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JEREISTEROL A AND B : TWO 3β -METHOXY-SECOSTEROIDS FROM THE PACIFIC SPONGE *JEREICOPSIS GRAPHIDIOPHORA*

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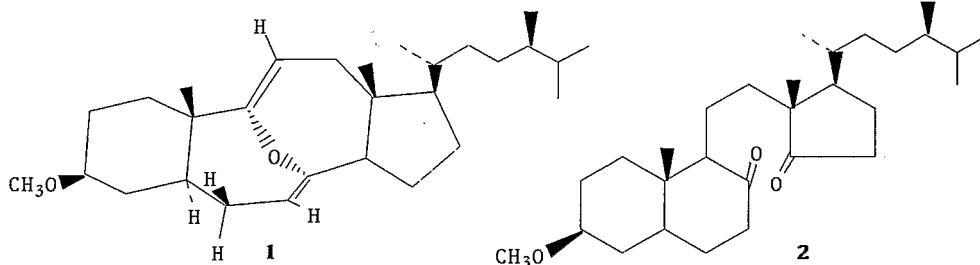
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Summary: Two 3β -methoxy secosteroids, named jereisterol A and B were isolated from the pacific sponge *Jereicopsis graphidiophora* Lévi & Lévi. Their structures, which combine rare 3β -methoxy and seco features, were determined as (24 R) 24-methyl- 3β -methoxy- $8\alpha,9\alpha$ -oxido- $8,9$ -secocyclosta-7,9(11)-diene (1) and (24R) 24-methyl- 3β -methoxy- $8,14$ -secocyclosta-8,14-dione (2).

Although a large number of new sterol structures have been discovered from marine organisms during the last twenty years¹, very few secosteroids have been reported. After the discovery in 1972 of the unique $3\beta,11$ -dihydroxy- $9,11$ -secogorgost-5-en-9-one from a gorgonian², three new $9,11$ -secosterols were isolated from the soft coral *Sinularia* sp.³, a polhydroxylated $9,11$ -secosterol from the sponge *Disidea herbacea*⁴ and a trihydroxylated $5,6$ -secosterol has been described from the sponge *Hippopsgorgia communis*⁵. Very recently an unique group of norsterols, the incisterols, derived from biodegradation of the sterol nucleus with loss of the ring A including 19-methyl group, has been isolated from the sponge *Dictyonella incisa*⁶, and the Authors have proposed these degraded sterols to originate biogenetically from a $5,6$ -secosterol through the further cleavage of the $9,10$ bond.

During our on going program to isolate novel bioactive molecules from New Caledonian marine invertebrates, the opportunity occurred recently to examine the lipidic extracts of the sponge *Jereicopsis graphidiophora*, Lévi & Lévi⁷. In a related paper⁸ we have reported the occurrence in *J. graphidiophora* of the first 3β -O-methylsterols, while the conventional 3β -hydroxysteroids were totally absent. Herein we describe the isolation and structure determination of two more 3β -methoxysteroids, jereisterol A (1) and B (2), which combine the 3β -methoxyl group with a rare seco-structure.

The sponge was ground, freeze-dried (0.7 K.) and extracted with *n*-hexane. Silica gel chromatography of the extract using increasing amounts of ethyl acetate in *n*-hexane gave a major fraction containing 3β -methoxysteroids together



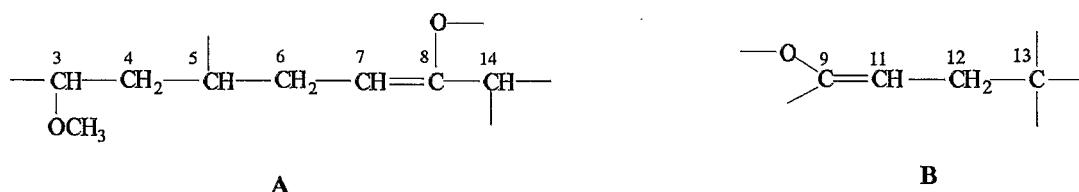
with some more polar fractions containing the oxygenated 3β -methoxysteroids.

Reverse-phase HPLC (Whatman Partisil ODS-2) of the fraction eluted with *n*-hexane/EtOAc 98:2 yielded, with MeOH/CH₂Cl₂ 9:1, jereisterol A (1, 0.8 mg), while HPLC with MeOH/H₂O 95:5 of the fraction eluted with *n*-hexane/EtOAc 9:1 yielded jereisterol B (2, 5 mg).

