BIODIVERSITY AND SUSTAINABILITY OF WETLAND RICE PRODUCTION: ROLE AND POTENTIAL OF MICROORGANISMS AND INVERTEBRATES

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Summary

This review considers some of the ecological foundations of sustainable wetland rice production related to microorganisms and invertebrates and their biodiversity. It summarizes (1) the aspects of sustainability of rice-producing environments involving microbial and invertebrate populations --maintenance of soil fertility, effects and control of rice pests and vector-borne diseases--, (2) how factors of crop intensification affect these populations and their biodiversity, (3) the agricultural practices that use microbial and invertebrate populations and their biodiversity, and (4) the current status of germplasm collections and the potential of biotechnology to use them to improve the sustainability of rice-producing environments.

The beneficial and detrimental roles of microorganisms and invertebrates in sustainable rice production have been identified and, sometimes, quantified. However, less is known about the possible long-term effects of crop intensification on these populations and their biodiversity.

Numerous methods using microorganisms and invertebrates to increase soil fertility and control pests and diseases have been tested. But the success of these methods is limited and their adoption almost negligible. This situation will probably remain unchanged as long as the methods designed are based on a very restricted knowledge on biodiversity, community structure, and trophic relationships at the ecosystem level, as it is currently.

Recent data on arthropods confirm that high biodiversity does not imply stability and low pest populations. Increases in pest and vector densities depend more on predator diversity, species resilience to perturbations, and biological attributes. Thus, increasing or preserving diversity per se does not necessarily contribute to pest stability, but developing effective trophic linkages might. This approach might also be valid for maintaining soil fertility through microbial management, the optimization of primary production in floodwater, and the optimization of nutrient recycling by invertebrate populations.

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1. INTRODUCTION

More than half of the world's population depends on rice which, in 1988, occupied 145 million hectares of land, with a global production of 468 million tonnes. An additional 300 million tonnes of rice will be needed in 2020 to meet the need of a fast-growing human population. This requires a 65% production increase within 30 years without much expansion of actual cultivated area (IRRI 1989). However, increased rice production should not be at the expense of future generations and should fulfill the concept of sustainability. It should be achieved through rice production management that (1) satisfies changing human needs and maintains production over time in the face of ecological difficulties and social and economic pressure, (2) maintains or enhances the quality of the environment, and (3) conserves or enhances natural resources. Aside from maintaining growth in productive agricultural systems and promoting growth in less productive systems, the major issues are (1) managing pests and nutrients in ways that reduce agrochemical use, (2) preserving the natural resource base, and (3) protecting the genetic base for agriculture.

This review considers the aspects of sustainability of rice production involving microbial and invertebrate populations, the effects of crop intensification on these populations, the agricultural practices that utilize microorganisms and invertebrates, and the current status of germplasm collections and their biotechnological use for improving the sustainability of rice-producing environments.

2. IMPORTANCE OF MICROORGANISMS AND INVERTEBRATES IN THE SUSTAINABILITY OF RICE-PRODUCING ENVIRONMENTS

In the ricefield agroecosystems, the major aspects of sustainability that involve microorganisms and invertebrates are the soil fertility maintenance and the detrimental effects and control of rice pests and vector-borne diseases.

2.1. Maintenance of soil fertility by microorganisms and invertebrates

From the point of view of yield sustainability, traditional wetland rice cultivation has been extremely successful. A moderate but stable yield has been maintained for thousands of years without deterioration of the environment (Bray, 1986). This is because flooding favors soil fertility and rice production by (1) bringing soil pH near to neutral; (2) increasing the availability of nutrients, especially P and Fe; (3) depressing soil organic matter decomposition and thus, maintaining soil N fertility; (4) favoring N₂ fixation; (5) depressing outbreaks of soil-borne diseases; (6) supplying nutrients from irrigation water; (7) depressing weed growth,

especially those of C₄ type, and (8) preventing water percolation and soil erosion (Watanabe et al., 1988).

