The epidemiological importance of the animal reservoir of Trypanosoma brucei gambiense in the Congo *

2-Characterization of the Trypanosoma brucei complex

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Summary

Biological and biochemical characterization of 36 human and 5 animal congolese stocks of *Trypanosoma brucei* were performed. One human and all the animal stocks showed a quick adaptation to rodent host whereas the other 35 human stocks were characterized by a low virulence degree (Group 1 of *T. gambiense*). The virulent stocks showed hybridization patterns specific to the *gambiense* subspecies. Our results confirm the absence of the *T. b. brucei* subspecies in the Congo and the low prevalence of domestic animals infected with *T. b. gambiense* (0.5%). Two cycles of human trypanosomiasis may thus occur in Central Africa: a predominant man-to-man cycle with group 1 trypanosomes and a minor cycle involving an animal reservoir.

Introduction

Studies carried out in the Congo on endemic animal trypanosomiasis within the sleeping sickness foci have revealed a high prevalence of *Nannomonas* infections (16.9%) whereas 0.5% of the domestic animals examined were infected with *Trypanosoma brucei* (Noireau et al., 1986). The results confirm previous observations of Van Hoof (1947) and Kageruka et al. (1977). These observations from Central Africa are different from those in West Africa where the prevalence rate of animal. *T. brucei* infections is high. The characterization of stocks from Ivory Coast and Liberia shows the possible presence of an animal reservoir of *T. b. gambiense*, indicating that the epidemiological cycles of human trypanosomiasis are probably different in West from those in Central Africa (Mehlitz et al., 1982; Zillman et al., 1984).

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Trop. Med. Parasit. 40 (1989) 9–11 © Georg Thieme Verlag Stuttgart · New York In this article, new data on the *T. brucei* complex are analysed from the characterization of *T. brucei* stocks isolated from human and animal hosts in the Congo, and a model applicable to the epidemiology of sleeping sickness in Central Africa is proposed.

Materials and Methods

Trypanosoma brucei stocks

36 stocks were isolated from patients with sleeping sickness between 1984 and 1987. Trypanosomiasis patients were originated from various foci in the country.

Among the seven *T. brucei* stocks isolated from domestic animals (Noireau et al., 1986), five could be analysed for biological and biochemical properties: MOVS/CG/85/ORBZV 503, MOVS/CG/85/ORBZV 504, MSUS/CG/85/ORBZV 505, MOVS/CG/85/ORBZV 506 and MSUS/CG/85/ORBZV 507. The origins of the 41 *T. brucei* stocks are shown in Table 1.

Study of growth rate of T. brucei stocks

The rodent model (Wistar white rat) was used in preference to *Mastomys natalensis* which was found to be less sensitive in our comparative trials. Recent report showed that the rate of proliferation among african trypanosomes was directly related to their virulence in rodent host (Diffley et al., 1987). A stock was considered to be virulent if the following criteria were fulfilled before the third passage:

- incubation time under five days
- exponential increase in the parasitaemia curve, reaching antiLog 8.1 or more before the 8th day, according to the method of Herbert and Lumsden (1976)
- survival of the infected rat under 10 days

DNA hybridization

This biochemical characterization technique was applied to the five animal stocks and one human stock (MHOM/CG/85/ORBZV 113). This last one was clearly differentiated from the other 35 *T. b. gambiense* stocks by its virulence to the rodent.

Preparation of DNA: the lysates of the trypanosome clones were incubated for an hour at 37 °C with 100 µg/ml of RNAase. 100 µg/ml of proteinase K were then added and digestion allowed to proceed for four hours at the same temperature. The lysates were then extracted by an equal volume of phenol saturated with Tris-Hcl 50 mM (pH 8.00), then by an equal quantity of (1:1) mixture of phenol saturated with Tris-Hcl 50 mM (pH 8.00) and chloroform.

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Table 1 Origin of Trypanosoma brucei stocks

| Geographic | Human | Animal host: | |
|------------|-------|--------------|-----|
| origin | host | Sheep | Pig |
| Bouenza | 18 | 3 | 1 |
| Couloir | 8 | 0 | 0 |
| Sangha | 10 | 0 | 0 |
| Pool | 0 | 0 | 1 |
| Total | 36 | 3 | 2 |

Table 2 Biological characterization of Trypanosoma brucei stocks

| Origin of | Number of | Virulence in the rodent: | |
|-----------|-----------|--------------------------|------------|
| stocks | isolates | High | Low |
| Man | 36 | 1 (2.8%) | 35 (97.2%) |
| Animal | 5 | 5 (100%) | 0 (0%) |

After addition of sodium acetate (final concentration: 250 mM) to the aqueous phase, the DNA was precipitated by two volumes of ethanol at -20 °C and finally dissolved in Tris-Hcl 10 mM, EDTA 0.1 mM (pH

Analysis of DNA: this was carried out by the previously described method (Paindavoine et al., 1986).

Results

Virulence of T. brucei stocks

The results are shown in Table 2. It seems that except for the stock MHOM/CG/85/ORBZV 113 all the human isolates showed a low degree of virulence in the rodent which is an usual characteristic of T. b. gambiense stocks in Central Africa. By contrast the five animal stocks were characterized by a very high degree of virulence.

DNA hybridization

The six stocks analysed by three different DNA probes corresponding to the AnTat 1.1, AnTat 1.8 and AnTat 1.13 surface antigens of T. brucei presented hybridization patterns specific to the gambiense sub-species. They may belong to a group of T. b. gambiense found in Central Africa, principally in Cameroon and Congo.