Jereisterol A (1) showed a molecular ion (EI) at *m/z* 428.3650 (C₂₉H₄₈O₂) and a major fragment at *m/z* 301.2167 (C₂₀H₂₉O₂) due to the loss of a C₉H₁₉ fragment and suggesting a conventional saturated steroid C₉-side chain. The ¹H-NMR spectrum also indicated a steroid structure. The methyl signals at δ 0.53 (3H, s), 0.79 (3H, d, *J*=7.0), 0.81 (3H, d, *J*=7.0), 0.86 (3H, d, *J*=7.0), 0.98 (3H, d, *J*=6.5) and 1.09 (3H, s) could be assigned to carbons 18, 26, 27, 28, 21 and 19 respectively of a "ergostane" skeleton. The 1H multiplet at δ 3.22 ppm and the 3H singlet at δ 3.36 are assigned to a 3β -methoxyl group. Remaining relevant ¹H NMR data, are summarized in Figure 1.

Sequential decoupling experiments identified the structural moieties A (C-3 to C-14) and B (C-9 to C-13). Evidence to connect the structural part from C-3 to C-7 and C-14, through a quaternary carbon, was provided by a clearly detected homoallylic coupling between the H-6 α signal at δ 2.12 ppm and the broad doublet downfield shifted to δ 2.38 ppm (H-14).

The ¹³C NMR spectrum⁹, even if quite poor, contained signals for rings A and B and for the side chain, and revealed



A

B

-OCH ₃	:	3.36 s			
H-3	:	3.22 m			
H-4 α	:	1.78 brd (<i>J</i> =12.5 Hz)			
H-4 β	:	1.95 m ^a			
H-5	:	2.79 tt (<i>J</i> =13.2, 3.7 Hz)			
H-6 α	:	2.12 brdt (<i>J</i> =18.4, 3.7 Hz)			
H-6 β	:	1.95 m ^a			
H-7	:	4.53 t (<i>J</i> =3.7 Hz)			
H-14	:	2.38 brdd (<i>J</i> =11.8, 5.9 Hz)			
			H-11	:	4.92 dd (<i>J</i> =8, 6 Hz)
			H-12	:	2.31 dd (<i>J</i> =14, 8 Hz)
					2.21 dd (<i>J</i> =14, 6 Hz)

Fig.1 ¹H NMR data (CDCl₃, 500 MHz) of jereisterol A (1)
^a overlapping signals

two sp^2 CH signals at relatively high field (δ 106.8 and 107.2 ppm), which, along with the 1H NMR high field shifted olefinic signals at δ 4.92 and 4.53 ppm, indicated the presence of enol ether structure.

Thus, the remaining oxygen atom indicated by MS was fixed between the olefin carbons C-8 and C-9 giving rise to the unique 24-methyl-3 β -methoxy-8 α ,9 α -oxido-8,9-secocholesta-7,9(11)-diene structure. The 8 α ,9 α -oxido stereochemistry is suggested by the chemical shift of the proton of C-5 downfield shifted to δ 2.79 ppm implying the oxygen function and H-5 to be located on same face of the molecule. The 24R stereochemistry is assigned by comparison of 1H and ^{13}C NMR spectra with known compounds¹⁰.

Jereisterol B (2). The molecular formula $C_{29}H_{50}O_3$ was established by MS, m/z 446.3745 (M^+). The MS spectrum also displayed, aside the molecular ion peak, an intense ion at m/z 319.2268 ($C_{20}H_{31}O_3$, due to the loss of a C_9H_{19} side chain), thereby locating all three oxygen functions in the nucleus.

The 1H NMR and ^{13}C NMR spectra¹¹ indicated a "3 β -methoxy-24-methylcholestane" skeleton. Two IR carbonyl bands at 1727 and 1700 cm^{-1} and the ^{13}C NMR signals at 224.0 and 214.0 ppm indicated the presence of two ketone groups in 2, which must be incorporated into a steroidal tricyclic nucleus to account for the molecular formula $C_{29}H_{50}O_3$, requiring five unsaturations. All the data can readily be explained if the two ketone groups are at C-8 and C-14 of a 8,14-seco steroid. The structural deduction was confirmed by synthesis of the model 3 β -acetoxy-8,14-secoergostane-8,14-dione. The synthesis was accomplished by oxidation with ruthenium tetroxide of 3 β -acetoxy-ergost-8(14)-ene¹² by using the procedure described by Sharpless *et al.*¹³ to yield the desired 8,14 seco-compound¹⁴ along with major amounts of the product of allylic oxidation, 3 β -acetoxy-ergost-8(14)-en-15-one, identified by comparison with published spectral data for the 3 β -hydroxycholesta-8(14)-en-15-one¹⁵.