There is however no assurance that, on the long term, crop intensification will not affect wetland soil fertility. Research on rice nutrition has shown that, at usual levels of inorganic fertilizer applied to ricefields, most N absorbed by the plant originates from the soil, where it is released by the turnover of a microbial biomass representing only a small percentage of total soil N (Watanabe et al., 1988). Crop residues, rhizosphere exudates, and photosynthetic aquatic biomass (algae and aquatic plants) contribute nutrients that allow microbial biomass replenishment. Crop residues are incorporated at the beginning of the cropping season while nutrients accumulating in the photosynthetic aquatic biomass (including biologically fixed N₂) are continuously recycled and reincorporated into the soil by zooplankton and soil fauna, which are therefore key components of ricefield fertility (Roger and Kurihara 1988). Some of the inputs allowing the replenishment of microbial biomass have been quantified, but a comprehensive understanding of the mechanisms involved in this aspect of N cycling is still to be developed. It is important to understand and predict how factors associated with crop intensification, (e.g., agrochemicals) may affect the soil microbial biomass, directly or indirectly, by decreasing the productivity of the photosynthetic aquatic biomass and the populations of invertebrates responsible for recycling soil nutrients.

2.2. Yield losses caused by microbial and invertebrate pests of rice Of the approximately 100 insect species and 74 diseases and physiological disorders associated with rice (Teng, 1990), 30 insects and 16 diseases are considered economically important (Riessig et al, 1986). Table 1 presents a summary of estimates of yield loss due to pests and diseases.

The important insect pests of wetland rice are brown planthopper (BPH), leaffolder, stem borer, and green leafhopper (a vector of tungro virus). Important but localized losses have been attributed to rice bug (*Leptocorisa* spp.), gall midge larvae (*Pachydiplosis oryzae*), rice hispa (*Dicladispa armigera*), and armyworm (*Mythimna separata*) (Teng, 1986, 1990). Scarce information exists on losses caused by nematodes. Important diseases are tungro virus, sheath blight, bacterial blight, and blast (on susceptible varieties). In recent years, tungro has become a major problem in many tropical areas because of its potential to cause total loss and the lack of corrective measures once its symptoms are observed. Few data are available on yield losses caused by other diseases; but generally, under favorable conditions, most pathogens have the potential to cause severe losses (Teng et al., 1990).

Generalized crop loss figures most commonly cited are those by Cramer (1967) who concluded that more of the rice potential production is lost due to pests (55%) than is harvested (

45%). He estimated the percentage of the potential harvest lost due to pests was 34% for insects, 10% for diseases, and 11% for weeds. Although these figures appear to be high, Teng (1990) found that other authors have felt that there is no sound evidence to the contrary and that these values may be under-estimations of actual losses in some years. Other generalized estimates for losses caused by insects of tropical rice are 35% - 44% (Pathak & Dhaliwal, 1981), 24% in East and Southeast Asia (Ahrens et al. ,1982), 35% in India, and 16-30% in the Philippines (Way, 1976).

2.3. Vector-borne diseases

Wetland rice culture and irrigation schemes in tropical and subtropical regions create ecological conditions favorable for the propagation of vector-borne diseases. The most important of these diseases are malaria, schistosomiasis, and Japanese encephalitis, whose vectors require an aquatic environment. Invertebrate vectors of human diseases in rice-growing environments are basically mosquitoes and aquatic snails.

The reproduction of mosquitoes in ricefields is affected by plant height, water depth, soil and other environmental conditions, and cultural practices. Generally, larval populations are low after transplanting, peak a few weeks later, and decline as the plants reach a height of 60-100 cm. Mosquito reproduction in ricefields ranges from 2 to 20 m⁻² day⁻¹ (Roger and Bhuiyan, 1990).

Aquatic snails are very common in ricefields where they can develop large populations, especially at the beginning of the cropping season when organic manure is applied. Populations up to 1000 m⁻² have been observed in Philippine ricefields. Behavioral experiments showed that snails having to choose between various soils were most often attracted (75%) to rice soils than to other soil types (Roger and Kurihara 1988).

3. EFFECTS OF CROP INTENSIFICATION ON MICROBIAL AND INVERTEBRATE POPULATIONS

3.1. General effects of crop intensification on biodiversity in ricefields

Traditional ricefields, some of which have been cultivated for several hundred years, may be considered as climax communities. Modern technologies -- which utilize fertilizer-responsive varieties, fertilizers, pesticides, and optimum water and crop management practices -- have tremendously increased yields and production but have, indeed, caused profound modifications to traditional rice-growing environments.

