Effect of natural naphthoquinones in BALB/c mice infected with Leishmania amazonensis and L. venezuelensis

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Abstract

Plumbagin, 3,3'-biplumbagin and 8,8'-biplumbagin are naphthoquinones isolated by activity-directed fractionation from a Bolivian plant, Pera benensis, used in folk medicine as treatment of cutaneous leishmaniasis caused by Leishmania braziliensis. BALB/c mice were infected with L. mexicana or L. venezuelensis and treated 24 h after the parasitic infection with plumbagin (5 or 2.5 mg/kg /day), 3,3'-biplumbagin, 8,8'-biplumbagin (25 mg/kg/d) or Glucantime[®] (200 mg/kg/d). Lesion development was the criteria employed to evaluate the inhibitory effect. The bisnaphthoquinones were less potent than Glucantime against amazonensis and L. venezuelensis. Plumbagin and Glucantime delayed the development of L. amazonensis and L. venezuelensis. Assays of a single local treatment on footpad infection two weeks after the parasitic inoculation with L. amazonensis showed that 8,8'-biplumbagin (50 mg/kg/d) was as potent as Glucantime (400 mg/kg/d).

Introduction

Cutaneous and mucocutaneous leishmaniasis are endemic diseases in the tropical subandean regions. Cutaneous leishmaniasis is popularly known as espundia in the area of Bolivia called Oriente by the natives. The use of medicital plants for the specific treatment of cutaneous leishmaniasis is quite widespread, specially Pera benensis (Eurhorbiaceas). The fresh stem barks are applied directly on the lesion.

We have previously reported the study of the chemical identification of active compounds and the leishmanial and trypanocidal activities in vitro of three active naphthoquinones (Fig. 1), plumbagin, 3,3'-biplumbagin and 8,8'-biplumbagin (Fournet et al., 1990). These compounds isolated by activity-directed fractionation from the stem barks and root barks of Pera benensis, displayed activity in vitro at 10 µg/ml against three strains of promastigote forms of Leishmania species, L. amazonensis (PH 8 and H-142), L. braziliensis (M)2903) and L. donovani (2682) and six strains of epimastigete forms of Trypanosoma cruzi. Plumbagin was also active

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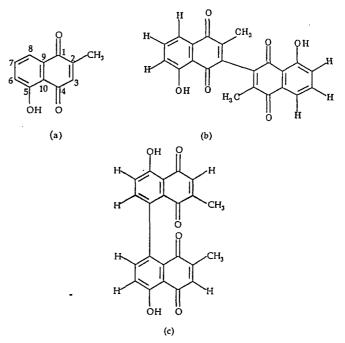


Fig. 1 Structures of (a) plumbagin; (b) 3,3'-biplumbagin; (c) 8.8'-biplumbagin

against amastigote forms of L. amazonensis (PH 8) infecting the mouse peritoneal macrophages.

The aims of this present paper were to evaluate the activity of naphthoguinones in BALB/c mice infected with L. amazonensis or with L. venezuelensis, two species of American cutaneous leishmaniasis. We have used L. venezuelensis because this parasite produces for the hamster a rapid growing granuloma at the site of inoculation, containing abundant amastigotes. After a few months, we have observed necrosis on the nose and the head of the animal.

The mouse footpad infection has been used as model for these experiments (Avila et al., 1990; Coleman et al.,

Materials and methods

Animals

Female or male BALB/c mice were supplied by Charles River Breeding Laboratory and then were bred in IBBA (Bolivia). Mice weighed 18-20 g and eight weeks old when bioassays were initiated.

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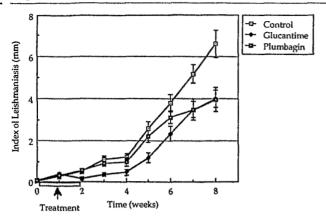


Fig. 2 Effects of plumbagin (2.5 mg/kg/d) and Glucantime (200 mg/kg/d) on the development of *L. amazonensis* (PH 8) in BALB/c mice (n = 10,-/+ S.E.M.). Treatments were given for 14 d period commencing 1 d after inoculation of *L. amazonensis* S.E.M. = -/+ Standard error of the mean

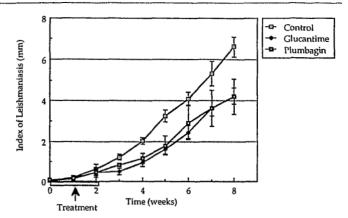


Fig. 3 Effects of plumbagin (5 mg/kg/d) and Glucantime (200 mg/kg/d) on the development of L. venezuelensis (H-3) in BALB/c mice (n = 6, -/+ SEM). Treatments were given for 14 d period commencing 1 d after inculation of L. venezuelensis.

Table 1 Effect of Glucantime (200 mg/kg/d), 3,3'-biplumbagin (25 mg/kg/d), 8,8'-biplumbagin (25 mg/kg/d) on the development of L. amazonensis (PH 8) in BALB/c mice (-/+SEM). Drug were given for 14 d-period commencing 1 d after inoculation of L. amazonensis.

