabin Sylvestre, the courageous 7 years old nephew of F. Sylvestre.

Type material

Holotype

Slide PC0608732 and sediment PC0608730 deposited at the Laboratoire de Cryptogamie, Muséum national d'histoire naturelle (MNHN) Paris, France. Specimen on slide PC0608732 (Fig. 45) represents the holotype designated here.

Isotype

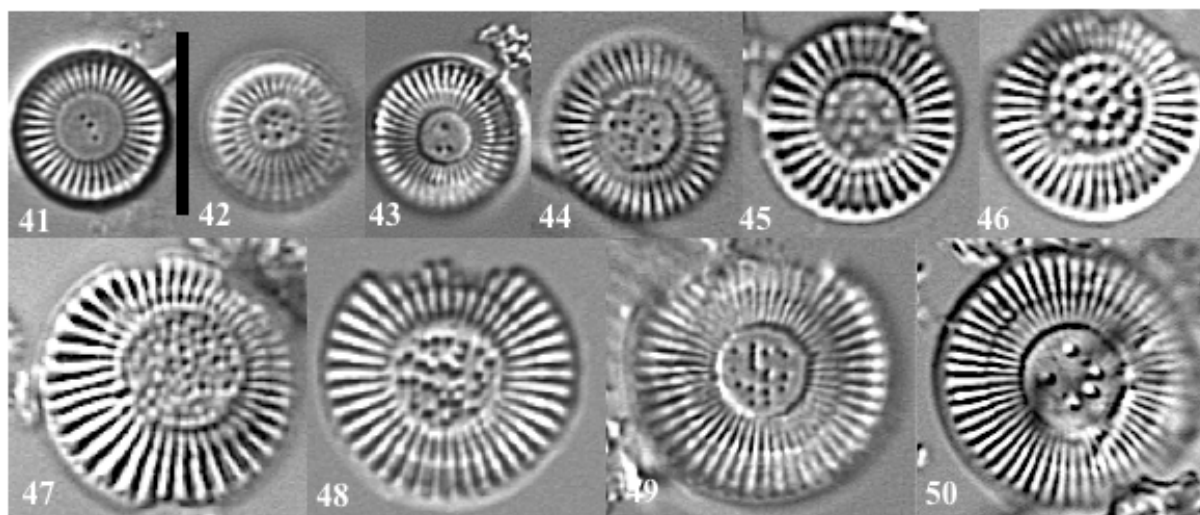
Slide ZU 11/31 and sediment R1285 deposited at the Friedrich Hustedt Diatom Center in Bremerhaven, Germany.

Other material examined

Modern specimens collected from Cenote Juarez and Lake Amatitlan (see Table 1).

Type locality

GUATEMALA • Department of Petén, Lake Petén-Itzá; 16°15'50" N, 89°15'00" W; lacustrine sediment in core PI-06; sample GLAD9-PET06-6B-18E1- 35.4–36.4 cm (51.53 m below lake floor) consisting of dark gray clayish sediment; core collected in February 2006.



Figs 41–50. *Discostella gabinii* Paillès & Sylvestre sp. nov., Lake Petén-Itzá (Guatemala); LM valve views. **41–42.** Modern specimens of *D. gabinii* sp. nov. from Cenote Juarez. **43–44.** Modern specimens of *D. gabinii* sp. nov. from Lake Amatitlan. **45–50.** Type material of fossil lacustrine diatom *D. gabinii* sp. nov. **45.** Holotype (MNHN, slide PC060873). **48–50.** A shadow line is visible in large specimens. Scale bar = 10 μ m.

Description

Light microscopy (Figs 41–50)

Cells quadrangular in connective view. Valvar views circular and flat, 8–18 µm in diameter with a small central area ($\frac{1}{3}$ of the valve radius). Central area with 5 to >30 scattered large areolae, the number being independent of valve size (Figs 41–50). When numerous, the scattered areolae give the impression of a colliculate/granular flat center. The marginal area of the valve face has radial striae numbering from 10 to 14 in 10 µm. The striae are long ($\frac{2}{3}$ of the valve radius) and of equal length. On large specimens, marginal striation is crossed circumferentially by a ring ('Schattenlinie' = 'shadow line') close to the valve center (Figs 48–50).

Scanning electron microscopy (Figs 51–58)

Valves flat to barely concave externally with gently sloping mantle. Central area covered with several scattered punctae separated by knots (colliculate) bearing papillae (Figs 51–52). Radiating striae starting on the mantle as crescents of three to five rows of fine areolae (60–70 areolae/10 µm), merging into two rows near the central area and ending with a single large pore (Figs 53–54). The central area is thus bordered by a ring of large areolae. On the valve face, striae are depressed, whereas they are smooth on the mantle. Near the valve margin, every third to fifth striae, pores just below the crescent of fine areolae mark the external openings of marginal fultoportulae (Fig. 54). Interstriae are narrow, domed and granular on the valve face, whereas smooth on the mantle. The mantle is unornamented except for the large round openings of marginal fultoportulae and few papillae. The external opening of the rimoportula was not observed, although it should be positioned at the same level since it is within the ring of marginal fultoportulae.

Interior views of the valve show a flat to slightly concave but smooth central area with none or single areola (Figs 55–56). The internal lamina spread from the valve center to $\frac{2}{3}$ of the valve radius. The alveoli are thus medium sized, oblong and of unequal length, those bearing marginal fultoportulae being longer (Fig. 56). Marginal fultoportulae with two laterally positioned satellite pores surrounding a short tubulus are located at the distal extremity of every third to fourth alveoli (Fig. 57). One nearly sessile rimoportula with vertically orientated lips located between two costae at the edge of an alveolus and within the ring of marginal fultoportulae (Fig. 58). Girdle bands present, an open valvocopula with two copulae (Fig. 56); a row of fine pores is noted on the interior of the girdle band (Fig. 58).

Time range

Present since at least 84 ka in the geological record, present in Lake Amatitlan and Cenote Juarez (20°48'09.6" N, 87°27'23.8" W) in March 2008.