In general, a disturbance to a stabilized ecosystem reduces the number of species while provoking "blooms" of certain others. Those effects have been observed in ricefields (Roger and Kurihara, 1988). However, the quantitative knowledge on the long-term effects of crop

47

intensification on species diversity is extremely scarce. The only reference on the species abundance in traditional ricefields is a study conducted in 1975 by Heckman (1979) in Thailand where 590 species were recorded in one field within one year (Table 2). Few records of aquatic invertebrates can be compared with Heckman's record of 183 species (Table 2). In a 2-year study of pesticide applications on Malaysian ricefields, Lim (1980) recorded 39 taxa of aquatic invertebrates. Single sampling by Takahashi et al. (1982) in four Californian ricefields recorded 10-21 taxa. In 18 sites in the Philippines and India, the highest number of aquatic invertebrate taxa recorded by single sampling at one site was 26; the lowest, 2 (Roger et al., 1987). Similarly, records of numbers of arthropod species in Japanese ricefields estimated in 1954-55 by net sweeping (Kobayashi et al., 1973) seem to indicate a higher biodiversity than in recent data collected by Heong et al. (unpubl. data) in 5 fields in the Philippines using the suction method (Table 2). All the above data were obtained using different sampling methods and time frame. The marked decrease of values recorded since 1975 might probably be taken as a rough indication of a decrease in total number of species after crop intensification; however, this does not demonstrate the generally accepted concept that crop intensification decreases biodiversity in ricefields.

Crop intensification has reduced the number of edible species traditionally harvested from ricefields. Heckman (1979) reported that 1 vegetable and 16 edible animal species (snail, prawn, crab, large water bug, fish, frog) were collected in a single ricefield within 1 year. Such a diversity is not common anymore, and pesticides may have rendered these edible species unfit for human consumption.

Agrochemical use, besides increasing rice yield, may also cause uncontrolled growth of single species that might, directly or indirectly, have detrimental effects. One of those effects is the outbreak of pests (Heinrichs, 1988) and other organisms that may affect the fertility- or health-related aspects of the ecosystem; for example, (1) blooms of unicellular algae, observed after fertilizer application, which cause N losses by volatilization, (2) proliferation of ostracods and chironomid larvae, observed after insecticide application, which inhibits the development of efficient N₂-fixing blue-green algae (BGA) blooms; and (3) proliferation of snails or mosquito larvae that may occur after insecticide application and favor vector-borne diseases (Roger and Kurihara 1988).

3.2. Effects on soil and water microbial populations

Most of the information on the impacts of crop intensification on ricefield microflora deals with pesticide use that may (1) alter activities related to soil fertility and (2) reduce pesticide efficiency because of shifts in microbial populations toward organisms more efficient in their degradation. More than 200 papers -- reviewed by Roger (1990) -- have been published on this topic, but more than half of the studies are short-term laboratory experiments in test tubes or

flasks that cannot be extrapolated to field conditions. Field and long-term laboratory studies on soil with pesticide levels near the recommended field dose allow to draw the following conclusions.

Pesticides have three major effects on ricefield algae: (1) a selective toxicity that affects preferentially green algae and thus promote BGA growth; (2) a short-term promoting effect of insecticides on microalgae caused by a temporary decrease of invertebrate that graze on algae; (3) a selective effect of insecticides on BGA flora by causing a recruitment of algal grazers which results in the dominance of strains forming mucilaginous macrocolonies (such as *Nostoc* spp.) resistant to grazing.

Field and laboratory studies showed that pesticides applied on soil at the recommended rates and intervals had either no effect on microbial populations or their activities, or had an effect that was followed by recovery after 1-3 weeks. Herbicides seem to have more short-term negative effects on soil microflora than insecticides. A few studies indicate that repeated application of the same pesticide may cause its rapid inactivation because of the enhanced growth of related specific decomposing microorganisms. This was observed in gamma-BHC, diazinon, aldicarb, and nitrophenols, but not in carbofuran and benthiocarb. Repeated application of a pesticide may also change the metabolic pattern of its decomposition. In the case of benthiocarb such a change produced a very phytotoxic compound (Moon and Kuwatsuka, 1984).

Because of the lack of field studies over several crop cycles there is no information on the long-term effects of pesticide use on wetland soil microflora. No method is yet available to quantify the biodiversity of soil microflora.

3.3. Effects on invertebrate populations

Studies of the effects of pesticide on floodwater populations show that insecticides are usually the most active compounds. Their application usually causes a general decrease in floodwater invertebrates followed by the proliferation of primary consumers -- ostracods, chironomid and mosquito larvae, and molluscs -- (Ishibashi and Ito, 1981; Roger and Kurihara, 1988), while populations of predators such as odonate larvae are reduced (Takamura and Yasuno, 1986). The rapid recovery of ostracods after pesticide application results from their resistance to pesticides and the large number of eggs they produce parthenogenetically (Lim and Wong 1986).