Comparison of the 1H NMR data of 2¹¹ with those of the synthetic 3 β -acetoxy-8,14-secoergostane-8,14-dione¹⁴ showed identical chemical shift values for C-18, C-19 and C-21 methyl resonances; the small differences observed in the pattern of the C-26, C-27 and C-28 methyl resonances gives strong support to the assignment of the 24R-configuration to 2 (the synthetic model has the 24S-configuration). Comparison of the ^{13}C NMR data of 2 with those of the model equally supports the 8,14-seco steroid-8,14-dione structure and the 24R-configuration for the natural compound.

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7) The sponge was collected in course of the Campaign MUSORSTOM IV of the ORSTOM-CNRS Programme "Substances Marine d'Interet Biologiques" (SMIB) in the North of New Caledonia at a depth of 225 m. A sample is kept at the ORSTOM Centre de Nouméa under the reference R 1369. The sponge has been identified by C.Lévi as a new genus; C.Lévi and P.Lévi, *Bull. Mus. Natn. Hist. Nat. Paris* IV series, 5, 101 1983.

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9) ^{13}C nmr data (CDCl_3) of jereisterol A (1); C-1: 36.0, C-2: 27.0, C-3: 79.5, C-4: 33.0, C-5: 37.8, C-6: 34.8, C-7: 106.8*, C-11: 107.2*, C-12: 38.3, C-13: 44.5, C-14: 59.5, C-15: 22.2, C-16: 27.9, C-17: 53.6, C-18: 11.9, C-19: 17.5, C-20: 36.0, C-21: 19.7, C-22: 33.8, C-23: 31.0, C-24: 39.1, C-25: 32.5, C-26 and C-27: 18.3 and 20.2, C-28: 15.5 OCH_3 ; 55.6 ppm. Signals for quaternary C-8, C-9 and C-10 confused in the noise.

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11) Spectral data of jereisterol B (2): IR (CHCl_3): 2950, 2936, 2870, 1727, 1700, 1460, 1371, 1213, 1029 cm^{-1} ; ^1H -nmr (CDCl_3): 0.61 (3H, s, CH_3 -18), 0.81, 0.82, 0.87 (each 3H, d, J = 6.5 Hz, 7.0 and 7.0 Hz, CH_3 -26, 27 and 28), 0.86 (3H, s, CH_3 -19), 1.08 (3H, d, J = 6.5 Hz, CH_3 -21), 2.36 (1H, m, CH), 3.20 (1H, m, H-3), 3.36 (3H, s, OCH_3); ^{13}C nmr (CDCl_3); C-1: 36.6, C-2: 27.2, C-3: 79.5, C-4: 32.4, C-5: 43.8, C-6: 30.1, C-7: 42.1, C-8: 211.0, C-9: 63.4, C-10: 41.6, C-11: 17.7, C-12: 37.6, C-13: 52.6, C-14: 224.0, C-15: 37.9, C-16: 23.6, C-17: 47.1, C-18: 18.4, C-19: 12.4, C-20: 34.5, C-21: 18.5, C-22: 34.0, C-23: 31.0, C-24: 39.1, C-25: 32.5, C-26: 20.4, C-27: 18.4, C-28: 15.5, OCH_3 ; 55.7 ppm.

12) prepared from ergosterol acetate by hydrogenation at 3 atm for 20 h at room temperature in the presence of platinum oxide according to W.H.Lee, B.N.Lutsky and G.J.Schroepfer, *J. Biol. Chem.* 244, 5440 (1969).

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14) Spectral data of the synthetic 3β -acetoxy-8,14-secoergostane-8,14-dione:
 ^1H -nmr (CDCl_3): 0.63 (3H, s, CH_3 -18), 0.81 (6H, d, J = 6.7 Hz), 0.87 (3H, d, J = 7.0 Hz, CH_3 -26, 27 and 28), 0.86 (3H, s, CH_3 -19), 1.08 (3H, d, J = 6.5 Hz, CH_3 -21), 2.03 (3H, s, CH_3CO), 2.35 (1H, m, CH), 4.75 (1H, m, 3-H); ^{13}C nmr (CDCl_3); C-1: 36.5, C-2: 26.9, C-3: 73.0, C-4: 32.4, C-5: 43.6, C-6: 30.1, C-7: 41.9, C-8: 210.7, C-9: 63.0, C-10: 41.6, C-11: 17.6, C-12: 37.6, C-13: 52.5, C-14: 223.9, C-15: 37.8, C-16: 23.5, C-17: 47.1, C-18: 18.4, C-19: 12.4, C-20: 34.8, C-21: 18.6, C-22: 33.5, C-23: 31.3, C-24: 39.3, C-25: 31.7, C-26: 20.4, C-27: 17.8, C-28: 15.5, CH_3CO : 21.2 and 170.1 ppm.

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