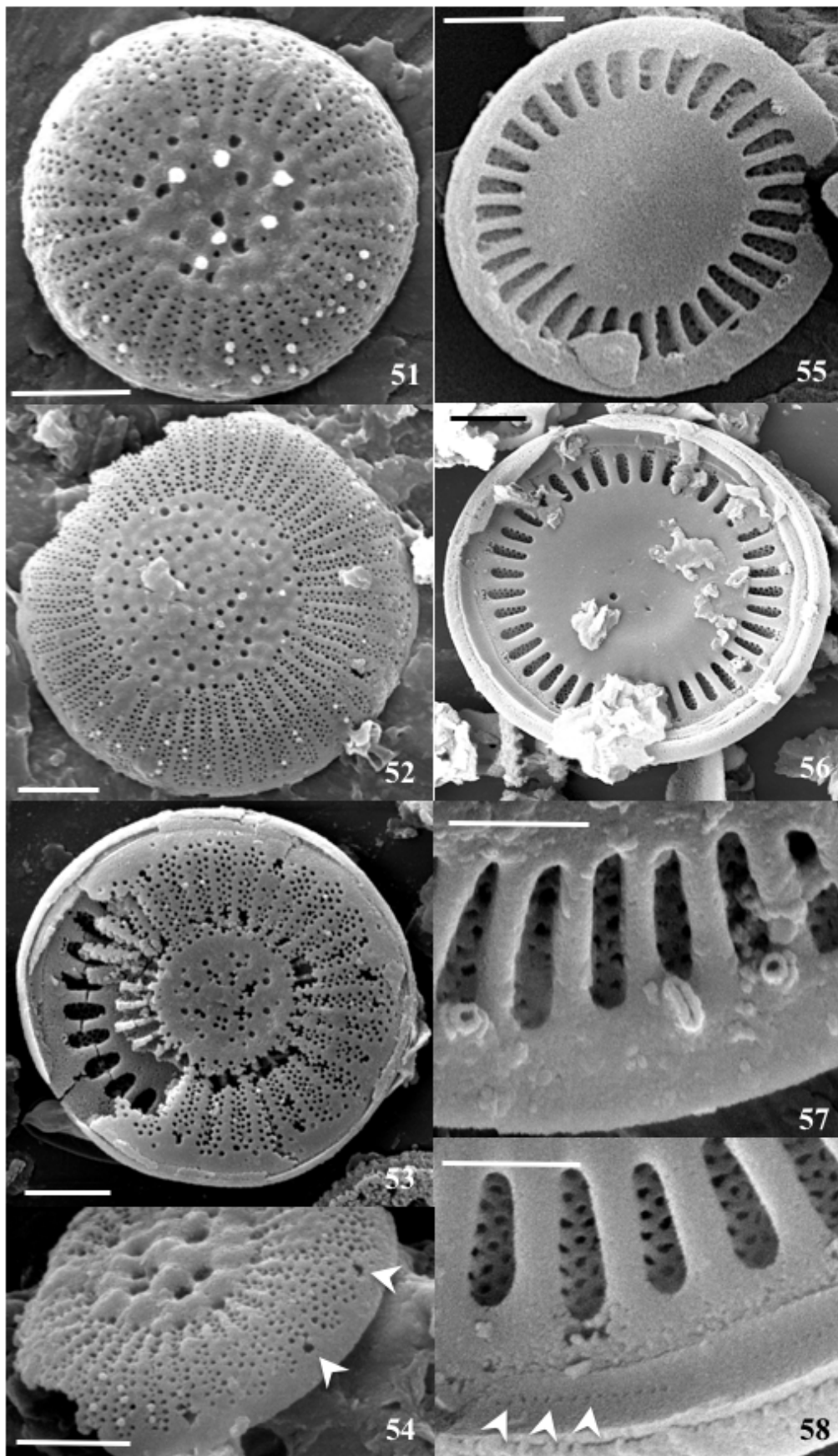
Remarks

With marginal fultoportulae and rimoportula being located between costae on the marginal side of the alveolus, *Discostella gabinii* sp. nov. belongs undoubtedly to the genus *Discostella*. Amongst the 15 species of *Discostella* described so far (Kociolek *et al.* 2018), *D. gabinii* sp. nov. showed some resemblance to *D. areolata* (Hust.) Houk & Klee. However, in LM they look somewhat different, the unique holotype of *D. areolata* having coarser striation (6–9 striae in 10 µm) and a large colliculate central area (Houk *et al.* 2010: table 330, figs 1–7). A reexamination of the original material of *D. areolata* from Hustedt by Tagliaventi & Cavinaci (2002) provided unambiguous SEM images of external views but only ambiguous internal views since *D. areolata* was rare and mixed with *D. stelligera* (Cleve & Grunow) Houk & Klee var. *robusta* (Hust.) Houk & Klee in the original material. The central area of *D. areolata* is concave or convex, smooth or consisting of alternating impressions and protrusions of various size with small punctae being mainly located in the depressions. Sometimes domed radiating striae resembling a poorly defined rosette are present in the central area. In *D. gabinii* sp. nov., the central

area is always flat, indeed colliculate but with large punctae inserted in the depressions. Moreover, in *D. areolata*, striae are depressed and costae elevated on their entire length, whereas in *D. gabinii* sp. nov., this feature is restricted to the valve face, the mantle being smooth. Internally, two types of central area could be attributed to *D. areolata*: smooth with no central fultoportula or smooth with a punctum. These variations are also visible in *D. gabinii* sp. nov. Despite uncertainties related to the species described as *D. areolata*, *D. stelligera* var. *robusta* and *D. stelligera* var. *hyalina* (Hust.) Houk & Klee, the structure of marginal costae and the position of marginal fultoportulae and rimoportula are quite different compared to that of *D. gabinii* sp. nov. Marginal costae can be forked or not. Furthermore, marginal fultoportulae (composed of one tube and two satellite pores placed horizontally) and rimoportula (vertically orientated slit) are inserted within the alveolar chamber.

Another somewhat similar species is *D. elentarii* (Alfinito & Tagliaventi) Houk & Klee with flat valves, although it has a large central area with radiate rows of granules and scattered punctae, coarsely striated (9–10 striae in 10µm) and reduced marginal area, and a marginal row of small spinae. Internally, it has similar smooth central area (sometimes with a faint stellate pattern) and similar structure and position of mfp and rm. The only difference is that, internally, in *D. elantarii* costae are broadening toward the valve margin with a punctum in the middle giving the impression of forked costae. After reexamination of *D. elantarii* by Knapp *et al.* (2006), it appears that the correct striae density is 8–14 and that each collared marginal fultoportula and the single rimoportula are surrounded by satellite pores covered by a cribrum. Although we did not use a field emission variable pressure SEM, such structures are absent in *D. gabinii* sp. nov. Interestingly, it is the only morphological feature used to differentiate *D. elantarii* from *D. stelligera* in SEM (Knapp *et al.* 2006). The presence of pores in the girdle band is also a subtle character shared by *D. elentarii* and *D. gabinii* sp. nov. that requires further investigation. Despite morphological similarities with *D. areolata* and *D. elentarii*, *D. gabinii* sp. nov. possesses distinctive characteristics that are sufficient to define a new species. Stelligeroid species of *Cyclotella* have been transferred to the genus *Discostella* on the basis of the unique position of strutted and labiate processes (Houk *et al.* 2010). However, difficulties arise because these species are often heterovalvate and size and morphological variations exist. As reported by Tagliaventi & Cavinaci (2002), Alfinito & Tagliaventi (2002) and Knapp *et al.* (2006), only minute distinctive features allow one to differentiate *D. areolata*, *D. stelligera*, *D. stelligera* var. *robusta*, *D. stelligera* var. *hyalina* and *D. elantarii*. This latter species is endemic to New Zealand and coexists with *D. stelligera* in two lakes. Knapp *et al.* (2006) suggest that considering the difficulty in differentiating them, they could be sibling species and *D. elantarii* may descend from *D. stelligera*.