Nematodes and oligochaetes are probably the only soil invertebrates studied in wetland ricefields. Usually, the specific abundance of parasitic nematodes is higher in wetlands than in uplands, but apparently this results from submersion rather than from higher agrochemical use in wetlands (J. C. Prot, ORSTOM, pers. comm.). Benthiocarb had no marked effect on the number of nematode species and their average populations during the crop cycle (Ishibashi and Ito, 1981). Studies at IRRI showed a 70% reduction in soil oligochaetes populations when the amount of furadan applied was increased from 0.1 to 1.5 kg a.i. ha⁻¹ (I. Simpson, ODI, pers. comm.).

3.4. Effects on rice pests

The effects of new rice technologies on the carrying capacity of the ecosystem for insect pests were summarized by Heinrichs (1988). The availability of short-duration varieties and irrigation water has made rice cultivation throughout the year possible, thus eliminating fallow periods that often depress insect pests. Stable water supply has favored aquatic pests such as the caseworm *Nymphula depunctalis* (Heinrichs and Viajante, 1987). Increased N fertilizer use on responsive new varieties has favored brown planthopper (BPH) (Denno and Roderick, 1990). In general, BPH survive better, molt into larger adults, and are more fecund if they develop on N-rich host plants. The increased yield potential of modern varieties has also resulted in the misconception by farmers that greater returns will arise from pesticide application. Many agricultural authorities have thus subsidized and encouraged insecticide use. This in turn, resulted in pesticide misuse, accelerated development of resistance of pests, destruction of natural control, and pest resurgence and outbreaks.

The effects of rice production intensification on microbial pests have been summarized by Teng (1990). Crop intensification has generally resulted in increased prevalence, incidence, and severity of diseases caused by bacteria, viruses, and fungi. Bacterial blight (*Xanthomonas campestris* pv. *oryzae*) and sheath blight (*Rhizoctonia solani*) are directly attributable to cultural conditions of the modern high-yielding rice varieties which are grown with N fertilizer in large homogeneous areas. However, improved fertility associated with crop intensification has also resulted in the decrease of diseases such as brown spot (*Helminthosporium oryzae*). In areas with inefficient irrigation schemes, growing several crops a year has resulted in large areas with asynchronously planted rice, which is known to favor the devastating tungro virus. Disease epidemics cause instability in rice production over time because of the pathogen's ability to overcome resistance incorporated in rice varieties.

There is new evidence that crop intensification has no significant effect on the diversity of pathogen species in tropical rice. A study of 90 fields in the Philippines (Elazegui et al., 1990) showed that the number of pesticide and N fertilizer applications had no effect on the average number of pathogenic species encountered in the fields. However, transplanted rice was richer in pathogenic species and more diverse than direct seeded rice. A denser plant population might be less conducive for pathogen dispersal within a field.

3.5. Effects on vector-borne diseases

In traditional ricefields, although many vectors exist in the ecosystem, competition and predator pressure by fish and aquatic insects limit the productivity of any one vector. Ricefields contain a variety of insect predators of mosquito larvae such as backswimmers, gerrids, etc. (Hemiptera, Notonectidae), dragonfly and damselfly nymphs (Odonata), and adult and larval predacious water

beetles (Celeoptera, Dytiscidae) (Service 1977). Predator fauna vary according to rice cultivars, plant height, and water management (Mather and Trinh Ton That 1984).

In rice monoculture with a less diverse fauna and without control measures, the productivity of some vectors may be very high. Insecticides used to control rice pests and vectors may create secondary problems. The three major effects of insecticides as summarized by Roger and Bhuyian (1990) are (1) the temporary decrease in vector incidence since many agricultural insecticides are nonspecific and affect some vectors; extensive agricultural insecticide use probably explains the marked reduction of malaria and Japanese encephalitis in Japan after 1945; (2) the resurgence of resistant strains; in 1987, 50 malaria vectors resistant to one or more pesticides were recorded in the world; and (3) the adverse effects on the natural predators and competitors of vectors, causing blooming of mosquito larvae, and molluscs which are usually not affected by most rice pesticides and which multiply because of reduced predation or competition for food.