Discussion

Our results confirm the absence of the T. brucei brucei subspecies in the Congo. The excellent specificity demonstrated by the indirect immunofluorescent antibody test applied to massscreening in this country may be the consequence of this (Frezil et al., 1977; Noireau et al., 1987). All Trypanozoon isolated from sheep and pig are therefore likely to be infective to man although this cannot be formally proven. These epidemiological data are different from those obtained in West Africa where stocks which are pathogenic and nonpathogenic to man are found in domestic animals (Mehlitz et al., 1982). The differences in the prevalence of animals infected with T. brucei in West and Central Africa suggest that a T. b. gambiense animal reservoir is possible, even probable in West Africa, whereas man seems to be the only effective reservoir in the Congo.

We propose that there are probably two groups of trypanosomes infective to man in the Congo. The first group corresponds most certainly to the group 1 of T. b. gambiense, as defined recently by Gibson (1986). The low virulence of the trypanosomes in group 1, which is one of their determining characteristics according to this author, enables most of the Congolese stocks isolated from a human host to be classified in that group. The stock MHOM/CG/85/ORBZV 113 is dis-

tinctly differentiated from this group by its high virulence to the rodent. However, the DNA hybridization experiment shows that this stock exhibit VSG patterns specific of T. b. gambiense which excludes this stock from the group 2 described by Gibson (1986). In addition, using mechanical transmission procedure, MHOM/CG/85/ORBZV 113 was able to infect pigs while three congolese T. gambiense stocks of group 1 did not one (Noireau, unpublished data). The similar characteristics of the five animal stocks of T. brucei and MHOM/CG/85/ORBZV 113 suggest that they are closely related. These stocks may be found both in man and domestic animals, unlike those in group 1 which do not seem to have an animal reservoir. Two cycles may thus occur in Central Africa, each characterized by a different group of T. b. gambiense: a man-to-man cycle (type 1) predominant in the Congo (T. b. gambiense of group 1), and a cycle involving an animal reservoir (type 2), epidemiologically less important and limited to a few foci. The reasons for the low prevalence of T. b. gambiense stocks characteristic of the second type of cycle are not known. It is probable that their hosts, whether animal or man, constitute an effective reservoir. Indeed, we have noticed that the animals infected with T. brucei seem to tolerate their infection. Similarly, the patient from whom the virulent T. b. gambiense stock was isolated, presented at diagnosis a clinical picture of chronic disease without the acute manifestations such as those observed in East Africa.

References

Difley, P., J. O. Scott, K. Mama, T. N. R. Tsen: The rate of proliferation among african trypanosomes is a stable trait that is directly related to virulence. Am. J. Trop. Med. Hyg. 36 (1987) 533-540

Frezil, J. L., J. Coulm, J. Alary: L'immunofluorescence indirecte et la stratégie de lutte contre la trypanosomiase humaine en Afrique Centrale. Med. Trop. 37 (1977) 285-289

Gibson, W. C.: Will the real Trypanosoma b.gambiense please stand up. Parasit. Today. 2 (1986) 255-257

Herbert, W. J., W. H. R. Lumsden: Trypanosoma brucei: A rapid "matching" method for estimating the host's parasitemia. Exp. Parasit. 40 (1976) 427-431

Kageruka, P. J., J. Colaert, Ngimbi Nkuku-Pela: Strain of Trypanosoma (Trypanozoon) brucei isolated from pigs in Bas-Zaire. Ann. Soc. belge Med. Trop. 57 (1977) 85-88

Mehlitz, D., U. Zillmann, C. M. Scott, D. G. Godfrey: Epidemiological studies on the animal reservoir of gambiense sleeping sickness. Part 3-Characterization of Trypanozoon stocks by isoenzymes and sensitivity to human serum. Tropenmed. Parasit. 33 (1982) 113-

Noireau, F., J. P. Gouteux, A. Toudic, F. Samba, J. L. Frezil: Importance épidémiologique du réservoir animal a Trypanosoma brucei gambiense au Congo. 1. Prévalence des trypanosomoses animales dans les foyers de maladie du sommeil. Tropenmed. Parasit. 37 (1986) 393-398

Noireau, F., J. L. Lemesre, M. Y. Nzoukoudi, M. T. Louembet, J. P. Gouteux, J. L. Frezil: Serodiagnosis of sleeping sickness in the Republic of the Congo: comparison if Indirect Immunofluorescent Antibody Test and Card Agglutination Test. Trans. Roy. Soc. Trop. Med. Hyg. 82 (1988) 237–240

Trop. Med. Hyg. 82 (1988) 237-240

Paindavoine, P., E. Pays, M. Laurent, Y. Geltmeyer, D. Le Ray, D. Mehlitz, M. Steinert: The use of DNA hybridization and numerical taxonomy in determining relationship between Trypanosoma brucei stocks and subspecies. Parasit. 92 (1986) 31-50

Van Hoof, L. M. J. J.: Observations on trypanosomiasis in Belgian Congo. 2-Variations in virulence and pathogenicity for man and animals. Trans. Roy. Soc. Trop. Med. Hyg. 40 (1947) 743-751

Zillmann, U., D. Mehlitz, R. Sachs: Identify of Trypanozoonstocks isolated from man and domestic dog in Liberia. Tropenmed. Parasit. 35 (1984) 105-108

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