Figs 51–58 (next page). Type material of fossil lacustrine diatom *Discostella gabinii* Paillès & Sylvestre sp. nov., Lake Petén-Itzá (Guatemala); SEM valve views. **51.** External valve view showing a colliculate central area with scattered areolae, knots and papillae, and a marginal area with finely punctuated radiating striae. **52.** External view of a large specimen with numerous areolae in the central area; the striae have rounded ends near the margin. **53.** Valve view of a complete corroded frustule showing valve interior with medium sized alveoli and marginal fultoportulae located in the middle of every 3rd to 5th alveoli; open valvocopula present. **54.** External openings of marginal fultoportulae located just below the crescent end of the striae (white arrowheads). **55.** Internal valve view with medium sized alveoli and a large expansion of the smooth central lamina; marginal fultoportulae located on every 3rd to 5th alveoli. **56.** Internal valve view with smooth central area with a single areola; open valvocopula present. **57.** Detail view of valve margin; marginal fultoportulae composed of one tube with two satellite pores positioned on the external ends of the alveoli; rimoportula with vertically orientated lips, located between two costae at the edge of an alveolus and within the ring of marginal fultoportulae. **58.** Detail view of the valve margin with an eroded marginal fultoportula at the edge of the alveolus; note a row of fine pores on the interior of the girdle band (white arrowheads). Scale bars = 51–56 = 2 µm; 57–58 = 1 µm.



Stratigraphic diatom succession

The base of the section (84 ka) is characterized by an assemblage dominated (58–90%) by *Aulacoseira granulata* (Ehrenb.) Simonsen and *A. ambigua* (Grunow) Simonsen (Fig. 59). *Cyclotella meneghiniana* and *Discostella stelligera* occurred punctually (<20%) between 82.3 and 80 ka. The *Aulacoseira* dominated assemblage persists up to 70 ka then greatly recedes (<20%) up to 1.5 ka. From around 60 ka, we observed successive occurrences of *D. stelligera*, *C. meneghiniana*, *Discostella gabinii* sp. nov. and *Cyclotella caspia* Grunow. At 45 ka, *Cyclotella petenensis* takes over the assemblage (73–97%) then declines abruptly at 31.5 ka. Prior to the collapse of *C. petenensis*, *Discostella gabinii* sp. nov. returned with fluctuating percentages for about 5 ka. Then, *Cyclocostis rolfii* gen et sp. nov. emerges at 26.9 ka, develops with fluctuating abundances with *Nitzschia amphibioides* Hust., *Mastogloia smithii* Thwaites, *M. elliptica* (C.Agardh) Cleve and *Navicula seminuloides* Hust. At 22.2 ka, *Cyclocostis rolfii* gen et sp. nov. disappears definitely while *Cyclotella petenensis* reoccurs. At first, *C. petenensis* coexists with *Discostella gabinii* sp. nov. (10–60%) then it takes over when *D. gabinii* sp. nov. declines. The dominant *C. petenensis* persists until 16.1 ka and does not reoccur thereafter in the sequence.

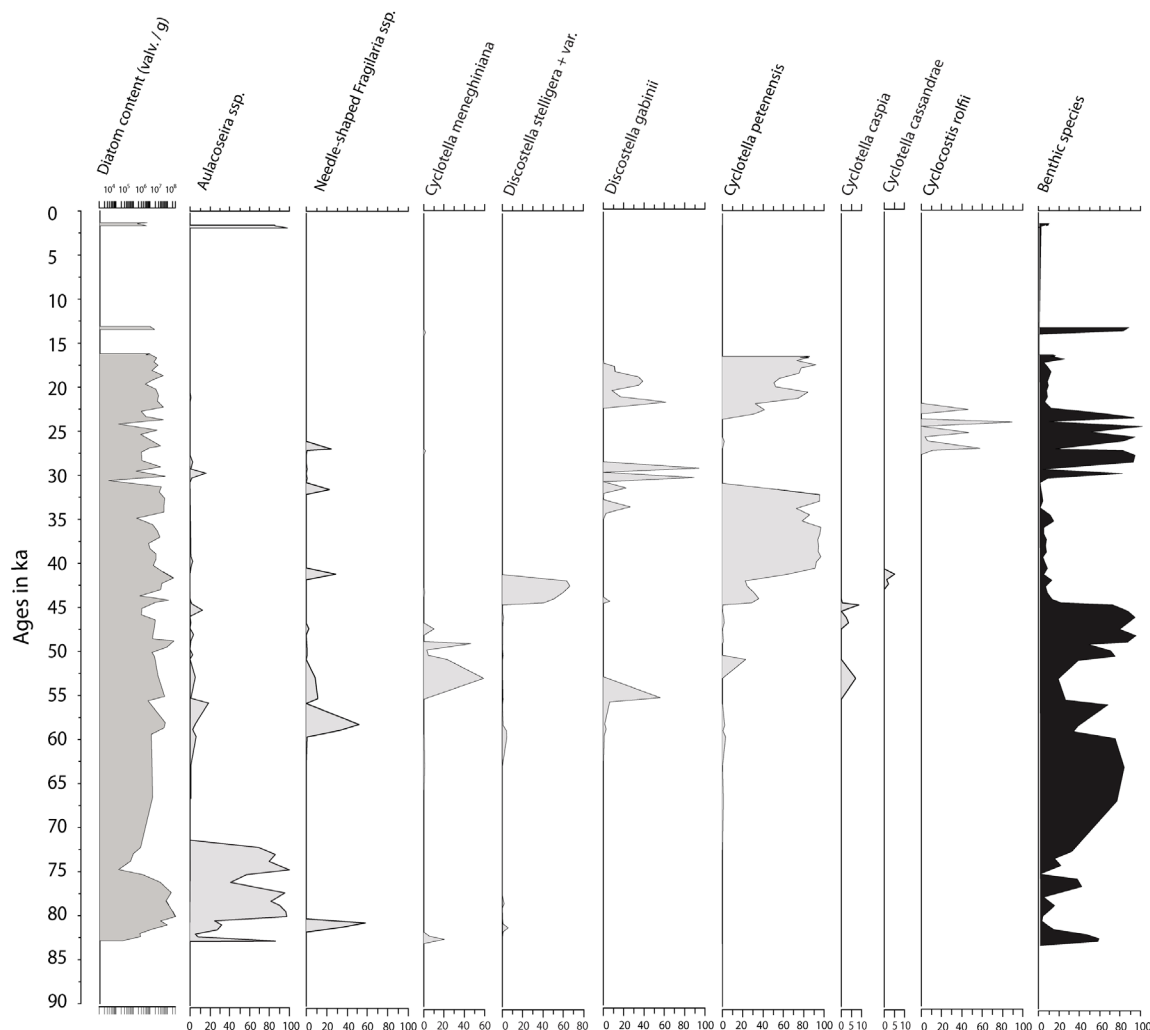


Fig. 59. Diagram showing the succession of Stephanodiscaceae Glezer & Makarova in Pleistocene sediments (0–84 ka) from Lake Petén-Itzá (Guatemala).

Ecology and associated diatom flora

In the modern dataset (Pérez *et al.* 2013), *Discostella gabinii* sp. nov. was initially identified as “*Cyclotella* sp22” (code CP22) and another species was identified as “*Discostella* aff. *pseudostelligera*” (CYAP). When analyzing the fossil flora and diagnosing *D. gabinii* sp. nov., we re-examined modern samples and observed that the two species were similar. “*Discostella* aff. *pseudostelligera*” and “*Cyclotella* sp22” were therefore combined together under the name *Discostella gabinii* sp. nov.