4. USE OF MICROBIAL AND INVERTEBRATE BIODIVERSITY TO ENHANCE AGRICULTURAL SUSTAINABILITY OF WETLAND SOILS

4.1. Microbial management of wetland rice soils

The microbial management of wetland soils was recently reviewed by Roger et al. (1991). Using biological N_2 fixation as an alternative or supplementary N source for rice has been the major approach. Whereas N_2 -fixing green manures (azolla and legumes) have been used for centuries in some rice-growing areas, research on N_2 -fixing BGA and bacterial inoculants for wetland rice is relatively recent, being initiated in the early 1950s for BGA and in the 1960s for bacteria.

Biomass estimates, N₂-fixation measurements, and inoculation experiments indicate that BGA, as an additional N source for rice, has a potential of 20-30 kg N ha⁻¹ crop⁻¹ which may translate to a yield increase of 200-350 kg ha⁻¹. Recent data show that BGA are ubiquitous in rice soils and that foreign strains usually do not get established in the field. Thus, the principle of BGA inoculation should be reconsidered and more attention should be paid in promoting indigenous strains.

Reported effects of bacterial inoculation of rice have been inconsistent. Most strains tested had been N₂-fixing forms, but there had been no clear evidence that promotion of rice growth and N uptake was due to increased biological N₂ fixation. Therefore, several authors refer to the production of plant growth regulators to explain the beneficial effect of bacterial inoculation. No experiment has yet supported this hypothesis. The few data on strain establishment showed that, in most cases, inoculated strains did not multiply. Given the current status of knowledge on bacterial inoculation of rice, no positive conclusion can be drawn on its potential.

There are several reports on the existence of varietal differences in the ability to support associative BNF (Nfs character). The idea of breeding varieties with higher N_2 -fixing potential (Nfs) is attractive since it would enhance BNF without additional cultural practices. This promising approach is still limited by the lack of an efficient screening method.

4. 2. Use of biodiversity to control insect pests

Natural parasitoids, predators, and pathogens that attack insect pests are abundant in ricefields. Outbreaks occur when the equilibrium is disrupted and the full reproductive capacities of pest species are released. Thus, enhancing the action of natural control agents is of paramount importance in an integrated pest management (IPM) program. The various aspects of IPM in Asia were recently reviewed (Teng and Heong, 1988).

Pesticides can markedly affect natural enemy fauna and should be used judiciously. In a study comparing 330 crops in insecticide-treated and untreated fields, only 50% of the fields showed measurable yield losses due to pests (Litsinger, 1984). The removal of pesticide subsidies in Indonesia led to a drastic decrease in pesticide use with no measurable reduction in the national yield average (Kenmore 1989). Recent data in the Philippines also showed no significant difference in average yields between farms using and not using pesticides (Elazegui et al., 1990).

IPM developed from the concept of integrating control tactics into an acceptable system. This tactics include ways to maximize natural control, and the use of resistant varieties and chemicals only when necessary. It is sometimes defined as the farmers' "best mix" of control tactics based on crop yield, profit, and safety (Kenmore et al., 1985). Since IPM advocates the conservation of natural enemy populations in ricefields, more basic knowledge on population dynamics and trophic linkages is needed. Increasing general biodiversity per se may not be sufficient to promote ecosystem stability and sustain low-pest situations. A recent study of arthropod diversity in Philippine ricefields has confirmed this hypothesis. Of the five locations where samples were collected, the IRRI experimental farm had significantly higher biodiversity (Fig. 1), but pest population was also significantly higher (Table 3). Lowest biodiversity and phytophages population were recorded in traditional rice terraces.

4.3. Use of biodiversity to control rice diseases

Natural control is not as effective with pathogens as with insect pests, but there is evidence that antagonistic bacteria can control some of the agents of rice diseases. Rosales et al (1986) found that 60% of 139 ricefield bacterial isolates inhibited the *in vitro* growth of *Fusarium moniliforme* (agent of bakanae disease). Disease control by seed treatment of 18 bacteria isolates tested in artificially infested nurseries ranged from 70 to 96%. Mew and Rosales (1986)

reported reductions in sheath blight severity by bacterization with *Pseudomonas* and *Bacillus* strains. These methods are not currently used in the field.

Spatial and temporal diversity of rice is a well-known strategy for disease control. Blast has been controlled by using variety mixtures (Bonman et al., 1986) and one control of tungro is by rotation of host genes conferring resistance (Manwan et al., 1987). The principle for using host genetic diversity to control pathogens depends on an understanding of the population genetics of the pathogen and the frequency of virulence genes in different ecosystems. While much of the theory has been developed, few empirical data yet exist in the tropics to extend the concept to pathogens other than blast and tungro.