Weeks post infection	Control**	Diameter of les Glucantime**	sion* 3,3'- biplumbagin**	8,8'- biplumbagin**	
2	0.3 (0.15)	0.3 (0.08)	0.5 (0.13)	0.4 (0.11)	
4	1.6 (0.3)	0.9 (0.23)	1.4 (0.34)	1.08 (0.28)	
6	3.8 (0.6)	1.7 (0.52)	2. 7 (0.55)	2.6 (0.33)	
8	6.0 (0.88)	3.23 (0.87)	4.58 (0.84)	5.05 (0.62)	
9	6.9 (0.70)	3.66 (1.03)	5.05 (0.83)	5.63 (0.74)	

^{*}Average measurement (in mm) for 8 mice. ** -/+ Standard error of the mean (S.E.M.)

Leishmania strains

L. amazonensis (IFLA/BR/67/PH 8) and L. venezuelensis (VE/74/PM-H3) were used. The source and history of this isolate have been described by Bonfante-Garrido (1983). BALB/c mice (n = 10, n = 8 or n = 6) were infected subcutaneously in the right rear footpad with 1×10^6 amastigotes obtained from infected hamsters. The parasites were delivered in 200 μ l phosphate buffered saline (PBS), with control mice receiving PBS only.

The growth of the lesion was determined weekly by measuring the diameter of both rear feet with a direct reading vernier caliper (Ref: Kroelin 10DI 00T6). The size of lesion in millimeters (Index of Leishmaniasis) was calculated by subtracting the measurements obtained for the uninfected foot from that the infected foot. Measurements started one day prior to the inoculation of amastigotes and continued for 8 or 9 weeks.

Drug treatment

Two experiments were conducted. Mice in the first experiment were treated by subcutaneous route. Glucantime was given at a dose of 200 mg/kg/d, plumbagin at 5 or 2.5 mg/kg/d, 3,3'-biplumbagin and 8,8'-biplumbagin at 25 mg/kg/d. Drug treatment started one day after the inoculation of amastigotes and continued once daily for 14 days.

In the second experiment, mice were treated directly on the infected rear footpad with a single dose 14 days after inoculation of parasites. For this experiment, mice were treated with Glucantime at

400 mg/kg/d, plumbagin at 10 mg/kg/d, 3,3'-biplumbagin and 8,8'-bi-plumbagin at 50 mg/kg/d.

The naphthoquinones were dissolved in 40 µl of polysorbate (Tween 80, Prolabo). For each experiment was calculated the mean and standard error of the mean (S.E.M.).

Results

Activity of naphthoquinones of Leishmania amazonensis

Separate experiments were conducted in which plumbagin was administered at different doses (7.5, 5, and 2.5 mg/kg daily) beginning 24 hr prior to infection with L. amazonensis (PH 8). Mice treated with 7.5 mg died within four weeks after the beginning of experiment. Figure 2 shows the combined results obtained with mice treated with 2.5 mg/kg daily of plumbagin compared to mice treated with 200 mg/kg daily of Glucantime. After eight weeks mice treated with plumbagin or with Glucantime had an average lesion size of 4 mm compared with 6.8 mm for control. We did not observe toxic effect of plumbagin at 2.5 mg/kg daily. We have obtained the same effects when plumbagin was administered at 5 mg/kg.

Table 1 presents the experiments of treatment with bis-naphthoquinones, 3,3'-biplumbagin and 8,8'-biplumbagin. These compounds were less toxic than plumbagin but less efficient at 25 mg/kg daily. After nine weeks, mice treated

infected with L. amazonensis (LV/78) or L. donovani (LV9). A bis-naphthoquinone isolated from the Indian plant, diospyrine, a dimer of 7-methyl-juglone is also active in vitro against L. donovani (Hazra et al., 1987). Several authors have reported an antiprotozoal activity of naphthoquinones (Callahan et al., 1988; Pinto et al., 1987; Wright and Phillipson, 1990), particularly of lapachol and β-lapachone isolated from Tabebuia rosea (Bignoniaceae) against Plasmodium falciparum (Carvalho et al., 1988) and against Trypanosoma cruzi (Goncalves et al., 1980), and a derived of lapachone, lapinone (Hudson et al., 1985) against Plasmodium vivax. Recently synthetic naphthoquinones (566C80) have been described as active against Pneumocystis carinii (Wellcome Foundation, 1990). Several authors have described the activity of naphthoquinones against skin diseases, plumbagin (Gujar, 1990) and 2-hydroxy-1,4naphthoquinones for prevention of dermatitis on the scalps (Tsucha and Yutaka, 1990).

In conclusion, the results of this study show that treatment with a topical application directly on the lesion of leishmaniasis of stem barks of *Pera benensis* may be effective against leishmaniasis. It could be possible to propose an effective ointment prepared locally with a low concentration of plumbagin or an other derived of this napthoquinone less toxic as 8,8'-biplumbagin. These formulations would be developed in endemic regions of cutaneous and mucocutaneous leishmaniasis, in particular in areas of colonization of Bolivia when occure the lack of usual drugs as pentavalent antimonials.

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