Conductivity, which is related to the precipitation gradient and marine influence on the Yucatan Peninsula, is the main variable that structures diatom, ostracod and cladoceran communities (Pérez *et al.* 2013). *Discostella gabinii* sp. nov. occurs in 11 water bodies of the Yucatan Peninsula (Table 1). In the Guatemalan highlands, its maximum occurrence (13%) was in Lake Amatitlan, a hypereutrophic alkaline lake spreading over 15.2 km² at 1200 m a.s.l. altitude. Water was calcium-bicarbonate rich, warm (22.6°C), with electrical conductivity of 630 µS/cm and high dissolved oxygen content (17.8 mg/L). Subdominant species were *Aulacoseira granulata*, *Cyclotella meneghiniana* and *Nitzschia pseudofonticola* Hust. In Cenotes Juarez, *D. gabinii* sp. nov. reached 4.8% in an assemblage dominated by *Achnanthisidium exiguum* (Grunow) Czarn. and *A. lineare* W.Sm. Water was warm (27.9°C) and alkaline with relatively high dissolved oxygen content (8.7 mg/L). Conductivity was 643 µS/cm. Water analyses determined Ca²⁺ (68.3 mg/L) and Mg²⁺ (23 mg/L) as the dominant cations and HCO₃⁻ (292.7 mg/L) as important anion. Overall, *D. gabinii* sp. nov. seems to tolerate varying conductivities but is most abundant in alkaline, low conductivities (600–650 µS/cm) and calcium-bicarbonated waters.

Cyclotella petenensis, although considered to be fossil at the time of description (Paillès *et al.* 2018), was identified as *C. meneghiniana* (CYMG) in the modern dataset mainly due to the fact that specimens were small in size, tangentially undulated with <5 valve face fulcra on the raised part. Once diagnosed as a new species in the sedimentary record, a re-examination of modern samples revealed that *C. petenensis* was present in low percentages (<4%) in five water bodies. Only in Lake Yalahau (Yucatan lowlands), *C. petenensis* reached 17.8% (Table 1). Of all water bodies investigated, Lake Yalahau had the highest diatom species richness. In this lake, water is shallow, warm (28.8°C) and alkaline (pH 8.9) with a high dissolved oxygen content (8.7 mg/L). Electrical conductivity is high 2350 µS/cm. Water was magnesium (136.8 mg/L) and bicarbonate (707.4 mg/L) rich. Its diatom population was composed of 33% of *C. meneghiniana* accompanied by *Brachysira australofollis* Lange-Bert. & Gerd Moser, *B. neoexilis* Lange-Bert., *Encyonema densistriata* Novelo, Tavera & Ibarra and *Fragilaria famelica* (Kütz.) Lange-Bert. In coastal Lake Progreso where *C. petenensis* represents 4% of the flora, water conductivity was 2040 µS/cm. In the modern samples, it appears that *C. petenensis* seems to favor waters with conductivities close to 2000 µS/cm.

Discussion

In terms of morphology, Stephanodiscaceae encompass a wide range of valve structures and thus genera. Interestingly, *Cyclocostis* gen. nov. possess distinctive morphological characters that can be found in species of genera *Cyclostephanos*, *Cyclotella* (*Lindavia*, *Discostella*, *Pantocsekiella*), *Stephanocostis*, *Pliocaenicus*, and *Paleotertiarius*. Conversely, the combination of these peculiar morphological features, which differentiate it from these genera, can be used to justify the definition of a new genus. In terms of valve structure, when looking at internal valve views of species of *Stephanodiscus* Ehrenb., *Stephanocostis* and most species of *Cyclostephanos*, the striking features are 1) the absence of alveoli and thus ribs, these being rather hyaline interfascicles and 2) the presence of domed criba covering internal areolae. Nevertheless, internal domed criba are always present in *Paleotertiarius* and *Pliocaenicus* and in some species of *Lindavia*. But *Paleotertiarius* and *Pliocaenicus* as well as some species of *Cyclostephanos* (*C. novaezeelandiae* and *C. dubius*) are also characterized by ghost or reduced alveoli. In contrast, internal alveolate structure – simple or complex – is a widespread structural component of

Table 1 (continued on the next page). Selected limnological and chemical data from lakes (single measurements in 2005 or 2008) containing *Discostella gabinii* Paillès & Sylvestre sp. nov. and *Cyclotella petenensis* Sylvestre, Paillès & Escobar in the Yucatan Peninsula after Pérez *et al.* (2013). n.d. = not determined.

Guatemalan highlands				Peten lowlands			
Lake name	Amatitlan	Atescatempa	Guija	Izabal	Salpeten	Bacalar-1	Bacalar-2
Lake ID	19	22	21	1	26	13	13
Latitude N	14°26'03.7"	14°13'01.1"	14°15'43.7"	15°29'24.5"	16°58'38.2"	18°39'54.0"	18°39'54.0"
Longitude W	90°32'58.6"	89°41'39.2"	89°32'11.3"	89°08'32.7"	89°40'30.9"	88°23'27.0"	88°23'27.0"
Species (%)							
<i>C. petenensis</i>	0.0	0.0	0.0	0.8	0.0	0.7	2.1
<i>D. gabinii</i> sp. nov.	13.2	0.3	5.0	0.0	2.0	0.2	0.2
Limnological parameters							
Max sampling depth (m)	0.5	0.5	0.5	14	38	65	16.3
Altitude (m a.s.l.)	1200	587	433	4	114	1	1
Surface area (km ²)	23	1.1	45	645	2.9	51	51
Temperature (°C)	22.8	27.3	26.2	26.4	25.4	28.8	27.1
DO ₂ (mg/L)	18.7	6.7	7.7	7.6	0.8	0.4	7.9
pH	9.3	8.0	8.4	8.3	7.3	6.9	7.8
Cond. (µS/cm)	630	283	206	216	4250	1400	1226
Secchi depth (m)	0.1–0.8	0.1	1.4	n.d.	0.8	10.3	10.3
Chemical variables (mg/L)							
HCO ₃ ⁻	234.8	182.9	121.9	119.6	140.3	305.6	187.3
SO ₄ ²⁻	n.d.	n.d.	n.d.	8.4	n.d.	1337.2	1300
Cl ⁻	n.d.	n.d.	n.d.	6.6	n.d.	81.3	85
Na ⁺	123.8	23.1	16.5	7.5	128.2	70.9	44.9
K ⁺	n.d.	n.d.	n.d.	1.9	n.d.	0.7	0.7
Ca ²⁺	23.2	23.3	26.9	25.9	795.8	460.3	399.6
Mg ²⁺	16.5	10.5	8.07	7.3	360.7	80.6	77.6
Yucatan lowlands					Cenotes	Coastal	Ponds
Lake name	Ocom	Coba	Yalahau-05	Yalahau-08	Juarez	Progreso	Belize 2
Lake ID	15	61	18	18	60	35	7
Latitude N	19°28'28.6"	20°29'40.2"	20°39'25.9"	20°39'25.9"	20°48'09.6"	18°13'05.2"	17°18'17.9"
Longitude W	88°03'17.9"	87°44'19.2"	89°13'02.0"	89°13'02.0"	87°20'23.8"	88°24'35.2"	88°29'18.9"
Species (%)							
<i>C. petenensis</i>	0.0	0.0	1.3	17.8	0.0	4.0	0.0
<i>D. gabinii</i> sp. nov.	3.7	0.6	1.3	0.0	4.8	0.0	0.1

Table 1 (continued). Selected limnological and chemical data from lakes (single measurements in 2005 or 2008) containing *Discostella gabinii* Paillès & Sylvestre sp. nov. and *Cyclotella petenensis* Sylvestre, Paillès & Escobar in the Yucatan Peninsula after Pérez *et al.* (2013).