4.4. Use of biodiversity to control vector-borne diseases

Biological control of vectors has basically two major approaches: (1) maintaining species diversity and thus conserving natural predators, and (2) introducing new predators, competitors, parasites, or diseases of vectors.

Most of the information on conservation of natural predators of vectors refers to mosquitoes (Roger and Bhuiyan, 1990). Because of predators such as fish, Odonata, Notonectidae, and Discidae, survival percentage of mosquito larvae in ricefields from the first-instar through the pupal stage varies from 2 to 5%. Spiders also reduce the number of adult mosquitoes. Despite very high predation on larvae, there may still be large numbers of adults emerging and constituting a nuisance or disease hazard. If natural predators are destroyed, emerging mosquitoes are likely to be more numerous. Even if it is not envisaged to use predators as control agents, cultural practices favoring their existence should be encouraged (Mather and Trinh Ton That 1984).

Numerous competitor and predator species have been tested to control vectors. The most promising method for mosquito control is to stock food fish in and around ricefields. It reduces vector and weed incidence, increases rice yields, partly because of the fish excreta, and produces fish food (Self 1987). In the Philippines, the combined culture of larvivorous tilapia and common carp in ricefields, with supplemental feeding, produced about 700 kg fish ha-1 year-1 (Petr 1987). However, experience in the efficiency of introduced larvivorous exotic fish has been varied (Roger and Bhuiyan, 1990). Main constraints to ricefield fisheries and vector control by fish are the toxicity of agrochemicals, especially pesticides, and unreliable water supply.

The use of competitors is a strategy that seems to have been restricted to snail vectors. Large snails such as *Marisa* and *Thiara* were successfully introduced and supplanted schistosome vector snails in ponds and canals in several countries, but some of these large snails feed on transplanted rice seedlings (Roger and Bhuiyan, 1990).

Microbial agents with a potential to control mosquitoes include virus, bacteria, and fungi.

Bacillus thuringiensis serotype H-14 and several virulent strains of Bacillus sphaericus (Dame

et al. 1988) provide selective control of mosquito larvae, while causing relatively little harm to most of the predators of vectors and agricultural pests. Currently, only *B. thuringiensis* is used and commercialized.

Probably, the least attention to date has been given to insect predators. Their taxonomy and ecology need to be studied before their possible use in integrated pest control can be assessed and ways of multiplying them evaluated (Schaefer and Meisch 1988).

5. PRESERVATION OF MICROORGANISM BIODIVERSITY IN GERMPLASMS AND POTENTIAL OF BIOTECHNOLOGIES FOR THEIR UTILIZATION

While more than 85 000 accessions of rice are kept in IRRI's rice germplasm bank, other components of the ecosystem preserved in a living state include a few hundred N₂- fixing organisms (azolla, BGA, bacteria) with potential use as biofertilizers and a few hundred strains of rice pathogens isolated from the Philippines. There is no collection of invertebrate germplasms.

5. 1. Improved strains of N₂-fixing agents

With regard to the rapid progress in genetic engineering, one can speculate on the possibilities to select or design efficient strains of N_2 -fixing organisms.

Several authors have selected BGA with high N₂-fixing activity. A nitrogenase- depressed *Anabaena* mutant that excretes NH₄+ into the medium was found to provide N to rice in a N-free gnotobiotic culture more efficiently than the parent strain (Latorre et al., 1986). But in both cases, strains could not establish themselves in the soil, which is consistent with the observation that no inoculation experiment has yet reported the establishment of foreign BGA strains in soils (Roger et al. 1991). Significant progress has been made on BGA genetics, and "super N₂-fixing BGA" can be selected or probably designed and grown *in vitro*, but the characteristics that will enable them to survive, develop, and fix N₂ as programmed *in situ* are still unknown.

Azolla collections have been used to screen varieties adapted to specific environments, while some efficient strains have been adopted for practical use. These collections have also been used for achieving exchanges of algal symbionts between species, and for breeding improved *Azolla* hybrids (Roger et al. 1991).