Lake name	Yucatan lowlands				Cenotes	Coastal	Ponds
	Ocom	Coba	Yalahau-05	Yalahau-08	Juarez	Progreso	Belize 2
Limnological parameters							
Max sampling depth (m)	10	0.5	11.5	11.5	0.5	3.15	0.5
Altitude (m a.s.l.)	1	7	2	2	14	5	33
Surface area (km ²)	0.25	0.35	0.25	0.25	0.03	7.2	
Temp. (°C)	25	28.9	25	28.8	27.9	26.4	27.4
DO ₂ (mg/L)	0.8	8.7	0.7	8.7	8.7	7	7.5
pH	7.4	8.5	8.6	8.9	8.1	8.2	8.0
Cond. (µS/cm)	777	1213	1138	2350	643	2040	244
Secchi depth (m)	5.5	0.9	n.d.	1.1	1.6	1.3	n.d.
Chemical variables (mg/L)							
HCO ₃ ⁻	130.6	256.1	607.2	707.4	292.7	213.4	180.1
SO ₄ ²⁻	685.9	n.d.	337.1	n.d.	n.d.	n.d.	6.4
Cl ⁻	114.7	n.d.	145.4	n.d.	n.d.	n.d.	2.6
Na ⁺	125.2	126.5	224	336.5	53.3	483.6	3.66
K ⁺	3.5	n.d.	16.9	n.d.	n.d.	n.d.	1.6
Ca ²⁺	155.2	99.7	18.8	89.6	68.3	189.4	45.0
Mg ²⁺	44.6	5.4	101.4	136.8	23.0	59.6	2.28

Cyclotella s. str., *Lindavia*, *Pantocsekiella* and *Discostella*. As in *Paleotertiarius* and *Pliocaenicus*, the alveolus (Anonymous 1975; Houk *et al.* 2010) is a chamber opening to the inside of the cell by small or large opening with a perforate outer layer. Thus, the degree of occlusion of the alveolus is determined by centrifugal roofing. Consequently, contact between the cell and the exterior is highly reduced by the presence of internal central lamina (Servant-Vildary 1986). Similarly, in more simply structured valves of *Stephanodiscus*, *Cyclostephanos* and *Stephanocostis*, the presence of domed criba is assumed to reduce contact between the cell and the exterior.

In morphogenesis of centric diatoms, valves are systems of silica ribs, which grow out from a circular center during valve formation (Round *et al.* 1990). They are formed within silica deposition vesicle (SDV) enclosed in the silicalemma, starting with a small, thin disc composed of radial bunched siliceous strands from the center (Kaluzhnaya 2006). According to Bedoshvili & Likhoshway (2019), valve morphogenesis in centric diatoms begins with a formation of a ring (annulus) from which the ribs radiate centrifugally as the SDV grows and until the mantle is formed (=horizontal growth). Vertical growth occurs by silica deposition to thicken the valve and to differentiate fine structures such as areolae and various processes. Structurally, *Cyclcostis* gen. nov. appears simple: a web of radiating ribs starting from a central silicified ring covered by a coarsely perforated silica layer. The central ring represents a remainder of the annulus that was never filled in. Additionally, striae are reduced to two ribs covered with a perforated layer, and alveoli are widely open to the valve center. The frustule is strongly deformed and, consequently, has a larger surface area compared to a flat disc. Furthermore, the cell

cavity is reduced (see the thickness of the frustule in Fig. 33). It could be anticipated that such simple construction combined with a large surface area would greatly expose the cell to surrounding waters as well as facilitate communication with the outside environment.

The species of diatoms are discerned by phenotypic characters. According to Benton & Pearson (2001), the process of speciation is too slow to be observed directly. However, along an upper Miocene lacustrine sequence, populations of triangular *Cyclotella* sp. exhibit complexification of their alveoli suggesting a trend toward a reduced contact between the cell and its surrounding environment (Servant-Vildary 1986). Similarly, Theriot *et al.* (2006) observed a morphological shift in the *Stephanodiscus niagarae* C.Ehrenb./*S. yellowstonensis* E.C.Ther. & Stoermer complex between 13.7 and 10.0 ka. They suggest that directional morphological evolution strongly associated with continuous environmental change would account for the evolution of *S. yellowstonensis*. Additionally, in Pleistocene sedimentary sequences from lakes Ohrid and Prespa, morphological variations (valve diameter, striae morphology, number of ribs and valve face fultoportulae) in *Cyclotella* populations suggested the expression of environmental factors (Cvetkoska *et al.* 2014). Interestingly, these studies only concern centric diatoms, the only study on pennate diatoms in lacustrine sequences concluded that levels of morphological differentiation were likely a consequence of limited dispersal (Evans *et al.* 2009).

In the PI-6 sedimentary record, from around 60 to 16.1 ka, we observe a succession of *Cyclotella meneghiniana*, *Discostella stelligera*, *D. gabinii* sp. nov., *C. petenensis* and *Cyclocostis rolfii* gen et sp. nov. that could evoke fluctuations of the lacustrine environment. A similar interpretation was made by Bradbury (1971) in a >46 ka diatom record from Lake Texcoco in Mexico where *Cyclotella* cf. *stylorum* (probably *C. petenensis*), *C. quillensis* L.W.Bailey and *C. striata* (Kütz.) Grunow alternated and reflected changes in salinity. He noticed that their distribution throughout the core was an alternation of ecotypes. From a morphological point of view, *C. petenensis* and *C. cassandrae* are probably variations of *C. meneghiniana*, initially present in our record (Paillès *et al.* 2018). Similarly, *Discostella gabinii* sp. nov. could also be a variation of *D. stelligera* firstly present in the record. For the newly described *Cyclocostis* gen. nov., its basic structure and the strong deformation of the valve suggest enhanced communication between the cell and the exterior. During 5 ka, *Cyclocostis* gen. nov. alternates with benthic species such as *Nitzschia amphibioides*, *Mastogloia smithii*, *M. elliptica* and *Navicula seminuloides*. As such, its punctual presence could also be the expression of local and rapid changes in the environment.

Overall, the succession of the species of *Cyclotella*, *Discostella* and *Cyclocostis* gen. nov. in our record could represent eco-phenotypic responses to environmental change/stress. Changes in water conductivity and/or water level fluctuations could be evoked as suggested by the presence of benthic mesosaline to hypersaline species developing between *Cyclotella*, *Discostella* and *Cyclocostis* gen. nov. episodes. Except for *Cyclotella cassandrae* and *Cyclocostis rolfii* gen et sp. nov. that were not identified in the modern dataset, all other centric species are still extant. Thus, a calibration of the modern data set in order to perform pH and conductivity transfer functions in the fossil record will be considered in future studies.