5.2. Characterization of inter- and intraspecific diversity of rice pathogens Sustainable rice ecosystems need to provide, *inter alia*, economic stability to the farmer as demonstrated by stable rice yields. One of the main causes of instability are the sporadic epidemics caused by subpopulations of plant pathogens increasing in frequency relative to the prevailing rice genotype. This results in the so-called "boom-and-bust" cycles of varietal

resistance breakdown (Teng 1990). Understanding the co-evolutionary processes between pathogen populations and rice requires the monitoring of pathogen species at the community level, which was constrained historically by the lack of rapid and accurate methods of characterization. Pathogen identification commonly requires time-consuming axenization of the pathogen to fulfill Koch's postulates. Tools such as polyclonal and monoclonal antibodies have reduced the time needed for identification, but antibodies have only been developed against a few rice pathogen species and this is an area which requires greater effort.

Within the same species, pathogen populations also exhibit much genetic variability on the same rice genotype. The traditional method of studying such variability has been to "type" the subpopulations using a set of rice differential genotypes representing a range of susceptibilities to the pathogen. With modern molecular markers and selected serological techniques, the tedious and complicated process of race typing has been greatly simplified and made more reliable. Intraspecific diversity of *Xanthomonas campestris* has been recently studied at the DNA level through the detection of restriction fragment length polymorphism (RFLP) (Raymundo et al, 1990). DNA probes were used to study the partitioning of variability in pathogen populations. Several putative transposable elements were also identified and used to examine the DNA profiles of a collection of strains. The results indicate a particular evolutionary relationship between pathogenic races and rice host resistance. The intraspecific diversity revealed by RFLP typing allows the selection for appropriate tester strains to identify unrecognized races and resistance genes.

6. CONCLUSION

The beneficial and detrimental roles of many groups of microorganisms and invertebrates in sustainable rice production have been identified, and sometimes, quantified. However, knowledge on the possible long-term effects associated with the intensification of rice cultivation on these populations is limited. The study of crop intensification effects in long-term experiments should have high priority. In particular, the N fertility of wetland rice soils depends upon the turnover of a soil microbial biomass representing only a very small percentage of total soil N. Therefore, a general understanding of (1) the pathways that allow microbial biomass replenishment and (2) the long-term effects of crop intensification on microbial and invertebrate populations involved in this replenishment is needed.

Estimates of biodiversity in ricefields are extremely scarce; there are no irrefutable data to demonstrate the generally accepted concept that crop intensification decreases biodiversity in ricefields.

Recent data on arthropods confirm that high biodiversity is not synonymous with stability and with low pressure of insect pests. Increases in pest and vector densities may depend primarily

on reduced predator diversity and the resilience and biological attributes of a particular pest. Thus, increasing or preserving diversity *per se* does not necessarily contribute to pest stability, but developing effective trophic linkages might. This approach might also be valid for maintaining soil fertility through microbial management, the optimization of the primary production in floodwater, and the optimization of nutrient recycling by invertebrate populations.

Numerous methods using of microorganisms and invertebrates to increase soil fertility (especially through biological N fixation) and to control pests and diseases have been tested. But the success of these methods is limited and their adoption is almost negligible. This situation will probably remain unchanged as long as the methods designed are based on an extremely restricted knowledge on biodiversity, community structure, and trophic relationships at the ecosystem level, as it is currently.

However, current knowledge shows that there is potential in designing methods that use microbial and invertebrate populations to sustain a management system that reduces and optimizes agrochemical use.

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Table 1. Estimates of losses due to rice pests (adapted from Teng, 1990)

Agent	% loss	Location	Reference ¹
Incost nosts	,		
Insect pests All insects	24%	Asia	(Ahrens et al. ,1982)
7th mocoto	35% - 44%	riola	(Pathak & Dhaliwal, 1981),
	35%	India	(Way, 1976),
	16% - 30%	Philippines	(Way, 1976).
	6%	Bangladesh	(Alam, 1961).
	10% - 20%	Sri Lanka	(Fernando (1966)
Rice stem borers	30-70%	Bangladesh ² ,	(Alam et al., 1972)
	3-20%	Bangladesh ³ ,	(Alam, 1961)
	3-95%	India	(Ghose et al., 1960)
	up to 95%	Indonesia	(Soenardi, 1967)
	33%	Malaysia	(Wyatt, 1957)
Leafhoppers	50% - 80%	Bangladesh	(Alam, 1961)
Brown planthopper	1% - 33%	India	(Jeyaraj et al, 1974).
Rice bugs Leptocorisa	10%		(Pruthi, 1953).
Gall midge larvae	12% - 35%	India	(Reddy 1967),
Pachydiplosis oryzae	50% - 100%	Vietnam	(Reddy 1967)
Rice hispa	10% - 65%	Bangladesh	(Barr et al., 1975)
Dicladispa armigera	up to 50%	India	(Barr et al., 1975).
Leaffolders	up to 50%	India	(Balasubramaniam et al., 1973).
<u>Diseases</u>			
Blast	up to 100%		(Teng et al, 1990)
	5% - 10%	India	(Padmanabhan 1965),
	3%	Japan	(Teng, 1990)
	8- 14%	China	(Teng, 1986).
Brown spot	80%	India	(Padmanabhan, 1973)
·	14% - 41%	India	(Vidhyasekaran & Ramados, 1973)
Sheath blight	9-13%	China	(Teng, 1986).
Tungro virus	100%	Indonesia	(Chang et al. 1985)
	40% - 60%	Bangladesh	(Reddy, 1973)
	50%	Thailand	(Wathanakul & Weerapat, 1969)
	30%	Philippines	(Teng, 1990)
Bacterial blight	up to 60%	India	(Srivastava, 1972)
	5-6 %	China	(Teng, 1986).
Stem rot	5% - 10%	India	(Chauhan et al., 1968)
	5-6 %	China	(Teng, 1986).

¹ references are listed in Teng (1990); ² outbreak; ³ chronic.

Table 2. Summary of quantitative records of species/taxa in wetland ricefields

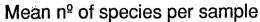
1 Number of engine records	d by Usale	man in 1075 in a 1 vas	r study of	a cinale				
1. Number of species recorder field in northeastern Thailan			study of	a single				
meid in northeastern rhanait	u (o saiii)	ings)						
Sarcodina	3 1	Cyanophyta	11					
Ciliata	83	Algae	166					
Rotifers	50	Pteridopyta	3					
Platyhelminthes	7	Monocotyledonae	25					
	7		10					
Nematoda		Dicotyledonae	18					
Annelida	11	Pisces	. •					
Mollusca	12	Amphibia/Reptilia	10					
Arthropoda	146	TOTAL	<u>590</u>					
2. Number of species/taxa of	aquatic i	nvertebrates, excluding	protozoas,	recorded				
by different authors	-							
· Heckman (1979) (species), one	traditional	field, 1-year study (Thaila	nd) 1	83				
• Lim (1980) (taxa), 2-year study of pesticide application (Malaysia)39								
• Takahashi et al. (1982) (taxa) 4 fields, single samplings (California)								
• IRRI 1985 and Roger et al. 1985								
with pesticide applied (Philipp				26				
with position applied (i timp	moo and m	<i>σ.</i> α,						
3. Records of arthropod speci	oe in ricof	ialde over one cron	cycle					
Kobayashi et al (1973) : study ii			aping	450				
(Shikoku, Japan)				450				
 Heong et al. (unpubl. data): stud 	ly in 1989 of	5 ricetields by suction						
(Philippines):								
	Fiv	e fields combined		.240				

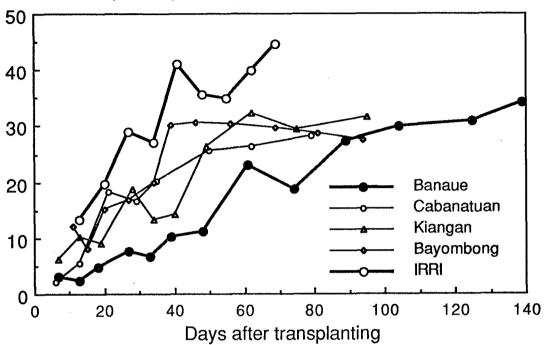
Table 3. Arthropod community structure in five ricefields in the Philippines*.

Sampling site	IRRI	Bayombong			Banaue
J		Canabatuan Farmer fields		Kiangan	
nature	experimental farm			rice terrace	
Species nº	31	16	20	17	8
Abundant spp. nº	13	5	11	10	6
% contribution	82	8 0	60	73	59
Evenness	0.58	0.52	0.62	0.70	0.86
Total nº x 1000	22	15	8	11	6
% phytophages	6 4	45	43	45	57
% predators	26	53	52	5 1	35

^{*} Average values per sampling . Heong et al. unpublished

Fig. 1. Dynamics of arthropod diversity during a crop cycle in five ricefields in the Philippines.





* Banaue: rice terrace where rice has been grown without agrochemicals for centuries.

Cabanatuan, Kiangan, Bayombong: farmers' fields with agrochemical use.

IRRI: experimental plot on the IRRI farm.