Acknowledgements

Drilling of Lake Petén-Itzá was funded by grants from the US National Science Foundation (ATM-0502030), the International Continental Scientific Drilling Program, the Swiss Federal Institute of Technology, and the Swiss National Science Foundation. We are indebted to Anders Noren, Kristina Brady and Amy Myrbo of LacCore (National Lacustrine Core Facility), Department of Earth Sciences, University of Minnesota-Twin Cities for their expertise in core acquisition, curation and sampling. We are also very grateful towards D. Hodell, M. Brenner, J. Curtis, L. Perez, A. Schwalb, S. Kutterolf for providing material and data, R. Klee, V. Houk and P. Kociolek, for looking at specimens and encouraging

this publication and L. Ector for providing extensive literature. SEM investigations were funded by the Institute for Research and Development (France). The Laboratoire Préparation Micropaléontologique (CEREGE, France) provided all facilities for sample processing. Finally, the constructive comments of two anonymous reviewers is gratefully acknowledged.

References

- Àcs E., Ari E., Duleba M., Dressler M., Genkal S.I., Jako E., Rimet F., Ector L. & Kiss K. 2016. *Pantocseckiella*, a new centric diatom genus based on morphological and genetic studies. *Fottea* 16 (1): 56–78. <https://doi.org/10.5507/fot.2015.028>
- Alfinito S. & Tagliaventi N. 2002. Morphology and taxonomy of *Cyclotella elentarii* spec. nova a newly described centric diatom from a New Zealand lake. *Algological studies* 105: 29–38. https://doi.org/10.1127/algol_stud/105/2002/29
- Anonymous. 1975. Proposals for standardization of diatom terminology and diagnoses. *Beiheft zur Nova Hedwigia* 53: 323–354.
- Bedoshvili D. & Likhoshway Y. 2019. Cellular mechanisms of diatom valve morphogenesis. In: Seckbach J. & Gordon R. (eds) *Diatoms: Fundamentals and Applications*: 99–114. Scrivener Publishing, Austin. <https://doi.org/10.1002/9781119370741.ch5>
- Benton M.J. & Pearson P. 2001. Speciation in the fossil record. *Trends in Ecology & Evolution* 16 (7): 405–411. [https://doi.org/10.1016/S0169-5347\(01\)02149-8](https://doi.org/10.1016/S0169-5347(01)02149-8)
- Bradbury J.P. 1971. Paleolimnology of Lake Texcoco, Mexico. Evidence from diatoms. *Limnology and Oceanography* 16 (2): 180–200. <https://doi.org/10.4319/lo.1971.16.2.0180>
- Bradbury J.P. 2000. Limnology history of Lago Pátzcuaro, Michoacán, Mexico for the past 48,000 years: impacts of the climate and man. *Palaeogeography, Palaeoclimatology, Palaeoecology* 163: 69–95. [https://doi.org/10.1016/S0031-0182\(00\)00146-2](https://doi.org/10.1016/S0031-0182(00)00146-2)
- Brébisson A. de 1838. *Considérations sur les Diatomées et Essai d'une Classification des Genres et des Espèces appartenant à cette Famille*. Brée l'Ainé Imprimeur-Librairie, Falaise. <https://doi.org/10.5962/bhl.title.64353>
- Chávez L. & Haberyan K.A. 1996. Diatom assemblages from the Camastro diatomite, Costa Rica. *Revista de Biología tropical* 44 (2): 899–902.
- Cohuo S., Macario-González L., Pérez L., Sylvestre F., Paillès C., Curtis J.H., Kutterolf S., Wojewódka M., Zawisza E., Szeroczyńska K. & Schwalb A. 2018. Climate ultrastructure and aquatic community response to Heinrich Stadials (HS5a–HS1) in the continental northern Neotropics. *Quaternary Science Reviews* 197: 75–91. <https://doi.org/10.1016/j.quascirev.2018.07.015>
- Cvetkoska A., Hamilton P., Ognjanova-Rumenova N. & Levkov Z. 2014. Observations of the genus *Cyclotella* (Kützing) Brébisson in ancient lakes Ohrid and Prespa and a description of two new species *C. paraocellata* sp. nov. and *C. prespanensis* sp. nov. *Nova Hedwigia* 98 (3–4): 313–340. <https://doi.org/10.1127/0029-5035/2014/0154>
- Escobar J., Hodell D.A., Brenner M., Curtis J.H., Gilli A., Mueller A.D., Anselmetti F.S., Ariztegui D., Grzesik D.A., Perez L., Schwalb A. & Guilderson T.P. 2012. A ~43-ka record of paleoenvironmental change in the Central American lowlands inferred from stable isotopes of lacustrine ostracods. *Quaternary Science Reviews* 37: 92–104. <https://doi.org/10.1016/j.quascirev.2012.01.020>
- Evans K.M., Chepurinov V.A., Sluiman H.J., Thomas S.J., Spears B.M. & Mann D.G. 2009. Highly differentiated populations of the freshwater diatom *Sellaphora capitata* suggest limited dispersal and opportunities for allopatric speciation. *Protist* 160: 386–396. <https://doi.org/10.1016/j.protis.2009.02.001>

- Genkal S.L. 1977. On counting of some taxonomically significant structural elements of valves in the diatom algae of the family Thalassiosiraceae Lebour emend. Hasle (Bacillariophyta). *Botaničeskij žurnal* 62 (6): 848–851. [In Russian.]
- Glezer Z.I. & Makarova I.V. 1986. Nouveaux ordres et familles de Diatomées (Bacillariophyta). *Botaničeskij žurnal* 71 (5): 673–676.
- Guiry M.D. & Guiry G.M. 2020. *AlgaeBase*. World-wide electronic publication, National University of Ireland, Galway. Available from <http://www.algaebase.org> [accessed 12 Jul. 2020].
- Habeyran K.A. & Horn S.P. 1999. A 10,000 years diatom record from a glacial lake in Costa Rica. *Mountain Research and Development* 19 (1): 63–70. <https://doi.org/10.2307/3674114>
- Haberyan K.A. & Horn S.P. 2005. Diatom paleoecology of Laguna Zoncho, Costa Rica. *Journal of Paleolimnology* 33 (3): 361–369. <https://doi.org/10.1007/s10933-004-6789-4>
- Håkansson H. 1990. *Cyclotella meneghiniana* Kütz. (Bacillariophyceae), its morphology and reappraisal of similar species. *Beiheft zur Nova Hedwigia* 100: 19–37.
- Håkansson H. 2002. A compilation and evaluation of species in the general *Stephanodiscus*, *Cyclostephanos* and *Cyclotella* with a new genus in the family Stephanodiscaceae. *Diatom Research* 17 (1): 1–139. <https://doi.org/10.1080/0269249X.2002.9705534>
- Håkansson H., Hadju H., Snoeijs P. & Loginova L. 1993. *Cyclotella hakanssoniae* Wendker, and its relationship to *C. caspia* Grunow and other similar brackish water *Cyclotella* species. *Diatom Research* 8: 333–347. <https://doi.org/10.1080/0269249X.1993.9705266>
- Hodell D.A., Anselmetti F.S., Ariztegui D., Brenner M., Curtis J.H., Gilli A., Grzesik D.A., Guilderson T.J., Müller A.D., Bush M.B., Correa-Metrio A., Escobar J. & Kutterolf S. 2008. An 85-ka record of climate in lowland Central America. *Quaternary Science Reviews* 27: 1152–1165. <https://doi.org/10.1016/j.quascirev.2008.02.008>
- Houk V., Klee R. & Tanaka H. 2010. Atlas of freshwater centric diatoms with a brief key and descriptions. Part III. Stephanodiscaceae A, *Cyclotella*, *Tertiarius*, *Discostella*. *Fottea* 10 (supplement): 1–498.
- Houk V., Klee R. & Tanaka H. 2014. Atlas of freshwater centric diatoms with a brief key and descriptions. Part IV. Stephanodiscaceae B, *Stephanodiscus*, *Cyclostephanos*, *Pliocaenicus*, *Hemistephanos*, *Stephanocostis*, *Mesodictyon* & *Spicaticribra*. *Fottea* 14 (supplement): 1–532.
- Kaluzhnaya O. 2006. Valve morphogenesis in the centric diatom *Cyclotella baicalensis*. In: Likhosway Y. (ed.) *Nineteenth International Diatom Symposium*: 31–38. Biopress, Bristol.
- Khursevich G. & Kociolek J.P. 2012. A preliminary worldwide inventory of the extinct, freshwater fossil diatoms from the orders Thalassiosirales, Stephanodisciales, Paraliales, Aulacoseirales, Melosirales, Coscinodisciales and Biddulphiales. *Beiheft zur Nova Hedwigia* 141: 315–364.
- Klee R. & Houk V. 1996. Morphology and ultrastructure of *Cyclotella woltereckii* Hustedt (Bacillariophyceae). *Archiv für Protistenkunde* 147: 19–27. [https://doi.org/10.1016/S0003-9365\(96\)80005-4](https://doi.org/10.1016/S0003-9365(96)80005-4)
- Knapp J.M., Furey P.C. & Lowe R.L. 2006. A comparison of the morphology and ultrastructure of the diatoms (Bacillariophyceae) *Discostella stelligera* and *D. elentarii* from two lakes in Fjorland, New Zealand. *New Zealand Journal of Marine and Freshwater Research* 40: 429–438. <https://doi.org/10.1080/00288330.2006.9517433>
- Kociolek J.P. & Khursevich G.H. 2013. Morphology of some lacustrine centric species from the western United States assigned to the genus *Cyclotella* (Bacillariophyta), including four described as new. *Phytotaxa* 127 (1): 81–99. <https://doi.org/10.11646/phytotaxa.127.1.11>

- Kociolek J.P., You Q., Stepanek J.G., Lowe R.L. & Wang Q. 2016. New freshwater diatom genus, *Edtheriotia* gen. nov. of the Stephanodiscaceae (Bacillariophyta) from south-central China. *Phycological Research* 84: 274–280. <https://doi.org/10.1111/pre.12145>
- Kociolek J.P., Balasubramanian K., Blanco S., Coste M., Ector L., Liu Y., Kulikovskiy M., Lundholm N., Ludwig T., Potapova M., Rimet F., Sabbe K., Sala S., Sar E., Taylor J., Van de Vijver B., Wetzel C.E., Williams D.M., Witkowski A. & Witkowski J. 2018. DiatomBase. *Discostella* Houk & Klee, 2004. Available from <http://www.diatombase.org/aphia.php?p=taxdetails&id=465545> [Accessed 19 Mar. 2020].
- Krahn K., Wetzel C., Ector L. & Schwalb A. 2018. *Achnanthidium neotropicum* sp.nov., a new freshwater diatom from lake Apastepeque in El Salvador (Central America). *Phytotaxa* 382 (1): 89–101. <https://doi.org/10.11646/phytotaxa.382.1.4>
- Krammer K. & Lange-Bertalot H. 1986. Bacillariophyceae. 1. Teil: Naviculaceae. In: Ettl H., Gerloff J., Heynig H. & Mollenhauer D. (eds) *Süßwasserflora von Mitteleuropa, Band 2/1*: 1–876. Gustav Fischer Verlag, Stuttgart, Jena.
- Krammer K. & Lange-Bertalot H. 1988. Bacillariophyceae. 2. Teil: Bacillariaceae, Epithemiceae, Surirellaceae. In: Ettl H., Gerloff J., Heynig H. & Mollenhauer D. (eds) *Süßwasserflora von Mitteleuropa, Band 2/2*: 1–596. Gustav Fischer Verlag, Stuttgart, Jena.
- Krammer K. & Lange-Bertalot H. 1991a. Bacillariophyceae. 3. Teil: Centrales, Fragilariaceae, Eunotiaceae. In: Ettl H., Gerloff J., Heynig H. & Mollenhauer D. (eds) *Süßwasserflora von Mitteleuropa, Band 2/3*: 1–576. Gustav Fischer Verlag, Stuttgart, Jena.
- Krammer K. & Lange-Bertalot H. 1991b. Bacillariophyceae. 4. Teil: Achnanthaceae, Kritische Ergänzungen zu Navicula (Lineolatae) und Gomphonema, Gesamtliteraturverzeichnis Teil 1–4. In: Ettl H., Gerloff J., Heynig H. & Mollenhauer D. (eds) *Süßwasserflora von Mitteleuropa, Band 2/4*: 1–437. Gustav Fischer Verlag, Stuttgart, Jena.
- Kutterolf S., Schindlbeck J.C., Anselmetti F.S., Aritzegui D., Brenner M., Curtis J., Schmid D., Hodell D.A., Mueller A., Pérez L., Pérez W., Schwalb A., Frische M. & Wang K.-L. 2016. A 400-ka tephrochronological framework for Central America from Lake Petén Itzá (Guatemala) sediments. *Quaternary Science Reviews* 150: 200–220. <https://doi.org/10.1016/j.quascirev.2016.08.023>
- Lange C. & Syversten E.E. 1989. *Cyclotella litoralis* sp. nov. (Bacillariophyceae) and its relationship to *C. striata* and *C. stylorum*. *Nova Hedwigia* 48 (3–4): 341–356.
- Lange-Bertalot H. & Metzeltin D. 2009. A dystrophic mountain lake in Panama - Hot spot of new and rare neotropical diatoms. *Nova Hedwigia* 135: 137–165.
- Loginova L.P. 1990. Classification of the diatom genus *Cyclotella*. In: Simola H. (ed.) *Proceedings of the 10th international diatom symposium*: 37–53. Koeltz Scientific Books, Koenigstein.
- Lowe R.L. 1975. Comparative ultrastructure of the valves of some *Cyclotella* species (Bacillariophyceae). *Journal of Phycology* 11 (4): 415–424. <https://doi.org/10.1111/j.1529-8817.1975.tb02805.x>
- McFarland B.H. & Collins G.B. 1978. A key to the species of the diatom genus *Cyclotella* (Kütz.) Bréb. based on new morphological data. Abstract. In: *26th annual meeting of North American Benthological Society*: 35. Winnipeg.
- Metcalf S.E., O'Hara S.L., Caballero M. & Davies S.J. 2000. Records of Late Pleistocene – Holocene climatic change in Mexico: a review. *Quaternary Science Reviews* 19: 699–721. [https://doi.org/10.1016/S0277-3791\(99\)00022-0](https://doi.org/10.1016/S0277-3791(99)00022-0)
- Nakov T., Guillory W.X., Julius M.L., Theriot E.C. & Alverson A.J. 2015. Towards a phylogenetic classification of species belonging to the diatom genus *Cyclotella* (Bacillariophyceae): transfer of

- species formerly placed in *Puncticulata*, *Handmannia*, *Pliocaenicus* and *Cyclotella* to the genus *Lindavia*. *Phytotaxa* 217 (3): 249–264. <https://doi.org/10.11646/phytotaxa.217.3.2>
- Ortega B., Vázquez G., Caballero M., Israde I., Lozano-Garcia S., Schaaf P. & Torres E. 2010. Late Pleistocene: Holocene record of environmental changes in lake Zirahuen, Central Mexico. *Journal of Paleolimnology* 44: 745–760. <https://doi.org/10.1007/s10933-010-9449-x>
- Paillès C., Sylvestre F., Escobar J., Tonetto A., Rustig S. & Mazur J.C. 2018. *Cyclotella petenensis* and *Cyclotella cassandrae*, two new fossil diatoms from Pleistocene sediments of Lake Petén-Itzá, Guatemala, Central America. *Phytotaxa* 351 (4): 247–263. <https://doi.org/10.11646/phytotaxa.351.4.1>
- Pérez L., Lorenschat J., Massafiero J., Paillès C., Sylvestre F., Hollwedel W., Brandorff G.-O., Brenner M., Islebe G., del Socorro Lozano M., Scharf B. & Schwalb A. 2013. Bioindicators of climate and trophic status in aquatic ecosystems of the northern Neotropics. *Revista de Biología Tropical* 61 (2): 603–644. <https://doi.org/10.15517/rbt.v61i2.11164>
- Prasad A.K.S.K. & Nienow A. 2006. The centric diatom genus *Cyclotella* (Stephanodiscaceae: Bacillariophyta) from Florida Bay, USA, with special reference to *Cyclotella choctawhatcheeana* and *Cyclotella desikacharyi*, a new marine species related to the *Cyclotella striata* complex. *Phycologica* 45 (2): 127–140.
- Rosenmeier M.F., Brenner M., Kenney W.F., Whitmore T.J. & Taylor C.M. 2004. Recent eutrophication in the southern basin of Lake Petén Itzá, Guatemala: human impact on a large tropical lake. *Hydrobiologia* 511 (1–3): 161–172. <https://doi.org/10.1023/B:HYDR.0000014038.64403.4d>
- Ross R., Cox E.J., Karayeva N.I., Mann D.G., Paddock T.B.B., Simonsen R. & Sims P.A. 1979. An amended terminology for the siliceous components of the diatom cell. *Beiheft zur Nova Hedwigia* 64: 513–533.
- Round F.E., Crawford R.M. & Mann D.G. 1990. *The Diatoms. Biology and Morphology of the Genera*. Cambridge University Press, Cambridge.
- Serieyssol K. 1981. *Cyclotella* species of Late Miocene from St. Bazile, France. In: Ross R. (ed.) *Proceedings of the 6th International Diatom Symposium*: 27–42. Koeltz Scientific Books, Koenigstein.
- Servant-Vildary S. 1986. Fossil *Cyclotella* species from Miocene deposits of Spain. In: Ricard M. (ed.) *Proceedings of the 8th International Diatom Symposium*: 495–511. Otto Koeltz, Koenigstein.
- Slate J.E., Jonson T.C. & Moore T.C. 2013. Impact of pre-Columbian agriculture, climate change, and tectonic activity inferred from a 5,700-years paleolimnological record from lake Nicaragua. *Journal of Paleolimnology* 50: 139–149. <https://doi.org/10.1007/s10933-013-9709-7>
- Tagliaventi N. & Cavacini P. 2002. A study on the morphology of *Cyclotella areolata* Hustedt, *C. stelligera* Cleve et Grunow var. *robusta* and *C. stelligera* Cleve et Grunow var. *hyalina* Hustedt, using original material. *Algological studies* 107: 1–16. https://doi.org/10.1127/algol_stud/107/2002/1
- Tanaka H. 2007. Taxonomic studies of the genera *Cyclotella* (Kützinger) Brébisson, *Discostella* Houk et Klee and *Puncticulata* Håkansson in the family Stephanodiscaceae Glezer et Makarova (Bacillariophyta) in Japan. *Bibliotheca Diatomologica* 53: 1–204.
- Temoltzin-Loranca Y., Velez M.I., Moreno E. & Escobar J. 2018. Late Holocene environmental change in Lake Boquete and its watershed: human or natural causes? *Boletín de la Sociedad Geológica Mexicana* 70 (1): 121–131. <https://doi.org/10.18268/BSGM2018v70n1a7>
- Theriot E.C., Håkansson H., Kocielek J.P., Round F.E. & Stoermer E.F. 1987. Validation of the centric diatom genus name *Cyclotella*. *British Phycological Journal* 22: 345–347. <https://doi.org/10.1080/00071618700650411>

Theriot E. & Serieyssol K. 1994. Phylogenic systematics as a guide to understanding features and potential morphological characters of the centric diatoms family Thalassiosiraceae. *Diatom Research* 9 (2): 429–450. <https://doi.org/10.1080/0269249X.1994.9705318>

Theriot E.C., Fritz S.C, Whitlock C. & Conley D. 2006. Late Quaternary rapid morphological evolution of an endemic diatom in Yellowstone Lake, Wyoming. *Paleobiology* 32 (1): 38–54. [https://doi.org/10.1666/0094-8373\(2006\)032\[0038:LQRMEQ\]2.0.CO;2](https://doi.org/10.1666/0094-8373(2006)032[0038:LQRMEQ]2.0.CO;2)

Wetzel C.E. & Ector L. 2014. *Planothidium lagerheimii* comb. nov. (Bacillariophyta, Achnanthes) a forgotten diatom from South America. *Phytotaxa* 188 (5): 261–267. <https://doi.org/10.11646/phytotaxa.188.5.3>

Whitmore T.J., Brenner M., Curtis J.H., Dahlin B.H. & Leyden B.W. 1996. Holocene climate and human influences on lakes of the Yucatan Peninsula Mexico: an interdisciplinary, palaeolimnological approach. *The Holocene* 6: 273–287. <https://doi.org/10.1177/095968369600600303>

Manuscript received: 6 April 2020

Manuscript accepted: 24 September 2020

Published on: 30 November 2020

Topic editor: Christian de Muizon

Desk editor: Radka Rosenbaumová

Printed versions of all papers are also deposited in the libraries of the institutes that are members of the *EJT* consortium: Muséum national d'histoire naturelle, Paris, France; Meise Botanic Garden, Belgium; Royal Museum for Central Africa, Tervuren, Belgium; Royal Belgian Institute of Natural Sciences, Brussels, Belgium; Natural History Museum of Denmark, Copenhagen, Denmark; Naturalis Biodiversity Center, Leiden, the Netherlands; Museo Nacional de Ciencias Naturales-CSIC, Madrid, Spain; Real Jardín Botánico de Madrid CSIC, Spain; Zoological Research Museum Alexander Koenig, Bonn, Germany; National Museum, Prague, Czech Republic.